

Development of a ^{177}Lu -Labeled RGD Derivative for Targeting Angiogenesis

Chang Hwan Ju,^{1,2} Jae Min Jeong,¹ Yun-Sang Lee,¹ Young Joo Kim,¹ Byung Chul Lee,¹
Dong Soo Lee,¹ June-Key Chung,¹ Myung Chul Lee,¹ and Seo Young Jeong²

Abstract

Various Arg-Gly-Asp (RGD) derivatives have been labeled with various radioisotopes for targeting $\alpha_v\beta_3$ integrin, which is expressed during angiogenesis in tumor. In this study, 2-(4'-isothiocyanatobenzyl)-1,4,7-triazacyclononane-1,4,7-triacetic acid (NOTA-SCN) and its c(RGDyK) conjugate (NOTA-SCN-c(RGDyK)) were labeled with ^{177}Lu , which is a near ideal radionuclide for treating tumors because it emits therapeutic beta particles and gamma rays for monitoring. ^{177}Lu (250 MBq) was labeled with 50 μg NOTA-SCN-c(RGDyK) quantitatively. The specific activity of ^{177}Lu -NOTA-SCN-c(RGDyK) was 1.44×10^5 Ci/mol. Biodistribution study was performed in Balb/c mice xenografted with CT-26 (mouse colon cancer) cells. The highest uptake was found in kidneys ($7.56\% \pm 0.71\%$ ID/g at 1 hour), and tumor uptake was $1.70\% \pm 0.33\%$ ID/g at 1 hour postinjection. Moderate tumor-to-blood (2.36 ± 0.29) and tumor-to-muscle (2.06 ± 0.40) ratios were observed. This study shows that ^{177}Lu -NOTA-SCN-c(RGDyK) is a potential therapeutic agent for angiogenic tumors, but special care is required to prevent kidney toxicity.

Key words: angiogenesis, Lu-177, lutetium, NOTA, radionuclide therapy, RGD

Introduction

Angiogenesis is required for tumor proliferation and metastasis,¹ and the integrins, which are composed of α and β subunits, participate in this process via cell-cell and cell-matrix interactions.² In particular, $\alpha_v\beta_3$ integrin is found where cell-cell and cell-matrix interactions occur and is highly expressed on osteoclasts, inflammatory cells, angiogenic cells, and tumor cells. Further, peptides with the arginine-glycine-aspartic acid (RGD) amino acid sequence strongly bind to $\alpha_v\beta_3$ integrin,³ and many studies, in which $\alpha_v\beta_3$ integrin was targeted using RGD derivatives, have reported that these derivatives inhibit angiogenesis.⁴ It has been also reported that anti- $\alpha_v\beta_3$ integrin monoclonal antibodies inhibit angiogenesis without affecting pre-existing blood vessels.⁵⁻⁷ Further, $\alpha_v\beta_3$ integrin is viewed as a promising candidate for tumor imaging and therapy, because its expression is highly specific. Accordingly, many radiolabeled RGD derivatives have been devised to target $\alpha_v\beta_3$ integrin, for

example, the interaction between ^{125}I -labeled c(RGDyK) and $\alpha_v\beta_3$ integrin has been used to target angiogenesis.⁸ However, the high gut activity of ^{125}I -labeled c(RGDyK) caused by hepatobiliary excretion is problematic. Several other radiolabeled RGD derivatives have been reported to improve pharmacokinetics and tumor affinities.⁹ Dimerized and multimerized derivatives have been developed to improve tumor accumulation and pharmacokinetics. Although these derivatives showed increased affinity for $\alpha_v\beta_3$ integrin, uptakes in nontargeted organs such as kidney and liver also increased.¹⁰⁻¹³

Beta emitters, especially ^{131}I , are used therapeutically,^{14,15} and ^{90}Y - and ^{111}In -labeled RGD peptides have shown potential for therapy and imaging, respectively.¹⁶ ^{177}Lu is known to be a promising radionuclide for therapeutic applications because it has suitable nuclear physical properties ($T_{1/2} = 6.73$ days, $E_{\beta(\text{max})} = 0.49$ MeV, $E_{\gamma} = 208$ keV [11%])¹⁷ and is amenable to large-scale production. In particular, it has the advantages of a wide distribution without significant decay loss

¹Department of Nuclear Medicine, Cancer Research Institute, Seoul National University College of Medicine, Seoul, Korea.

