Epstein-Barr Virus, Beta-Catenin, and E-cadherin in Gastric Carcinomas

Activated beta-catenin is suggested to inhibit NF-kappaB activation, and we previously demonstrated that NF-kappaB nuclear positivity was more frequent in Epstein-Barr virus (EBV)-infected gastric carcinomas. It is controversial that beta-catenin and E-cadherin are prognostic markers in gastric carcinomas. To define a relationship between beta-catenin and EBV, and the prognostic value of beta-catenin and E-cadherin, we analyzed in situ hybridization for EBV-encoded small RNAs, betacatenin, and E-cadherin immunohistochemistry, and clinicophatological features in 111 gastric carcinomas, EBV infection was detected in seven carcinomas (6.3%): none of seven showed beta-catenin nuclear accumulation, and five out of seven revealed beta-catenin membranous loss or cytoplamic expression. Eighty cases (72.1%) showed beta-catenin alteration; i.e., loss of membrane staining in 65 (58.6 %), cytoplasmic expression in 35 (31.5%), and nuclear accumulation in 15 (13.5%). E-cadherin alteration was observed in 34 cases (30.6%) and correlated with betacatenin alteration. On multivariate analysis, the combined immunoexpression group of beta-catenin nuclear accumulation/ E-cadherin alteration and the advanced TNM cancer stage group showed poor patient's survival (p<0.05). In conclusion, betacatenin activation through nuclear accumulation hardly occurred in EBV-infected gastric carcinomas. The combined immunoexpression pattern of beta-catenin and E-cadherin can be used as a prognostic marker in gastric carcinomas.

Key Words : Stomach Neoplasms; Herpesvirus 4, Human; Beta Catenin; Cadherins; Prognosis

INTRODUCTION

Gastric cancer is one of the leading fatal malignancies worldwide (1), and the most common malignancy in Korea (2). Now, gastric cancer has been reported to be the second leading cause of death due to malignancy in Korea, which was the first leading cause before 2005 (2). However, the etiology of gastric cancer has not yet been unveiled completely, and the previous studies have provided evidence that risk factors such as diet, infection including *Helicobacter pylori* and Epstein-Barr virus, and hereditary trait may be involved in (3). Recently, accumulation of molecular events has been known to be important in the development of gastric cancer (4). Furthermore, these biological markers as well as traditional histopathological parameters can be used as prognostic factors (4-8).

It was suggested that beta-catenin can regulate NF-kappaB negatively (9), and our previous study revealed that NFkappaB nuclear positivity was more frequent in Epstein-Barr virus (EBV)-positive gastric carcinomas than conventional gastric carcinomas (10).

Beta-catenin, a 92-kDa protein (11), plays multifunctional roles. First, beta-catenin is a component of the cell-cell adhesion complex, as an E-cadherin/beta-catenin complex. In Mok Jung^{*,†}, Jung Kee Chung^{*,†}, Young A Kim^{1, §}, Je Eun Kim^{1, §}, Seung Chul Heo^{*,†}, Young Joon Ahn^{*,†}, Ki-Tae Hwang^{*,†}, Byeong Gwan Kim^{1, 1}, Kook Lae Lee^{1, ¶}, Chul Woo Kim[§], Woo Ho Kim[§], Mee Soo Chang^{1, §}

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Secondly, beta-catenin acts as a coactivator of transcription factors involved in the *Wnt* signaling pathway (11). *Wnt* signaling is mediated by secreted proteins that interact with specific cell-membrane receptors and is involved in the growth and differentiation of cells during development. *Wnt* pathway dysregulation has been implicated in many cancers (11). Uncontrolled activation of this signaling pathway may induce inappropriate proliferation of target cells and may contribute to the development of malignancy. Dysfunction of this regulatory pathway may result in the accumulation of a hypophosphorylated stable form of beta-catenin in the cytoplasm (11), from whence it translocates to the nucleus, binding to the high mobility group domain factors Tcf/LEF (12) and stimulating transcription of target genes such as c-myc and cyclin D1 (13).

