Amniotic fluid volume in intra-amniotic inflammation with and without culture-proven amniotic fluid infection in preterm premature rupture of membranes

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Abstract

Objective—Previous studies reported that the clinical significance of intra-amniotic inflammation with a negative amniotic fluid (AF) culture is similar to that of intra-amniotic inflammation with microbiologically-proven AF infection. However, the magnitude of the fetal inflammatory response in these two conditions is different as gauged by umbilical cord C-reactive protein (CRP) concentrations. We undertook this study to determine if the frequency of oligohydramnios is different in these two conditions.

Methods—The amniotic fluid index (AFI) was measured in 205 patients with preterm premature rupture of membranes (PROM) (≤35 weeks). AF was cultured for aerobic and anaerobic bacteria and genital mycoplasmas. Intra-amniotic inflammation was defined as an elevated AF matrix metalloproteinase-8 (MMP-8) concentration (>23 ng/mL). Patients were divided into three groups according to the results of AF culture and the presence or absence of intra-amniotic inflammation: 1) without intra-amniotic inflammation and a negative culture (N=109); 2) with intra-amniotic inflammation and a negative culture (N=44); and 3) a positive culture (N=52).

Results—Patients with a positive culture had a higher frequency of oligohydramnios and a lower median AFI than those with a negative culture but with intra-amniotic inflammation (p<0.01). However, there was no significant difference in the median AFI or in the frequency of oligohydramnios according to the presence or absence of intra-amniotic inflammation among patients with a negative culture (p>0.1).

Conclusion—Oligohydramnios was more frequent in patients with culture-proven AF infection than in those with intra-amniotic inflammation and a negative AF culture.

Keywords

Amniotic fluid infection (AFI); amniotic fluid infection; fetal inflammatory response syndrome (FIRS); intra-amniotic inflammation; oligohydramnios, rupture of membranes
Introduction

Microbial invasion of the amniotic cavity is present in one third of patients with preterm premature rupture of membranes (PROM) and is strongly associated with impending preterm delivery, adverse pregnancy and neonatal outcome [4,24,25]. Previous studies have reported that the clinical significance of intra-amniotic inflammation with a negative culture for microorganisms is similar to that of microbiologically-proven amniotic fluid (AF) infection [25,33]. However, proven AF infection, which is severe enough to yield a positive culture, may reflect a higher microbial burden and elicit a more intense fetal inflammatory response than intra-amniotic inflammation with a negative culture. A previous study documented that fetuses born to mothers with culture-proven AF infection had a higher umbilical cord plasma C-reactive protein (CRP) than those born to mothers with culture-negative intra-amniotic inflammation [10]. This could have different implications for fetal target organs, including the kidney.

Oligohydramnios is frequently present in preterm PROM [18,27,29]. Though the reduced AF volume is partly attributed to the escape of fluid through the site of membrane rupture, microbial invasion of the amniotic cavity and the subsequent fetal inflammatory response syndrome (FIRS) is known to be associated with oligohydramnios, which has been attributed to the decreased fetal urine production [18,30]. The purpose of this study was to determine if proven AF infection is associated with changes in AF volume.

Materials and Methods

Study design

The relationship between AF volume and intra-amniotic inflammation and/or AF infection was examined in 205 singleton pregnancies admitted to our university hospital with the diagnosis of preterm PROM who met the following criteria: (1) preterm pregnancy [gestational age (GA) ≤35 weeks]; (2) AF obtained for microbiologic studies by transabdominal amniocentesis or at the time of cesarean delivery; and (3) AF index (AFI) determined before AF retrieval according to the method described by Phelan et al. [19]. Oligohydramnios was defined as AFI≤5cm.

Patients were divided into three groups according to the presence or absence of intra-amniotic inflammation and AF culture results: Group 1: patients without inflammation, without infection (a negative AF culture; N=109); Group 2: patients with intra-amniotic inflammation, without infection (a negative AF culture; N=44); and Group 3: patients with a positive AF culture (N=52). Retrieval of AF was performed after written informed consent was obtained. The Institutional Review Board of the participating institution approved the collection and use of these samples and information for research purposes.