²Department of Life and Nanopharmaceutical Sciences, College of Pharmacy, Kyung Hee University, Seoul, Korea.

Address correspondence to: Jae Min Jeong; Department of Nuclear Medicine, Seoul National University College of Medicine; 101 Daehangno Jongno-gu, Seoul 110-774, Korea
E-mail: jmjng@snu.ac.kr

due to a relatively long half-life, has adequate specific activity, and can be generated at excellent radionuclidic purities. Further, ^{177}Lu -labeled EDTMP and DOTMP showed potential for the palliation of bone metastases.¹⁸

In the present study, a c(RGDyK) derivative conjugated with a bifunctional chelating agent isothiocyanatobenzyl-1,4,7-triazacyclononane-1,4,7-triacetic acid (SCN-Bn-NOTA) was labeled with ^{177}Lu (Fig. 1).¹⁹ NOTA is a nine-membered cyclic compound and forms stable neutral complexes with metallic radionuclides, such as gallium and indium. In particular, gallium-NOTA complex is highly stable ($k = 10^{31}$) because the small ionic radius of Ga(III) (62 pm) matches that of the NOTA chelate cage. Further, Ga(III) has been used for radiolabeling various biomolecules.^{20,21} However, NOTA has not been previously labeled with ^{177}Lu because lutetium has relatively large ionic radius.

In the present study, NOTA-c(RGDyK) was labeled with ^{177}Lu and its biodistribution was investigated in tumor-xenografted mice.

Materials and Methods

Synthesis of NOTA-RGD

4'-SCN-Bn-NOTA and c(RGDyK) were purchased from Futurechem. All other reagents and solvents were purchased from Sigma-Aldrich.

NOTA-RGD was synthesized as previously described.¹⁹ In brief, 4'-SCN-Bn-NOTA (660 nmol, 0.3 mg) was added to c(RGDyK) (600 nmol, 0.37 mg) in 0.1 M sodium carbonate buffer (pH 9.5) and allowed to react for 20 hours at room temperature in the dark. NOTA-RGD peptides were then purified by high-performance liquid chromatography (HPLC; Agilent 1100 series) using an XTerra preparative column RP18 (10×250 mm; Waters) with 0%–100% ethanol gradient in 0.01% trifluoroacetic acid from 0 to 30 minutes at a flow rate of 3 mL/minute (retention time of NOTA-RGD was 13.2 minutes).

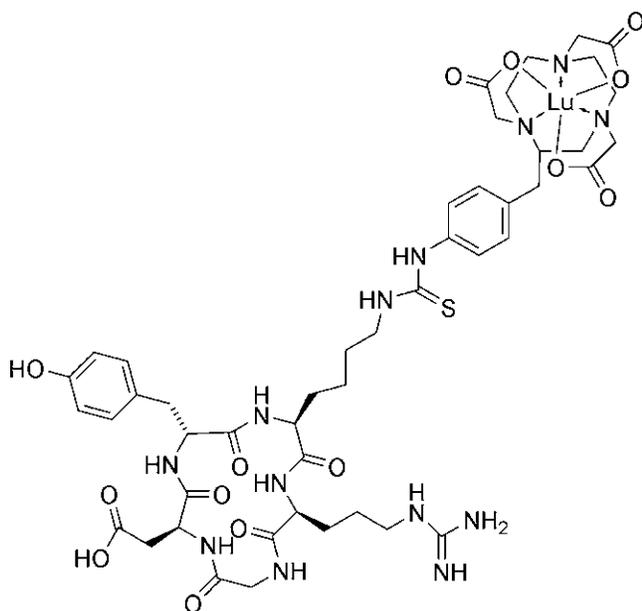


FIG. 1. Chemical structure of ^{177}Lu -NOTA-RGD. RGD, arginine-glycine-aspartic acid.