E-cadherin is a 120-kDa transmembrane glycoprotein that is responsible for calcium-dependent intercellular adhesion by homotypic interactions (14), and hence plays a critical role in cancer cell metastasis and invasion because anchorage of cells to substrate is critical for the integrity of many cell types including epithelial cells. Becker et al. first disclosed that E-cadherin is implicated in gastric carcinogenesis (15). Functional cadherin-dependent cell adhesion requires the formation of complexes between E-cadherin and catenins. However, cell growth suppressor activity of E-cadherin is not adhesion-dependent. E-cadherin protein can directly transduce a growth inhibitory signal, through the inhibition of beta-catenin signaling (16).

Previous immunohistochemical studies have shown a close relationship between alteration of E-cadherin or bata-catenin and lower 5-yr survival rates in diverse human cancers such as breast, urinary bladder, prostate, and esophagus cancer (17-20). However, there is a conflict concerning the prognostic significance of bata-catenin or E-cadherin in gastric cancer (5-8, 21-23).

In the present study, we investigated in situ hybridization for EBV-encoded small RNAs and immunoexpression of beta-catenin and E-cadherin proteins in 111 cases of gastric carcinomas, in order to define a relationship between betacatenin and EBV and to clarify the impact of beta-catenin or E-cadherin on the prognosis.

MATERIALS AND METHODS

Patients and tumor tissue

The present study was based on data from 111 Korean patients undergoing surgically resection for gastric carcinomas from 1990 to 1994 at the Seoul National University Boramae Hospital. Age, sex, tumor size and location, tumor differentiation according to the WHO classification, Lauren classification, and pathological tumor stage (TNM stage) according to the American Joint Committee on Cancer (AJCC) system were evaluated. Survival data were obtained from hospital records and patients' physicians in charge. The overall survival period of the patients was measured from the date of

operation to the date of death or the last follow-up.

EBER-in situ hybridization

Three micromete thick sections were cut from each paraffin block, deparaffinized and dehydrated, digested with proteinase K, and hybridized for two hours at 37°C with a fluorescein-conjugated EBV oligonucleotide probe targeting EBV-encoded RNA (EBER) (Novocastra, Newcastle-upon-Tyne, U.K.). Hybridization products were detected using an alkaline phosphatase-conjugated antibody to fluorescein isothiocyanate (affinity-isolated rabbit F[ab']). 5-Bromo-4chloro-3-indolylphosphate and nitroblue tetrazolium were used as an enzyme substrate to demonstrate the alkaline phosphatase activity. The slides were counterstained with Mayer's hematoxylin. Using a light microscope, black or dark navy-colored aggregates at the hybridization site were interpreted as positive staining (Fig. 1A). Rare reactive lymphocytes showed a positive signal of EBER, but only signals of EBER within the tumor cells were considered to be positive.



Fig. 1. Representative features of in situ hybridization for Epstein-Barr virus (A) and immunohistochemistry for beta-catenin (B-D) and Ecadherin (E-F). (A) Most of cancer cell nuclei reveal strong signals of black or dark navy aggregates on in situ hybridization for Epstein-Barr virus, while normal stroma cell nuclei produce no signal (\times 200). (B) Beta-catenin shows loss of membranous staining in cancer cells (left 2/3), while preserved membranous staining in normal epithelium (right 1/3) (\times 100). (C) Beta-catenin reveals cytoplasmic expression without nuclear accumulation in cancer cells (\times 200). (D) Beta-catenin demonstrates nuclear accumulation in cancer cells without cytoplasmic expression (\times 400). (E) E-cadherin discloses loss of membranous staining in cancer cells (right 2/3), while preserved membranous staining in normal epithelium (left 1/3) (\times 200). (F) E-cadherin exhibits well preserved membranous staining in cancer cells (\times 200).

Immunohistochemistry

Tumor tissue was retrieved from formalin-fixed, paraffinembedded specimen. After the antigen retrieval process by microwave, immunohistochemistry was performed with monoclonal antibodies against E-cadherin (1:100, Zymed Laboratories, San Francisco, CA, U.S.A.) and beta-catenin (1:100, Transduction Laboratories, Lexington, KY, U.S.A.), using avidin-biotin complex method. For an interpretation of immunohistochemical staining, cut-off point was adapted as reported previously (7, 21, 22, 24-26). Those cases with loss of membranous staining in more than 10% of tumor cells were considered to be an altered expression of E-cadherin protein (7, 24, 25). An altered expression of beta-catenin protein was divided into three patterns, that is, loss of membranous staining in more than 10% of tumor cells (7, 24, 25), cytoplasmic expression in any degree of tumor cells (21, 22, 26) and nuclear accumulation in any degree of tumor cells (21, 22, 26) patterns.