AF studies

AF was cultured for aerobic and anaerobic bacteria, as well as genital mycoplasmas (Mycoplasma hominis and Ureaplasma urealyticum). An aliquot of AF was transported to the laboratory and examined in a hemocytometer chamber to determine the white blood cell (WBC) count. The remaining fluid was centrifuged and stored in polypropylene tubes at −70 °C. Matrix metalloproteinase-8 (MMP-8) concentration was measured with a commercially available enzyme-linked immunosorbent assay (Amersham Pharmacia Biotech, Inc, Bucks, UK). The sensitivity of the test was 0.3 ng/mL. Intra- and inter-assay coefficients of variation were <10%, respectively. MMP-8 was used to assess the presence of intra-amniotic inflammation because previous studies indicated that it is a sensitive and specific index of inflammation [1,12,17]. Intra-amniotic inflammation was defined as an elevated AF MMP-8 concentration (>23 ng/mL), as previously reported [17].
Diagnosis of chorioamnionitis

Clinical chorioamnionitis was diagnosed in the presence of maternal temperature of $\geq 37.8 \, ^\circ\text{C}$ and two or more of the following criteria: (1) uterine tenderness; (2) malodorous vaginal discharge; (3) maternal leukocytosis (WBC count of $>15,000 \, \text{cells/mm}^3$); (4) maternal tachycardia ($>100 \, \text{beats/min}$); and (5) fetal tachycardia ($>160 \, \text{beats/min}$) [3]. Acute histologic chorioamnionitis was diagnosed if acute inflammatory changes were present on examination of the extra-placental membranes or the chorionic plate of the placenta, according to the criteria previously published [31]. Funisitis was diagnosed in the presence of neutrophil infiltration into the umbilical vessel walls or into Wharton’s jelly.

Statistical analysis

Proportions were compared with the Fisher’s exact test. A Kruskal-Wallis analysis of variance test was used for comparison of continuous variables among groups. Multiple comparisons between groups were performed with the Mann-Whitney U test. The amniocentesis-to-delivery interval was compared using the generalized Wilcoxon test for survival analysis. The interval-to-delivery of patients induced for maternal or fetal indications was treated as a censored observation, with a censoring time equal to the amniocentesis-to-delivery interval. A $P<0.05$ was considered significant.

Results

The frequency of oligohydramnios (AFI $\leq 5 \, \text{cm}$) was 29% (59/205). The prevalence of positive AF culture was 25% (52/205). Microorganisms isolated from the AF included *Ureaplasma urealyticum* (n=37), Candida species (n=4), Streptococcus species (n=4), *Escherichia coli* (n=3), coagulase-negative Staphylococcus (n=3), *Mycoplasma hominis* (n=3), and one isolate each of Peptostreptococcus species, Corynebacterium species, *Acinetobacter baumanii*, *Burkholderia cepalia*, *Staphylococcus hominis*, and *Staphylococcus epidermidis*. Six patients had polymicrobial infections (4 patients with 2 species, and 2 patients with 3 species of microorganisms).

Table 1 describes the clinical characteristics and pregnancy outcomes of the study population according to the results of AF culture and MMP-8 concentrations. Patients with intra-amniotic inflammation but a negative AF culture (Group 2) had a significantly lower median GA at delivery, and birth weight, and a higher rate of histologic chorioamnionitis and funisitis than those without intra-amniotic inflammation and a negative AF culture (Group 1). However, there was no significant difference in the frequency of oligohydramnios between groups 1 and 2 (26% vs. 16%; $p=0.2$).

There were no significant differences in the clinical characteristics and pregnancy outcomes between patients with intra-amniotic inflammation and a negative AF culture (Group 2) and those with positive AF culture (Group 3), except for the frequency of oligohydramnios, which was higher in group 3 than in group 2 (46% vs. 16%; $p<0.005$).

Figure 1 shows that patients with a positive AF culture had a lower median AFI than those with a negative culture but with intra-amniotic inflammation (median, 5.1 [range 0–17.5] vs. median 8.4 [range 0–21.9]; $p<0.01$). However, there was no significant difference in the median AFI according to the presence or absence of intra-amniotic inflammation in patients with a negative AF culture (median, 8.1 [range 0–31.0] vs. median 8.4 [range 0–21.9]; $p=0.8$).