Labeling with ^{177}Lu

^{177}Lu (250 MBq; Perkin-Elmer) was added to a solution of 50 μg NOTA-SCN-c(RGDyK) and 1.0 mg selenomethionine in 300 μL sodium acetate buffer (pH 5.5, 0.4 M) and heated at 90°C for 35 minutes. Labeling efficiencies were checked by instant thin layer chromatography-silica gel (ITLC-SG; Pall Co.) using 0.1 M citric acid as eluant (R_f values of ^{177}Lu -NOTA-RGD and free ^{177}Lu were 1.0 and 0.0, respectively) or by Whatman No. 1 paper (Kent) using 50% ethanol as eluant (R_f values of ^{177}Lu -NOTA-RGD and free ^{177}Lu were 0.0 and 1.0, respectively). And labeling efficiencies were confirmed by HPLC (XTerra preparative column RP18; 250 nm; 100%–30% of 10 mM HCl gradient, 0%–70% MeCN gradient for 20 minutes). The retention time of ^{177}Lu -NOTA-RGD in this system was 18.5 minutes. The radioactivity and the optical density at 240 nm of the peak fraction containing ^{177}Lu -NOTA-RGD were measured, and specific activity was calculated.

Stability of ^{177}Lu -NOTA-RGD

The stability of ^{177}Lu -NOTA-RGD was checked at room temperature over 1 week. Radiochemical purity for stability testing was checked by ITLC-SG and paper chromatography as described above.

Biodistribution study in mice with a colon cancer xenograft

The biodistribution study was performed at Seoul National University Hospital, which has full Association for the Assessment and Accreditation of Laboratory Animal Care International (2007) accreditation.

The mouse colon cancer cell line CT-26 was grown in DMEM medium containing 10% fetal bovine serum and harvested with trypsin. Cells were washed with 10 mL of phosphate-buffered saline. Each male Balb/C mouse was injected subcutaneously (s.c.) with 2×10^5 CT-26 cells in right shoulder. Fourteen (14) days postinjection, ^{177}Lu -NOTA-RGD (0.37 MBq/0.1 mL) was injected intravenously via a tail vein. Mice were sacrificed by cervical dislocation at 10 minutes, 1 hour, 4 hours, 12 hours, 24 hours, and 48 hours postinjection; thigh muscles, blood, and other organs were excised immediately, weighed, and counted using a Cobra II γ -scintillation counter (Packard Canberra Co.). Results were expressed as percentages of injected dose per gram of tissue (% ID/g).

Results

Radiolabeling

The ^{177}Lu -NOTA-RGD produced was analyzed by ITLC-SG, paper chromatography, and HPLC. For these analyses, ITLC-SG plates and Whatman No. 1 paper were eluted with 0.1 M citric acid and 50% ethanol, respectively. When ITLC-SG plates were eluted with 0.1 M citric acid, free ^{177}Lu moved with the solvent front and ^{177}Lu -NOTA-RGD remained at the origin, but when Whatman No. 1 paper plates were eluted with 50% ethanol, free ^{177}Lu remained at the origin and ^{177}Lu -NOTA-RGD moved with the solvent front. The labeling efficiency was over 99% and almost no free ^{177}Lu was found.

App: Y-90-RGD-SCN-NOTA, Run: a_7_y-90-nota-scn-RGD_100ug_SuL_080312 @ 3/12/2008 8:03:21 PM, Method: Y-90-RGD-SCN-NOTA, Inj: 1

