



저작자표시-비영리-변경금지 2.0 대한민국

이용자는 아래의 조건을 따르는 경우에 한하여 자유롭게

- 이 저작물을 복제, 배포, 전송, 전시, 공연 및 방송할 수 있습니다.

다음과 같은 조건을 따라야 합니다:



저작자표시. 귀하는 원저작자를 표시하여야 합니다.



비영리. 귀하는 이 저작물을 영리 목적으로 이용할 수 없습니다.



변경금지. 귀하는 이 저작물을 개작, 변형 또는 가공할 수 없습니다.

- 귀하는, 이 저작물의 재이용이나 배포의 경우, 이 저작물에 적용된 이용허락조건을 명확하게 나타내어야 합니다.
- 저작권자로부터 별도의 허가를 받으면 이러한 조건들은 적용되지 않습니다.

저작권법에 따른 이용자의 권리는 위의 내용에 의하여 영향을 받지 않습니다.

이것은 [이용허락규약\(Legal Code\)](#)을 이해하기 쉽게 요약한 것입니다.

[Disclaimer](#)

의학석사 학위논문

**Influence of Genetic Polymorphisms
on the Association between Phthalate
Exposure and Pulmonary Function in
Elderly Koreans**

국내 노령인구에서 프탈레이트
노출과 폐기능 간 관련성에 대한
유전자 다형성의 영향

2012년 8월

서울대학교 대학원
의학과 예방의학 전공
박 혜 인

국내 노령인구에서 프탈레이트
노출과 폐기능 간 관련성에 대한
유전자 다형성의 영향

지도교수 홍 윤 철

이 논문을 의학석사 학위논문으로 제출함
2012 년 8 월

서울대학교 대학원
의학과 예방의학 전공
박 혜 인

박혜인의 의학석사 학위논문을 인준함

2012 년 8 월

위 원 장 조 수 현 (인)

부위원장 홍 윤 철 (인)

위 원 조 비 룡 (인)

학위논문 원문제공 서비스에 대한 동의서

본인의 학위논문에 대하여 서울대학교가 아래와 같이 학위논문 제공하는 것에 동의합니다.

1. 동의사항

- ① 본인의 논문을 보존이나 인터넷 등을 통한 온라인 서비스 목적으로 복제할 경우 저작물의 내용을 변경하지 않는 범위 내에서의 복제를 허용합니다.
- ② 본인의 논문을 디지털화하여 인터넷 등 정보통신망을 통한 논문의 일부 또는 전부의 복제, 배포 및 전송 시 무료로 제공하는 것에 동의합니다.

2. 개인(저작자)의 의무

본 논문의 저작권을 타인에게 양도하거나 또는 출판을 허락하는 등 동의 내용을 변경하고자 할 때는 소속대학(원)에 공개의 유보 또는 해지를 즉시 통보하겠습니다.

3. 서울대학교의 의무

- ① 서울대학교는 본 논문을 외부에 제공할 경우 저작권 보호장치(DRM)를 사용하여야 합니다.
- ② 서울대학교는 본 논문에 대한 공개의 유보나 해지 신청 시 즉시 처리해야 합니다.

논문 제목: Influence of Genetic Polymorphisms on the Association between
Phthalate Exposure and Pulmonary Function in Elderly Koreans

학위구분: 석사 V□ · 박사 □
학 과: 예방의학과
학 번: 2010-23717
연 락 처: hyeyinpark@snu.ac.kr
저 작 자: 박 혜 인 (인)

제 출 일: 2012 년 8 월 3 일

서울대학교총장 귀하

ABSTRACT

Introduction: Phthalates are prevalent environmental exposure chemical that have raised concern regarding their various health effects, including pulmonary function. Meanwhile, elderly people are more susceptible to environmental exposure, and their decreasing lung function is an important health issue. The purpose of current study was to investigate the association between urinary phthalate metabolite levels and indices for pulmonary function and evaluate effect modification by genetic polymorphisms of oxidative-stress related genes, catalase (CAT), superoxide dismutase (SOD2), and myeloperoxidase (MPO) in elderly Koreans.

Methods: We conducted a panel study on 418 individuals over 60 years old in Seoul, Korea, and repeatedly measured urinary phthalate metabolite levels and ran pulmonary function tests. Genetic polymorphisms of CAT (rs769218 and rs769217), SOD2 (rs4880, rs2758331, and rs5746136) and MPO (rs2071409, rs7208693) were determined. Mixed effect model was used to investigate association of urinary phthalate metabolite levels with indices for pulmonary function and to examine the effect of CAT, SOD2 and MPO polymorphisms on the association.