Forty-six percent (24/52) of patients with a positive AF culture had oligohydramnios. Among patients with a positive AF culture, the median GA at amniocentesis and delivery, birth weight, and WBC count in AF, and the frequency of clinical chorioamnionitis (29% vs. 11%), histologic chorioamnionitis (100% vs. 78%), and funisitis (77% vs. 74%) were not significantly
different in patients with or without oligohydramnios. However, patients with oligohydramnios had a shorter interval-to-delivery than those without oligohydramnios (median, 38 hours [range 0–285 hours] vs. median 95 hours [range 7–748 hours]; p<0.001, Figure 2).

Cox proportional hazards model analysis indicated that the presence of oligohydramnios (AFI ≤5 cm) was an independent predictor of the interval-to-delivery after adjustment for GA and the AF culture results [hazards ration (HR), 1.8; 95%. Confidence interval (CI) (1.3–2.6)].

Comment

Principal findings of this study

1) Oligohydramnios is associated with microbiologically-proven AF infection; 2) among cases with proven AF infection, those with oligohydramnios had a shorter amniocentesis-to-delivery interval than those with an AFI >5 cm.

AF infection, FIRS and oligohydramnios

Oligohydramnios is often associated with microbial invasion of the amniotic cavity [18,27–30]. Microorganisms in the AF can gain access to the fetus and elicit a systemic FIRS [5,23,26]. Pro-inflammatory cytokines released during FIRS can have effects on multiple fetal organ systems such as the brain and heart, leading to blood flow redistribution and cardiac dysfunction [22,30,34]. Reduction of renal blood flow during FIRS can result in the decrease of fetal urine production and the subsequent oligohydramnios. Indeed, a previous study indicated that umbilical cord plasma concentrations of interleukin-6, which is the hallmark of FIRS, are significantly elevated in fetuses with oligohydramnios and preterm PROM compared with those without oligohydramnios [30]. Therefore, there is solid evidence linking microbial invasion of the amniotic cavity, FIRS and oligohydramnios.

Intra-amniotic inflammation in the absence of proven AF infection

A novel finding of this study is that among patients with intra-amniotic inflammation, those with proven AF infection had a decreased AF volume, whereas those with a negative AF culture did not. Substantial evidence indicates that intra-amniotic inflammation is a risk factor for adverse pregnancy and neonatal outcome, regardless of the presence or absence of a positive AF culture [2,7–9,16,25,33,34]. However, the data presented herein demonstrates that a culture-proven infection has clinical implications beyond the presence of intra-amniotic inflammation with a negative AF culture in preterm PROM. This finding suggests that FIRS, in cases with intra-amniotic inflammation with a negative culture, may be milder than the one associated with a positive AF culture. Milder cases of FIRS may not be severe enough to cause a reduction of fetal urine production, which appears to occur in cases with proven AF infection and more severe FIRS. Indeed, studies in adults indicate that there is disease progression which begins with a systemic inflammatory response and advanced to sepsis, severe sepsis and septic shock [11,20].

We propose that fetuses with a positive AF cultures have a more severe systemic inflammatory response compared to those with intra-amniotic inflammation with a negative culture. Support for this hypothesis is derived from a prior report indicating that among patients with intra-amniotic inflammation, those with proven infection had a higher concentration of umbilical cord plasma CRP than those with a negative AF culture [10].

Intensity of FIRS

This study suggests that fetuses with positive AF cultures have a greater degree of systemic fetal inflammation compared to those with intra-amniotic inflammation and negative AF cultures. Why the difference? Intra-amniotic inflammation in the absence of proven AF
infection is commonly considered to be attributed to infection in which organisms escaped detection by traditional microbiological methods [15,32]. It is possible that the microbial burden of patients with a positive culture is greater than that of those with a negative culture. Small inoculum size of microorganisms probably contributed to the failure of cultivation. It is also possible that microorganisms recovered with culture techniques may be more virulent than those who resist cultivation in the laboratory.