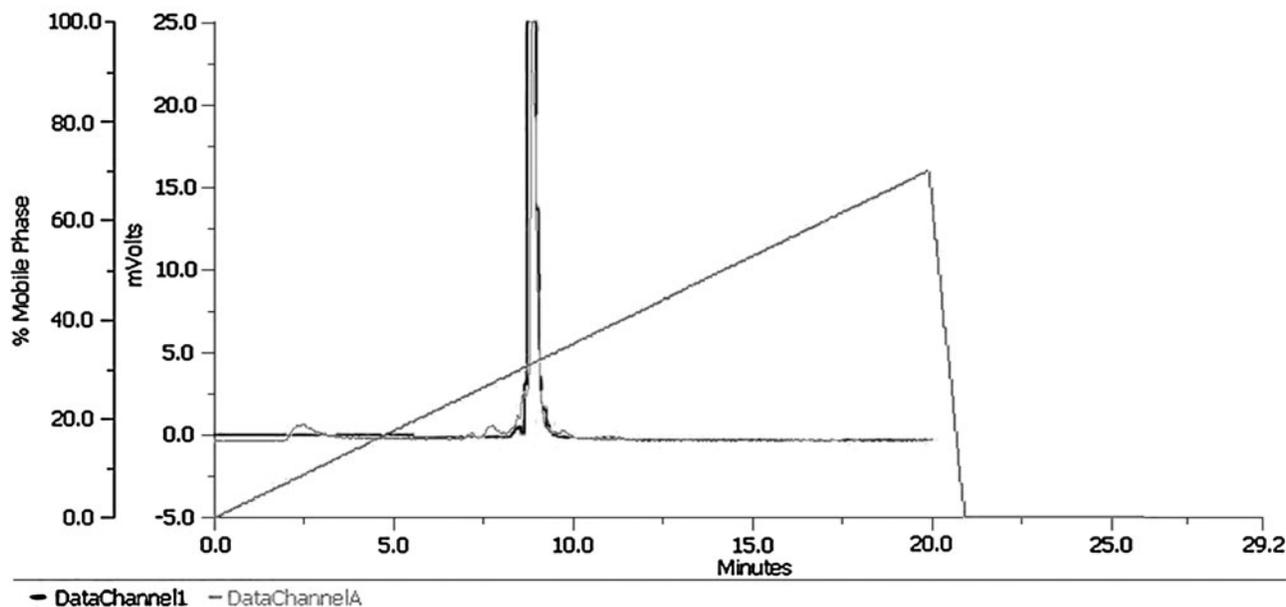


FIG. 2. High-performance liquid chromatography profiles of ¹⁷⁷Lu-NOTA-RGD. Zero percent to 70% MeCN gradient from 0 to 20 minutes in 10 nM HCl. High purity of radiolabeled product was demonstrated.

¹⁷⁷Lu-NOTA-RGD had a longer retention time (18.5 minutes) than unlabeled NOTA-RGD by preparative HPLC (Fig. 2). The specific activity of ¹⁷⁷Lu-NOTA-RGD was 5.33 GBq/μmol, and labeled ¹⁷⁷Lu-NOTA-RGD was found to be stable for more than 24 hours at room temperature by HPLC (Fig. 3).

Biodistribution in tumor xenograft model

The biodistribution of ¹⁷⁷Lu-NOTA-RGD was investigated in a mouse colon cancer CT-26-bearing Balb/C mice model at 14th day post-transplantation (Fig. 4). Levels of ¹⁷⁷Lu-NOTA-RGD uptake in the kidneys (7.56% ± 0.71%

ID/g at 1 hour) were high, suggesting that most was excreted via the kidneys. Although tumor uptake was not high (1.70% ± 0.33% ID/g at 1 hour), tumor-to-blood (2.36 ± 0.29 at 1 hour) and tumor-to-muscle ratios (2.06 ± 0.40 at 1 hour) were high and increased with time (Table 1). Bone uptakes were similar to those of other organs, such as liver, spleen, stomach, and intestine, and decreased with time, which indicates that ¹⁷⁷Lu-NOTA-RGD is sufficiently stable *in vivo*.

Discussion

In this study, a conjugate of NOTA and RGD derivatives was designed for diagnostic and therapeutic applications and investigated as a potential ligand for ¹⁷⁷Lu labeling. This type of derivative was chosen because RGD peptides specifically target the α_vβ₃ integrin expressed in tumors. In terms of preparation, c(RGDyK) and SCN-Bn-NOTA were conjugated by thiourea bond formation. This process leaves all carboxyl residues of NOTA intact and available for complex formation with ¹⁷⁷Lu.

The NOTA-c(RGDyK) conjugate was purified by HPLC and then labeled with ¹⁷⁷Lu at 90°C for 35 minutes. Free ¹⁷⁷Lu remained at the origin by paper chromatography when eluted with 50% ethanol and moved with the solvent front when eluted with a 0.1 M citric acid solution on ITLC-SG.