Results: Inverse association was demonstrated between sum of mono-(2-ethyl-5-hydroxyhexyl) phthalate and mono-(2-ethyl-5-oxohexyl) phthalate (Σ DEHP) levels and FEV1/FVC (P=0.0275) or FEF25-75 (P=0.002) after adjusting for age, number

of months past after each previous measurements, sex, body mass index, cotinine, mean temperature and dew point. The effect of Σ DEHP on lung function was significant only in subjects with GG genotype of CAT rs769218 and CC genotype of CAT rs769217, TT genotype of SOD2 rs4880 and CC genotype of SOD2 rs2758331, AA genotype of MPO rs2071409 and GG genotype of MPO rs7208693. Having all significant genotypes in three genes compared to other combinations showed significant difference in the phthalate-pulmonary function association (p-value for interaction: 0.0176 for Σ DEHP and FEV1/FVC, 0.0663 and FEF25-75).

Conclusions: In conclusion, we found urinary phthalate metabolites levels associated with decreasing pulmonary function in elderly Koreans, and suggested the effect modification of certain CAT, SOD2 and MPO polymorphisms on the phthalates-lung function association.

Keywords: phthalates, pulmonary function, genetic polymorphisms, elderly

Student number: 2010-23717

CONTENTS

Abstract	i
Contents	iii
List of tables and figures	iv
List of abbreviations	vi
Introduction	1
Materials and Methods	
1. Study subjects and sampling.....	3
2. Measurement of urinary biomarkers.....	4
3. Measurement of pulmonary function.....	6
4. Measurement of <i>CAT</i> , <i>SOD2</i> and <i>MPO</i> polymorphisms.....	6
5. Measurement of seasonal factors	8
6. Statistical analysis	8
Results	
1. Baseline characteristics by sex	11
2. Distribution of biomarker levels	11
3. Effects of urinary phthalate metabolites on lung function	12
4. Influence of <i>CAT</i> , <i>SOD2</i> and <i>MPO</i> polymorphisms.....	13
5. Comparison of association by smoking status and bronchial / lung disease history.....	15
Discussion	
1. Mean levels of phthalate metabolites.....	16
2. Association between phthalate exposure and pulmonary function, and influence by genetic polymorphisms of oxidative stress marker genes	16
3. Possible mechanism for phthalate-pulmonary function association	17
4. Possible mechanisms for influence of oxidative stress marker genes in the phthalate-pulmonary function association	18
5. Comparison of exposure effect by smoking versus phthalates on pulmonary function ..	19
6. Strength and limitation of the study.....	20
Conclusion	23
References	43
Abstract in Korean	51

LIST OF TABLES AND FIGURES

Table 1: Baseline characteristics of study subjects by gender.....	24
Table 2: Distribution of urinary phthalate metabolite levels	25
Table 3a: Association of urinary phthalate metabolite levels and pulmonary function measurements, by repeated measure analysis	26
Table 3b: Association of urinary phthalate metabolite levels (in interquartile range) and pulmonary function measurements, by repeated measure analysis	27
Table 4a: Effect of <i>CAT</i> , <i>SOD2</i> , <i>MPO</i> genotypes on phthalate-pulmonary function association	28
Table 4b: Effect of <i>CAT</i> , <i>SOD2</i> , <i>MPO</i> genotypes on relationship between interquartile increase of phthalate metabolites and pulmonary function parameters	30
Table 5: Effect of <i>CAT</i> , <i>SOD2</i> , <i>MPO</i> haplotypes on phthalate-pulmonary function association	32
Table 6a: Effect of grouped <i>CAT</i> , <i>SOD2</i> , <i>MPO</i> haplotypes on phthalate-pulmonary function association, by comparing having all 3 significant diplotypes versus having none, one or two of the 3 diplotypes	33
Table 6b: Effect of grouped <i>CAT</i> , <i>SOD2</i> , <i>MPO</i> haplotypes on phthalate-pulmonary function association, by comparing each of the possible combinations of having significant diplotypes	34
Table 6c: Effect of grouped <i>CAT</i> , <i>SOD2</i> , <i>MPO</i> haplotypes in diplotype combinations, on relationship between interquartile increase of phthalate metabolites and pulmonary function parameters.....	35
Table 7: Mean levels of pulmonary function tests by genotypes.....	36
Table 8a: Mean levels of pulmonary function indices by smoking status	37