Statistics

All statistical analyses were conducted using the SPSS 10.0 statistical software program (SPSS, Chicago, IL, U.S.A.). The chi-square test or Fisher's exact test (two-sided), Pearson correlation (two-tailed), and Kendall's tau-b correlation were applied. The cumulative survival rates were obtained by the Kaplan-Meier analysis, and differences in survival were compared using the log-rank test. Multivariate analysis was performed using the Cox proportional hazards model. Values of p<0.05 were considered to be statistically significant.

RESULTS

Cilnicopathological features

Clinicopathological features are summarized in Table 1 and 2. The follow-up period ranged from 2 to 167 months, with a median of 50 months; thirty-five patients (31.5%) were dead and 76 (68.5%) were alive. Among the clinicopathological features evaluated in the present study, Lauren classification showed correlations with tumor location, lymph node metastasis (pN stage), TNM stage, and patient's survival (p<0.05).

EBV-infected gastric carcinomas

EBV infection was revealed in seven carcinomas (6.3%) (Fig. 1A), and none of EBV-infected gastric carcinomas show-

Table 1. Relationship between Epstein-Barr virus infection, beta-catenin alteration or E-cadherin alteration, and clinicopathological features in 111 cases of gastric carcinomas

	~	Epstein-Barr virus		beta-catenin		E-cadherin	
	n	Infection	<i>p</i> value	Alteration	<i>p</i> value	Alteration	<i>p</i> value
Sex			0.31		0.31		0.38
Male	76	6 (8%)		57 (75%)		21 (62%)	
Female	35	1 (3%)		23 (66%)		13 (38%)	
Age (yr)		. ,		х <i>У</i>		. ,	
Range	26-79	31-67		26-79		26-69	
Median	57	62		57		55	
Tumor size							
Range (cm)	1.0-14.0	2.0-8.0		1.0-14.0		2.0-10.0	
Mean (cm)	4.7 ± 2.27	5.6 ± 2.28		4.9±2.44		4.8 ± 2.17	
Tumor site			0.15		0.76		0.80
Low 1/3	81	3 (4%)		59 (73%)		25 (31%)	
Middle 1/3	29	4 (14%)		20 (69%)		9 (31%)	
Upper 1/3	1	0		1 (100%)		0	
Lauren			0.16		0.90		0.14
Intestinal	44	1 (2%)		32 (73%)		10 (23%)	
Diffuse	67	6 (9%)		48 (72%)		24 (36%)	
TNM stage*			0.81		0.75		0.87
I	33	0		25 (76%)		8 (24%)	
II	14	2 (14%)		8 (57%)		5 (15%)	
IIIA	18	1 (6%)		13 (72%)		6 (18%)	
IIIB	13	0		10 (77%)		5 (15%)	
IV	33	4 (12%)		24 (73%)		10 (29%)	
Patient outcome			0.08		0.98		0.38
Alive	76	7 (9%)		54 (71%)		21 (28%)	
Dead	35	0		26 (74%)		13 (37%)	

*Pathologic tumor stage according to the American Joint Committee on Cancer system.

Table 2. Relationship between three altered patterns of beta-catenin expression and clinicopathological features in 111 cases of gastric carcinomas