Like ¹³¹I, ¹⁸⁸Re, ¹⁶⁶Ho, and ⁹⁰Y, ¹⁷⁷Lu is a promising therapeutic β-emitter and has adequate nuclear physical properties (T_{1/2} = 6.73 days, E_{β(max)} = 0.49 MeV, E_γ = 208 keV [11%]) for therapeutic applications. ¹⁷⁷Lu could be a viable alternative for ¹³¹I (T_{1/2} = 8.02 days, E_{β(max)} = 0.606 MeV, E_γ = 364 keV [81%]) for therapeutic applications and has an advantage of a relatively low energy (208 keV) and low abundance (11%) of major γ emission. In addition, ¹⁷⁷Lu can be produced in large scale with good radionuclidic purity and adequate specific activity.

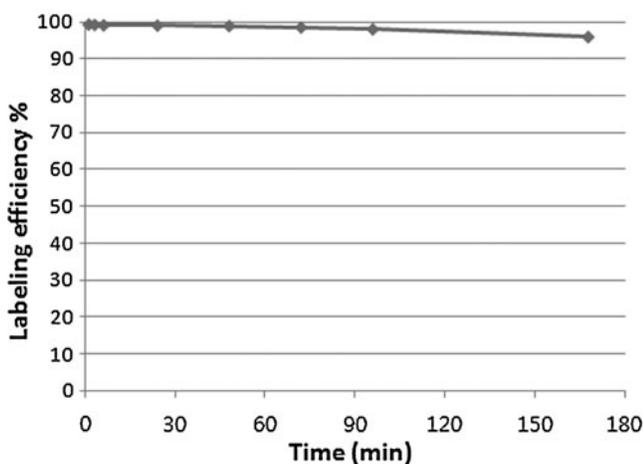


FIG. 3. Stability of ¹⁷⁷Lu-NOTA-RGD at room temperature for 7 days. Stabilities were checked by instant thin layer chromatography-silica gel and Whatman paper, using 0.1 M citric acid and 50% ethanol as eluants, respectively.