**FIRS in proven AF infection**

AF infection is associated with immune responses [6,13,21], and FIRS has been also understood as an immune response, which can signal the initiation of parturition for the fetus to exit a hostile intrauterine environment [23]. Of interest, our data showed that among cases with proven AF infection, patients with oligohydramnios had an increased risk for impending delivery compared to those with normal AF volume. This suggests that fetuses with oligohydramnios in the presence of proven AF infection probably have an advanced stage of FIRS. However, this cannot be addressed with current data because the difference in the magnitude of FIRS between cases with and without oligohydramnios was not examined. Such issue requires a large cohort consisting of fetuses whose umbilical cord blood was retrieved at the time of AFI determination.

**Unanswered questions, limitations and further considerations**

Further studies are required to determine if oligohydramnios in the presence of proven AF culture is associated with a worse neonatal outcome. This question cannot be addressed with the sample size in this study.

**Clinical implication in this study**

This investigation indicated that oligohydramnios is associated with microbiologically proven AF infection, and that among cases with proven AF infection, patients with oligohydramnios had an increased risk for impending delivery compared with those with a normal AFI. Indeed, several studies have reported a higher frequency of sepsis in neonates born to mothers with oligohydramnios [14,28–30]. Therefore, we propose that the presence of oligohydramnios combined with the results of AF culture should be considered when planning the management of patients presenting preterm PROM.

**Acknowledgments**

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**References**


Figure 1.
Amniotic fluid index (AFI) according to the presence or absence of intra-amniotic inflammation and AF culture results: the median AFI in patients with intra-amniotic inflammation and a negative AF culture was significantly higher than that in patients with a positive AF culture, but was not different from that in patients without intra-amniotic inflammation and with a negative AF culture.
Figure 2.
Survival analysis of interval-to-delivery among patients with preterm premature rupture of membranes and amniotic fluid infection according to the presence or absence of oligohydramnios (median, 38 hours; range, 0–285 hours vs. median, 95 hours; range, 7–748 hours; P<0.001)
Table 1

Clinical characteristics and pregnancy outcomes of the study population according to the results of AF culture and MMP-8 concentrations