		beta-catenin alteration					
	n	Membranous loss (%)	<i>p</i> value	Cytoplasmic expression (%)	p value	Nuclear accu- mulation (%)	<i>p</i> value
Sex			0.84		0.65		0.10
Male	76	45 (59%)		25 (33%)		13 (17%)	
Female	35	20 (57%)		10 (29%)		2 (6%)	
Age (yr)							
Range	26-79	26-79		26-76		36-69	
Median	57	56		57		53	
Tumor size							
Range (cm)	1.0-14.0	1.0-10.0		1.0-14.0		1.5-14.0	
Mean (cm)	4.7 ± 2.27	4.8 ± 2.30		5.1±2.45		5.5 ± 3.40	
Tumor site			0.65		0.74		0.77
Low 1/3	81	48 (59%)		25 (31%)		12 (15%)	
Middle 1/3	29	16 (55%)		10 (34%)		3 (10%)	
Upper 1/3	1	1 (100%)		0		0	
Lauren			0.76		0.96		0.55
Intestinal	44	25 (57%)		14 (32%)		7 (16%)	
Diffuse	67	40 (58%)		21 (31%)		8 (12%)	
TNM stage*			0.66		0.16		0.85
I	33	22 (67%)		7 (21%)		4 (12%)	
II	14	6 (43%)		5 (36%)		3 (21%)	
IIIA	18	10 (56%)		9 (50%)		2(11%)	
IIIB	13	8 (61%)		2 (35%)		1 (8%)	
IV	33	19 (58%)		12 (36%)		5 (15%)	
Patient outcome			0.36		0.86		0.08
Alive	76	46 (61%)		23 (30%)		8 (11%)	
Dead	35	19 (54%)		12 (34%)		7 (2%)	

*Pathologic tumor stage according to the American Joint Committee on Cancer system.

 Table 3. Comparison of beta-catenin and E-cadherin immunoexpression between Epstein-Barr virus (EBV)-infected gastric carcinomas and non-infected carcinomas

	EBV-infected cases (n=7) (%)	Non-infected cases (n=104) (%)	p value
beta-catenin alteration Membranous loss Cytoplamic expression Nuclear accumulation E-cadherin alteration (membranous loss)	5 (71%) 3 (43%) 3 (43%) 0 1 (14%)	75 (72%) 62 (60%) 32 (31%) 15 (14%) 33 (32%)	0.97 0.38 0.50 0.28 0.33

ed beta-catenin nuclear accumulation. However, five out of seven EBV-infected carcinomas revealed beta-catenin alteration; three cases showed cytoplasmic expression, and two out of these three cases had loss of membranous staining simultaneously. Another two cases revealed loss of membranous staining. Regarding E-cadherin, only one EBV-infected carcinoma disclosed E-cadherin alteration. However, there were no statistical significances in beta-catenin or E-cadherin immunoexpression patterns between EBV-infected gastric carcinomas and non-infected carcinomas (Table 3).

The EBV infection status showed a marginal impact on

the patient outcome (p=0.08) (Table 2) but was not associated with beta-catenin or E-cadherin alteration.

Expression of beta-catenin and E-cadherin

There were no significant relationships between clinicopathological features and immunoexpresssion of beta-catenin or E-cadherin (Table 2, 3). A representative immunohistochemical staining pattern is shown in Fig. 1B-F. In normal gastric epithelium, both E-cadherin and beta-catenin showed cytoplasmic membranous staining (Fig. 1B, E). Eighty cases (72.1%) showed beta-catenin alteration, i.e. loss of membrane staining in 65 cases (58.6%), cytoplasmic expression in 35 cases (31.5%), and nuclear accumulation in 15 cases (13.5%) (Table 1, 2). Regarding concurrent cytoplasmic and nuclear beta-catenin expression, nine of 35 cases displayed both cytoplasmic and nuclear beta-catenin. In six additional cases (5.4%), beta-catenin was seen only in the nuclei, leading to a total of 15 cases (13.5%) with beta-catenin nuclear accumulation. E-cadherin alteration (loss of membrane staining) was observed in 34 cases (30.6%) of gastric carcinomas.

The E-cadherin alteration was statistically significantly correlated with beta-catenin alteration (p=0.003); in partic-



Fig. 2. Kaplan-Meier survival plots. Combined immunoexpression of beta-catenin nuclear pattern/E-cadherin (A), TNM tumor stage (B), and beta-catenin nuclear accumulation (C). (A) Group 4 (with a combined pattern of beta-catenin nuclear accumulation/E-cadherin alteration) shows a poor patient survival (p=0.001). Group 1, a group with a combined pattern of beta-catenin no nuclear accumulation/normal E-cadherin; Group 2, beta-catenin nuclear accumulation/normal E-cadherin; Group 3, beta-catenin no nuclear accumulation/E-cadherin alteration; and Group 4, beta-catenin nuclear accumulation/E-cadherin alteration. (B) Advanced TNM tumor stage group has a lower rate of patient survival (p=0.008). (C) Beta-catenin nuclear accumulation group has a marginal impact on the patient survival (p=0.077).