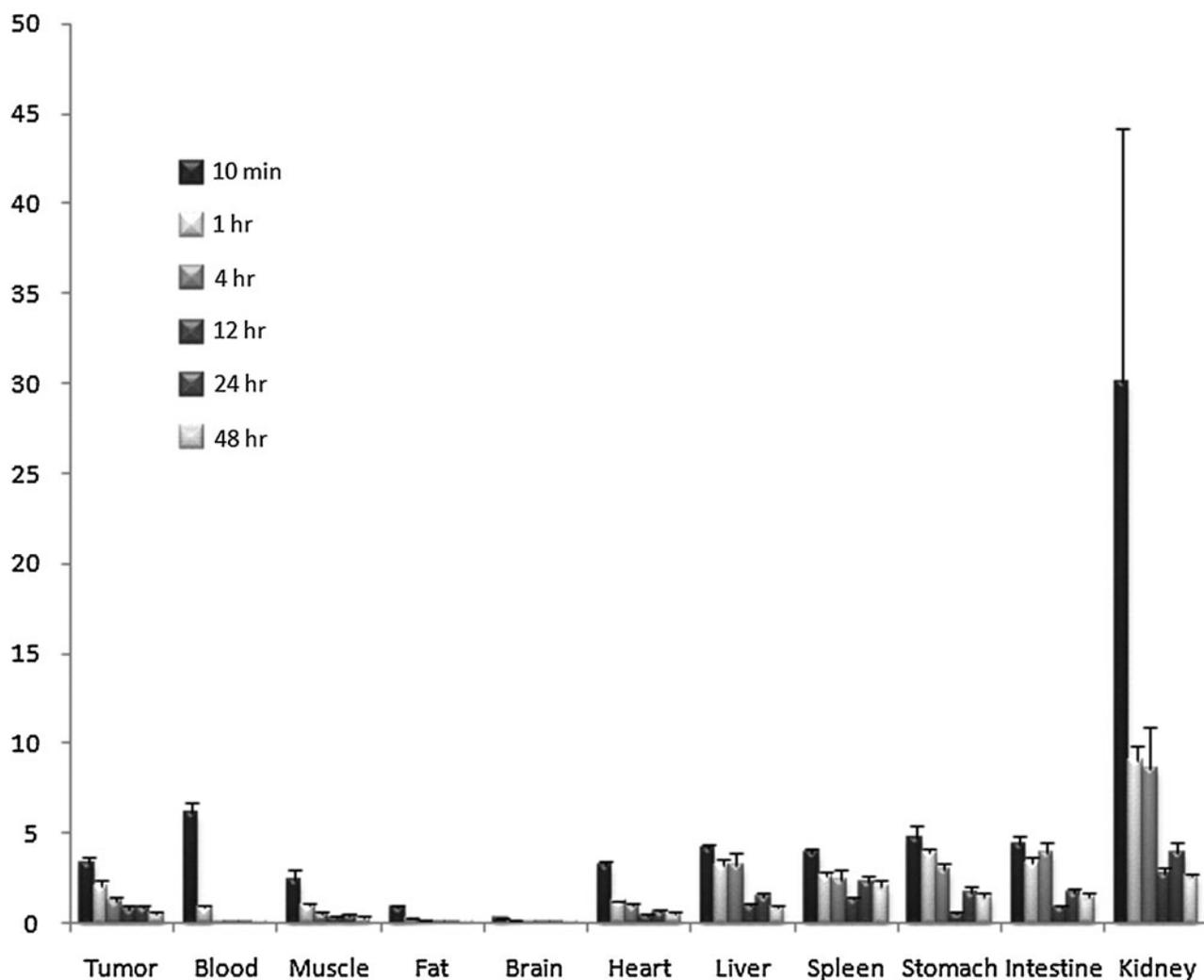


FIG. 4. Biodistribution of ^{177}Lu -NOTA-RGD in Balb/c mice with a CT-26 tumor after intravenous injection through tail vein.

^{188}Re ($T_{1/2} = 17$ hours, $E_{\beta(\text{max})} = 2.21$ MeV, $E_{\gamma} = 155$ keV [15%]) and ^{90}Y ($T_{1/2} = 64$ hours, $E_{\beta(\text{max})} = 2.18$ MeV) have been reported to be attractive radionuclides for cancer therapy, because they have favorable nuclear decay characteristics and can be produced from generators installed in hospitals. However, the availability of $^{188}\text{W}/^{188}\text{Re}$ generator is limited, because it is produced by a double neutron cap-

ture reaction, and only few reactors worldwide can provide the high thermal neutron fluxes ($>5 \times 10^{14}$ n/cm²/s) required to produce ^{188}W in the quantities required for the preparation of $^{188}\text{W}/^{188}\text{Re}$ generator.^{22–24} ^{90}Y is also obtained from a generator, but radionuclidic contamination by ^{90}Sr ($T_{1/2} = 28.3$ years) is problematic, and thus, the generator is not available commercially.²⁵

TABLE 1. TUMOR-TO-BLOOD AND TUMOR-TO-MUSCLE RATIOS OF ^{177}Lu -NOTA-RGD UPTAKES IN BALB/C MICE WITH CT-26 TUMOR AFTER INTRAVENOUS INJECTION THROUGH TAIL VEIN

Time	Tumor/blood	Tumor/muscle
10 minutes	0.56 ± 0.08	1.42 ± 0.31
1 hour	2.36 ± 0.29	2.06 ± 0.40
4 hours	15.09 ± 3.66	2.18 ± 0.44
12 hours	33.07 ± 8.36	2.50 ± 0.62
24 hours	33.77 ± 10.01	1.98 ± 0.41
48 hours	31.37 ± 7.10	1.68 ± 0.24

DOTA has been used as a bifunctional chelating agent for labeling peptides with lanthanide radionuclides, such as ^{177}Lu , because of its ability to form highly stable chelates.²⁶ However, in the present study, it was found that NOTA also forms a stable chelate with ^{177}Lu . Although NOTA has been previously reported to form a stable chelate with ^{68}Ga ,²⁷ this is the first report of the successful labeling of an RGD derivative with ^{177}Lu .