Table 8b: Association of urinary phthalate metabolite levels and pulmonary function measurements, by smoking status	38
Table 9a: Mean levels of pulmonary function indices by history of bronchial or lung disease	39
Table 9b: Association of urinary phthalate metabolite levels and pulmonary function measurements, on including or excluding subjects diagnosed or in current treatment of bronchial / lung disease	40
Table 10: Genetic information on selected <i>CAT</i> , <i>SOD2</i> , <i>MPO</i> SNPs.....	41
Fig.1: Linkage disequilibrium plots for selected <i>CAT</i> (rs769218-rs769217) and <i>SOD2</i> (rs4880-rs27583331-rs5746136) SNPs, using Japanese+Chinese SNP genotypes from HapMap data	42

LIST OF ABBREVIATIONS

MEHHP: mono-(2-ethyl-5-hydroxyhexyl) phthalate

MEOHP: mono-(2-ethyl-5-oxohexyl) phthalate

DEHP: di(2-ethylhexyl) phthalate

MnBP: mono-n-butyl-phthalate

DnBP: di-n-butyl phthalate

Σ DEHP: sum of MEHHP and MEOHP

FEV₁: forced expiratory volume in 1 second

FVC: forced vital capacity

FEV₁/FVC: ratio of FEV1 to FVC

FEF₂₅₋₇₅: forced expiratory flow between 25 and 75% of FVC

CAT: catalase

SOD: superoxide dismutase

MPO: myeloperoxidase

INTRODUCTION

Phthalate use and exposure are ubiquitous in industrialized countries, as they are most commonly used plasticizers, primarily in polyvinyl chloride (PVC). They are found in a wide variety of products including vinyl upholstery, food containers and wrappers, toys, floor tiles, lubricants, sealers and adhesives. Beyond use in PVC resins, they are also found in cosmetics such as perfume, eye shadow, moisturizer, nail polish, hair spray, and liquid soap, and as an inert ingredient in pesticides [1]. Phthalates contribute 10-60% of plastic products by weight because of their ability to increase flexibility and transparency [2].

Although most manifestation on the effect of phthalates on human health are focused on reproductive system such as decreased sperm production, infertility, and harming sexual development in male infants [3], increasing but limited evidence for effect of phthalate exposure on respiratory health has demanded attention for further investigation on the possible association [4-9].

Elderly are identified as susceptible group to environmental exposure [10], and investigating association between commonly exposed chemicals, namely phthalates, on elderly population's health is important especially in countries like South Korea where elderly population is rapidly increasing. It has not been demonstrated whether environmental exposure to phthalates contribute to reduction of pulmonary function in elderly, but the few epidemiologic studies on phthalate exposure and lung function in other subpopulation provide grounds for possible

association.

Oxidative stress is defined as an impaired balance between free radical production and antioxidant capacity resulting in excess oxidative products. Studies have demonstrated oxidative stress attributable to exposure of common environmental hazards including phthalates [11-14] and also investigated on induction of oxidative stress leading to airway inflammation [15, 16]. The role of key antioxidant enzymes, catalase (CAT), superoxide dismutase (SOD2), and myeloperoxidase (MPO) are important in the process of oxidative stress. Activities of such enzymes in acute lung injury or lung carcinogenesis have been suggested in previous reports [17, 18]. In addition, studies on genetic polymorphisms of oxidative stress markers with possible effect on pulmonary function are much recommended [19-22].

With given background information, further investigation for effect of environmental phthalate exposure on pulmonary function in elderly population is necessary, and identifying possible biological explanation is just as important. Therefore, we conducted repeated pulmonary function tests and analyzed urinary samples in Korean elderly to evaluate on the association between phthalates exposure and pulmonary function, and attempted to explore the effect of genetic polymorphisms of oxidative stress markers, *CAT*, *SOD2* and *MPO*, on the association.

MATERIALS AND METHODS

1. Study subjects and sampling

Five hundred and sixty participants over 60 years old and residing in Seongbuk-gu, Seoul, Republic of Korea were enrolled in the Korean Elderly Environmental Panel (KEEP) study. In this research, we focused on 418 subjects with available measurements on their phthalate metabolite and pulmonary function test from September 2008 until November 2009. Among them, 410 subjects with available data on *CAT*, *SOD2* and *MPO* polymorphisms were selected for genotype and haplotype analysis.