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	E-cadherin e	p	
	No alteration	Alteration	value
beta-catenin			0.003
No alteration	28	3	
Alteration	49	31	
beta-catenin membranous			0.012
Preservation	39	7	
Loss	38	27	
beta-catenin cytoplasmic			0.312
No expression	55	21	
Expression	22	13	
beta-catenin nuclear			0.040
No accumulation	70	26	
Accumulation	7	8	

ular, with beta-catenin membranous loss or nuclear accumulation (p=0.012 and p=0.040, respectively) (Table 4).

Survival analyses

The beta-catenin nuclear accumulation was associated with a poor patient survival in a subset of Lauren diffuse-type carcinomas. Additionally, in a subset of male patients, beta-catenin nuclear accumulation and E-cadherin alteration were related to a poor patient survival (p<0.05).

The combined expression pattern of beta-catenin/E-cadherin was reclassified into four combinations (considering Table 4); the combination of beta-catenin alteration/E-cadherin, the combination of beta-catenin membranous pattern/ E-cadherin, the combination of beta-catenin cytoplasmic pattern/E-cadherin, and the combination of beta-catenin nuclear pattern/E-cadherin. On univariate analysis, the combination of beta-catenin nuclear pattern/E-cadherin had a Table 5. Multivariate analysis of the prognostic factors for overall survival (Cox proportional hazards model) in the 111 patients with gastric carcinomas

Prognostic factors	Hazard ratio (95% confidence interval)	p value
TNM stage (AJCC system)		0.001
ll vs. l	1.044 (0.083-13.074)	
IIIA vs. I	9.095 (1.440-57.461)	
IIIB vs. I	14.923 (3.220-69.158)	
IV vs. I	22.371 (4.132-121.130)	
Combined pattern of beta-catenin		0.008
nucleus staining/ E-cadherin		
*Group 2 vs. Group 1	1.006 (0.178-5.688)	
Group 3 vs. Group 1	0.352 (0.092-1.350)	
Group 4 vs. Group 1	5.492 (1.621-18.601)	

*Group 1, a group with combined pattern of beta-catenin, no nuclear accumulation/normal E-cadherin; Group 2, beta-catenin nuclear accumulation/normal E-cadherin; Group 3, beta-catenin, no nuclear accumulation/E-cadherin alteration; Group 4, beta-catenin nuclear accumulation/E-cadherin alteration.

AJCC, American Joint Committee on Cancer.

significant impact on the patient survival (p<0.05) (Fig. 2A). In other words, a group with beta-catenin nuclear accumulation/E-cadherin alteration showed poor patient survival. Elaborating on this, in groups of combined beta-catenin nuclear pattern/E-cadherin, the patient survival rate of each group was 63% in beta-catenin nuclear accumulation/E-cadherin alteration, which contrasted with the remaining groups; 29%, 29%, and 21%, in a group with beta-catenin nuclear accumulation/normal E-cadherin, in a group with beta-catenin, and in a group with beta-catenin, no nuclear accumulation/ E-cadherin alteration, respectively (Fig. 2A).

On multivariate analysis, the combined beta-catenin nuclear pattern/E-cadherin, and cancer stage (TNM classifica-

DISCUSSION

In the present study, we suggest that beta-catenin activation through nuclear accumulation seldom occurs in EBVinfected gastric carcinomas, although beta-catenin expression can be altered in patterns of membranous loss and cytoplasmic expression. Furthermore, our previous paper showed that NFkappaB nuclear accumulation occurred more frequently in EBV-infected gastric carcinomas than in EBVnegative gastric carcinomas (p < 0.05) (10), suggesting EBVinfection could be related to NFkappaB activation. The betacatenin cytoplasmic stabilization is involved in the first step of beta-catenin activation, and nuclear transport is the second step (27), In the present study, it was not certain whether cytoplasmic expression might be related to activated form of beta-catenin or not, because we have not evaluated the free beta-catenin level in cytoplasm or beta-catenin-dependent gene expression level. However, beta-catenin nuclear accumulation can represent beta-catenin activation. Consequently, in EBV-infected gastric carcinomas, beta-catenin activation through nuclear accumulation hardly occurs, but NF- kappaB could be activated. It is plausible that NFkappaB activation might suppress beta-catenin activation in EBV-infected gastric carcinomas because previous papers suggest that beta-catenin activation can inhibit NFkappaB activation (9). Therefore, a negative crosstalk between betacatenin and NFkappaB activation seems to exist in EBVinfected gastric carcinomas.