The biodistribution model used in this study was a CT-26 (mouse colon carcinoma)-xenografted mouse model, and it was found that ^{177}Lu -NOTA-RGD was well taken up by tumors. Hydrophilicity and hydrophobicity are important determinants of renal excretion and hepatobiliary excretion. In the present study, high renal excretion and low hepatobiliary excretion were attributed to the hydrophilic nature of

¹⁷⁷Lu-NOTA residues. ¹⁷⁷Lu-NOTA-RGD shows high kidney uptake, and thus, it is probably cleared rapidly through kidneys. However, kidney uptake might be the dose-limiting factor for radionuclide therapy using ¹⁷⁷Lu-NOTA-RGD, and thus, ¹⁷⁷Lu-NOTA-RGD doses should be considered carefully to prevent kidney toxicity.

Conclusions

A NOTA-RGD conjugate was synthesized, labeled with ¹⁷⁷Lu, and its *in vitro* stability and *in vivo* biodistribution were investigated in tumor-bearing mice. ¹⁷⁷Lu has a considerable potential as a therapeutic radionuclide because of its suitable decay properties. ¹⁷⁷Lu-NOTA-RGD complex was prepared with excellent radiochemical purity and was found to have excellent *in vitro* stability. This biodistribution study in Balb/C mice showed high uptakes in kidneys and high tumor/blood ratios. These findings indicate that ¹⁷⁷Lu-NOTA-RGD might be a potential therapeutic agent for angiogenic tumors. However, caution must be exercised to prevent kidney toxicity.

Acknowledgments

This work was supported by an NRF of Korea grant (R0A-2008-000-20116-0) and the Converging Research Program (2010K001055) funded by the Ministry of Education, Science and Technology.

Disclosure Statement

No competing financial interests exist.

References

1. Carmeliet P, Jain RK. Angiogenesis in cancer and other diseases. *Nature* 2000;407:249.
2. Ruoslahti E, Pierschbacher MD. New perspectives in cell adhesion: RGD and integrins. *Science* 1987;238:491.
3. Plow EF, Haas TA, Zhang L, et al. Ligand binding to integrins. *J Biol Chem* 2000;275:21785.
4. Meyer A, Auernheimer J, Modlinger A, et al. Targeting RGD recognizing integrins: Drug development, biomaterial research, tumor imaging and targeting. *Curr Pharm Des* 2006;12:2723.
5. Brooks PC, Clark RA, Cheresch DA. Requirement of vascular integrin alpha v beta 3 for angiogenesis. *Science* 1994; 264:569.
6. Allman R, Cowburn P, Mason M. *In vitro* and *in vivo* effects of a cyclic peptide with affinity for the alpha(nu)beta3 integrin in human melanoma cells. *Eur J Cancer* 2000;36:410.
7. Brooks PC, Stromblad S, Klemke R, et al. Antiintegrin alpha v beta 3 blocks human breast cancer growth and angiogenesis in human skin. *J Clin Invest* 1995;96:1815.
8. Haubner R, Wester HJ, Reuning U, et al. Radiolabeled alpha(v)beta3 integrin antagonists: A new class of tracers for tumor targeting. *J Nucl Med* 1999;40:1061.
9. Haubner R. Alphavbeta3-integrin imaging: A new approach to characterise angiogenesis? *Eur J Nucl Med Mol Imaging* 2006;33(Suppl1):54.
10. Janssen M, Oyen WJ, Massuger LF, et al. Comparison of a monomeric and dimeric radiolabeled RGD-peptide for tumor targeting. *Cancer Biother Radiopharm* 2002;17:641.