<table>
<thead>
<tr>
<th></th>
<th>(−) AF culture</th>
<th>P&lt;sup&gt;a&lt;/sup&gt;</th>
<th>(−) AF culture</th>
<th>P&lt;sup&gt;b&lt;/sup&gt;</th>
<th>(+) AF culture</th>
<th>P&lt;sup&gt;c&lt;/sup&gt;</th>
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<tbody>
<tr>
<td>Maternal age (years)</td>
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<tr>
<td>(group 1; n=109)</td>
<td>30 (20–41)</td>
<td>NS</td>
<td>31 (23–43)</td>
<td>NS</td>
<td>29 (21–43)</td>
<td>NS</td>
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<tr>
<td>Nulliparity (n)</td>
<td>57 (52%)</td>
<td>NS</td>
<td>19 (43%)</td>
<td>NS</td>
<td>22 (42%)</td>
<td>NS</td>
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<td>GA at amniocentesis (week)&lt;sup&gt;d&lt;/sup&gt;</td>
<td>33.1 (20.4–35.0)</td>
<td>&lt;.001</td>
<td>30.6 (21.7–34.9)</td>
<td>NS</td>
<td>30.8 (22.4–34.7)</td>
<td>&lt;.001</td>
</tr>
<tr>
<td>GA at delivery (week)&lt;sup&gt;d&lt;/sup&gt;</td>
<td>34.7 (25.0–41.6)</td>
<td>&lt;.001</td>
<td>31.4 (22.3–35.0)</td>
<td>NS</td>
<td>31.5 (23.1–34.9)</td>
<td>&lt;.001</td>
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<tr>
<td>Birth weight (g)&lt;sup&gt;d&lt;/sup&gt;</td>
<td>2330 (660–4390)</td>
<td>&lt;.001</td>
<td>1680 (520–2630)</td>
<td>NS</td>
<td>1625 (570–2410)</td>
<td>&lt;.001</td>
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<tr>
<td>Amniocentesis-to-delivery interval (h)&lt;sup&gt;d&lt;/sup&gt;</td>
<td>127.8 (2.2–3544.5)</td>
<td>&lt;.001</td>
<td>47.0 (0–881.5)</td>
<td>NS</td>
<td>84.9 (0.2–748.4)</td>
<td>&lt;.001</td>
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<td>Preterm delivery (n)</td>
<td></td>
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<td>Within 36 weeks</td>
<td>90 (83%)</td>
<td>&lt;.005</td>
<td>44 (100%)</td>
<td>(−)</td>
<td>52 (100%)</td>
<td>&lt;.001</td>
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<tr>
<td>Within 34 weeks</td>
<td>32 (29%)</td>
<td>&lt;.001</td>
<td>36 (82%)</td>
<td>NS</td>
<td>44 (85%)</td>
<td>&lt;.001</td>
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<tr>
<td>Clinical chorioamnionitis</td>
<td>2 (2%)</td>
<td>NS</td>
<td>3 (7%)</td>
<td>NS</td>
<td>10 (19%)</td>
<td>&lt;.001</td>
</tr>
<tr>
<td>Histologic chorioamnionitis (n/N)</td>
<td>41/95 (43%)</td>
<td>&lt;.001</td>
<td>30/34 (88%)</td>
<td>NS</td>
<td>38/44 (86%)</td>
<td>&lt;.001</td>
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<tr>
<td>Histologic chorioamnionitis; delivered ≤48 h&lt;sup&gt;e&lt;/sup&gt;</td>
<td>5/16 (31%)</td>
<td>&lt;.005</td>
<td>14/15 (93%)</td>
<td>NS</td>
<td>16/18 (89%)</td>
<td>&lt;.005</td>
</tr>
<tr>
<td>Funisitis (n/N)</td>
<td>18/95 (19%)</td>
<td>&lt;.001</td>
<td>21/34 (62%)</td>
<td>NS</td>
<td>33/44 (75%)</td>
<td>&lt;.001</td>
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<td>Funisitis; delivered ≤48 h&lt;sup&gt;e&lt;/sup&gt;</td>
<td>1/16 (6%)</td>
<td>&lt;.005</td>
<td>9/15 (60%)</td>
<td>NS</td>
<td>14/18 (78%)</td>
<td>&lt;.001</td>
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<tr>
<td>AF WBC (cells/mm&lt;sup&gt;3&lt;/sup&gt;)&lt;sup&gt;d&lt;/sup&gt;</td>
<td>1 (0–243)</td>
<td>&lt;.001</td>
<td>414 (0–1000)</td>
<td>NS</td>
<td>314 (0–1000)</td>
<td>&lt;.001</td>
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<td>AF MMP-8 (ng/mL)&lt;sup&gt;d&lt;/sup&gt;</td>
<td>1.4 (0.3–22.3)</td>
<td>&lt;.001</td>
<td>184.4 (24.9–2874.5)</td>
<td>NS</td>
<td>255.6 (0.4–2370.6)</td>
<td>&lt;.001</td>
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<tr>
<td>Oligohydramnios (AFI ≤5)</td>
<td>28 (26%)</td>
<td>NS</td>
<td>7 (16%)</td>
<td>&lt;.005</td>
<td>24 (46%)</td>
<td>&lt;.05</td>
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</table>

Values are medians and ranges

AF, amniotic fluid; MMP-8, matrix metalloproteinase-8; Low MMP-8, matrix metalloproteinase-8 <23 ng/mL; High MMP-8, matrix metalloproteinase-8 >23 ng/mL; GA, gestational age; NS, not significant; WBC, white blood cell

<sup>a</sup> Comparison between groups 1 and 2.

<sup>b</sup> Comparison between groups 2 and 3.

<sup>c</sup> Comparison between groups 3 and 1.
# $P<.05$, by Kruskal-Wallis ANOVA test

§ To preserve a meaningful temporal relationship between the results of AF studies and histologic findings of the placenta obtained at delivery, cases delivered within 48 hours of amniocentesis were considered for the analysis.