In the present study, E-cadherin alteration (loss of membrane staining) was observed in 30.6% and beta-catenin alteration in 72.1% [loss of membrane staining in 58.6%, cytoplasmic expression in 31.5%, and nuclear staining in 13.5%] of gastric carcinomas. The figures of the E-cadherin alteration and beta-catenin nuclear accumulation are almost same as those in the paper by Grabsch et al., 30.7% and 13.2%, respectively (21). Additionally, the figures of the beta-catenin membranous loss, cytoplasmic expression and nuclear accumulation are similar to those in the paper by Nabais et al. (22, 26). Meanwhile, concerning beta-catenin, some studies did not evaluate nuclear accumulation of beta-catenin (5, 6, 8, 23, 28). However, beta-catenin nuclear accumulatin should be evaluated in the study of the Wnt signal activation because beta-catenin translocated into the nucleus is predominantly in an active and oncogenic form (11, 27, 29, 30). The Wnt signal activation induces beta-catenin stabilization and its accumulation in the cytoplasm, and then, beta-catenin is translocated to the nucleus, where it interacts with DNA binding proteins of the T-cell factor-lymphoid enhancer factor (Tcf/LEF) family, causing transcriptional activation of target genes (11, 27, 29, 30), such as c-myc and cyclin D1 oncogenes (13). Moreover, in the present study and others (26), certain cases showed beta-catenin only in the nuclear accumulation pattern, neither in cytoplasmic expression nor membranous loss. Therefore, beta-catenin nuclear accumulation should be assessed necessarily.

In the multivariate survival analysis, a group with combined beta-catenin nuclear accumulation/E-cadherin alteration showed a poor patient survival, compared to the other groups (Fig. 2A). In this combined pattern, oncogenic process by beta-catenin activation and cancer progression due to reduction of the cell-cell adhesion by E-cadherin loss can occur together and end up with a poor patient survival, accordingly. This result might apparently support previous suggestions that the beta-catenin or E-cadherin immunoexpression pattern correlates with patient survival (5-8), but the present study showed that only beta-catenin nuclear accumulation or only E-cadherin alteration had no significant impact on patient survival, or rather a combined pattern of betacatenin nuclear accumulation/E-cadherin alteration was an independent prognostic factor.

There have been controversies concerning the prognostic significance of E-cadherin or bata-catenin in gastric cancer; that is, Grabash et al. (21), Nabias et al. (22), and Joo et al. (23) showed no prognostic significance in their studies. This discrepancy might be partly due to the interpretation of immunohistochemical staining or case selection bias. First, we can find a fault with ignorance of beta-catenin nuclear accumulation in some papers (5, 6, 8, 23, 28). Secondly, subsets of cases can make wide differences. For example, in the present study, beta-catenin nuclear accumulation or E-cadherin alteration had a significant impact on patient survival in subsets such as Lauren diffuse-type patients and male patients, but a marginal impact in the total patient group as a whole. Additionally, Nabais et al. demonstrated that betacatenin or E-cadherin did not have a prognostic significance based upon different subsets of gastric carcinomas (22).

Previous studies analyzed the prognostic significance of each protein, beta-catenin or E-cadherin, rather than the combined pattern of beta-catenin/E-cadherin. To the best of our knowledge, there is a study about the combined expression pattern of beta-catenin/E-cadherin, in which it correlated with the patient survival in univariate analysis, but not in multivariate analysis (23), although it could be a critical drawback that beta-catenin nuclear accumulation was not analyzed in their study (23). Besides, the other combined pattern of abnormal E-cadherin/MUC1-positive was suggested to be an unfavorable prognostic factor (8).

In conclusion, beta-catenin expression can be altered in EBVinfected gastric carcinomas, but beta-catenin activation through nuclear accumulation hardly occurrs. The beta-catenin and E-cadherin alterations correlated with each other. The combined beta-catenin and E-cadherin pattern can be used as prognostic markers in Korean gastric carcinomas, and frequent alterations of beta-catenin and E-cadherin proteins are observed.

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