11. Jia B, Shi J, Yang Z, et al. ^{99m}Tc-labeled cyclic RGDfK dimer: Initial evaluation for SPECT imaging of glioma integrin alphavbeta3 expression. *Bioconjug Chem* 2006;17:1069.
12. Janssen ML, Oyen WJ, Dijkgraaf I, et al. Tumor targeting with radiolabeled alpha(v)beta(3) integrin binding peptides in a nude mouse model. *Cancer Res* 2002;62:6146.
13. Zhang X, Xiong Z, Wu Y, et al. Quantitative PET imaging of tumor integrin alphavbeta3 expression with ¹⁸F-FRGD₂. *J Nucl Med* 2006;47:113.
14. Moon EH, Lim ST, Jeong YJ, et al. Efficacy of I-123/I-131 metaiodobenzylguanidine scan as a single initial diagnostic modality in pheochromocytoma: Comparison with biochemical test and anatomic imaging. *Nucl Med Mol Imaging* 2009;43:436.
15. Kim KH, Kim SM, Seo YD. A discrepancy between ¹³¹I-metaiodobenzylguanidine (¹³¹I-MIBG) scintigraphy and ¹⁸F-FDG PET/CT after ¹³¹I-MIBG therapy in a patient with recurrent malignant pheochromocytoma. *Nucl Med Mol Imaging* 2009;43:582.
16. Yoshimoto M, Ogawa K, Washiyama K, et al. Alpha(v) beta(3) integrin-targeting radionuclide therapy and imaging with monomeric RGD peptide. *Int J Cancer* 2008;123:709.
17. Chakraborty S, Das T, Sarma HD, et al. Preparation and preliminary studies on ¹⁷⁷Lu-labeled hydroxyapatite particles for possible use in the therapy of liver cancer. *Nucl Med Biol* 2008;35:589.
18. Chakraborty S, Das T, Sarma HD, et al. Comparative studies of ¹⁷⁷Lu-EDTMP and ¹⁷⁷Lu-DOTMP as potential agents for palliative radiotherapy of bone metastasis. *Appl Radiat Isot* 2008;66:1196.
19. Jeong JM, Hong MK, Chang YS, et al. Preparation of a promising angiogenesis PET imaging agent: ⁶⁸Ga-labeled c(RGDyK)-isothiocyanatobenzyl-1,4,7-triazacyclononane-1,4,7-triacetic acid and feasibility studies in mice. *J Nucl Med* 2008;49:830.
20. Studer M, Meares CF. Synthesis of novel 1,4,7-triazacyclononane-*N,N',N''*-triacetic acid derivatives suitable for protein labeling. *Bioconjug Chem* 1992;3:337.
21. McMurry TJ, Brechbiel M, Wu C, et al. Synthesis of 2-(p-thiocyanatobenzyl)-1,4,7-triazacyclononane-1,4,7-triacetic acid: Application of the 4-methoxy-2,3,6-trimethylbenzenesulfonamide protecting group in the synthesis of macrocyclic polyamines. *Bioconjug Chem* 1993;4:236.
22. Jeong JM, Chung JK. Therapy with ¹⁸⁸Re-labeled radiopharmaceuticals: An overview of promising results from initial clinical trials. *Cancer Biother Radiopharm* 2003;18:707.
23. Pillai MR, Chakraborty S, Das T, et al. Production logistics of ¹⁷⁷Lu for radionuclide therapy. *Appl Radiat Isot* 2003;59:109.
24. Jeong JM, Knapp FF Jr. Use of the Oak Ridge National Laboratory tungsten-188/rhenium-188 generator for preparation of the rhenium-188 HDD/lipiodol complex for trans-arterial liver cancer therapy. *Semin Nucl Med* 2008;38: S19.
25. Chakravarty R, Pandey U, Manolkar RB, et al. Development of an electrochemical ⁹⁰Sr-⁹⁰Y generator for separation of ⁹⁰Y suitable for targeted therapy. *Nucl Med Biol* 2008;35:245.
26. Cacheris WP, Nickle SK, Sherry AD. Thermodynamic study of lanthanide complexes of 1,4,7-triazacyclononane-*N,N',N''*-triacetic acid and 1,4,7,10-tetraazacyclododecane-*N,N',N'',N'''*-tetraacetic acid. *Inorg Chem* 1987;26:958.
27. Jeong JM, Kim YJ, Lee YS, et al. Radiolabeling of NOTA and DOTA with positron emitting ⁶⁸Ga and investigation of *in vitro* properties. *Nucl Med Mol Imaging* 2009;43:330.