We conducted pulmonary function test and obtained urine samples during three medical examinations. All urine samples were placed at -20°C immediately following collection and stored until analysis for the three biomarkers: mono-(2-ethyl-5-hydroxyhexyl) phthalate (MEHHP), mono-(2-ethyl-5-oxohexyl) phthalate (MEOHP) and mono-n-butyl-phthalate (MnBP). Also, we used a structured questionnaire at initial examination, and obtained detailed information on demographics, lifestyle habits, medical history and environmental exposure.

The study protocol was approved by the institutional review board of Seoul National University Hospital (IRB No. H-0804-045-241). Written consent was received from all participants and was confirmed by the IRB.

2. Measurement of Urinary Biomarkers

Phthalates

In this study, we measured urinary levels of MEHHP and MEOHP as metabolites of di(2-ethylhexyl) phthalate (DEHP), and MnBP as di-n-butyl phthalate (DnBP) metabolite. We used high performance liquid chromatography tandem mass spectrometry (Agilent 6410 triple Quad LCMS, Agilent, Santa Clara, CA, USA) according to previously reported procedures [11]. To determine MEHHP and MEOHP levels, 500 μ l of urine was buffered with 30 μ l of 2.0 M sodium acetate (pH 5.0), and then spiked with a mixture of isotope phthalate monoester standards (100 ng/ml) and 10 μ l of β -glucuronidase. The sample was incubated at 37 °C for 3 h to deconjugate the glucuronidated phthalate metabolites. After the incubation, 100 μ l of 2 N HCl was added to collect phthalate monoester. The extract was dried with nitrogen gas and reconstituted with 1 ml of HPLC-grade H₂O in a 2 ml glass vial. The supernatant was purified by solid phase extraction with disposable Agilent SB-C18 1.8 μ m (2.1 mm \times 50 mm). The mobile phase was 0.1% acetic acid water: 0.01% acetic acid acetonitrile (90:10, v/v) at a flow rate of 0.25 ml/min and the eluate was monitored at target masses of 221, 293 and 291, and internal standard masses of 225, 297 and 295.

Cotinine

Urinary cotinine levels were determined for monitoring tobacco exposure. After

urine samples were centrifuged to remove particulate matter, cotinine level was analyzed by an enzyme-linked immunosorbent assay method based upon the competitive binding to antibody of enzyme labeled antigen and unlabeled antigen (Bio-Quant cotinine direct Elisa, San Diego, CA, USA). Briefly, a 10 μ l of urine and 100 μ l of Horseradish peroxidase-labeled Cotinine derivative are incubated in micro-plate wells coated with high affinity cotinine antibody for 60 min at room temperature (RT). The wells are washed thoroughly and 100 μ l of chromogenic substrate added. After incubation for 30 min at RT, the color produced is stopped using 100 μ l of stop solution and the wells are read at 450 nm and 650 nm. The intensity of the color developed is inversely proportional to the concentration of drug in the sample.

Creatinine

For the adjustment of urinary volume, we measured urinary creatinine levels using a HITACHI 7600 instrument (HITACHI, Tokyo, Japan). Briefly, 10 μ l of urine and 300 μ l of picric acid solution (Wako, Osaka, Japan) were mixed at room temperature for 3 min, and absorbance of the solution was measured at 505 nm. Seventy-five μ l of alkaline solution (Wako, Osaka, Japan) was added for 4 min, and absorbance of the solution was measured at 570 nm. The mean of the two values measured at 505 nm and 570 nm was used as the creatinine level.

3. Measurement of Pulmonary Function

Spirometric Measurements

Spirometric testing was conducted every medical examination according to 2005 European Respiratory Society/American Thoracic Society recommendations. All tests were performed using a Microlab® (SensorMedic, Yorba Linda, CA, USA) spirometer by one trained technician, and one best result from three consecutive pulmonary function tests was taken for recording. From the test, we obtained values of forced expiratory volume in 1 second (FEV₁, L), forced vital capacity (FVC, L), a ratio of FEV₁ to FVC (FEV₁/FVC, %) and forced expiratory flow between 25 and 75% of the FVC (FEF₂₅₋₇₅, L/sec).

4. Measurement of *CAT*, *SOD2* and *MPO* polymorphisms

Genomic DNA was extracted from peripheral blood lymphocytes using a QIAamp DNA Blood Mini Kit (Qiagen, Valencia, CA, USA) and genetic polymorphisms of *CAT* (rs769218 and rs769217), *SOD2* (rs4880, rs2758331, and rs5746136) and *MPO* (rs2071409, rs7208693) were determined using the TaqMan fluorogenic 5' nuclease assay (ABI, Foster City, CA, USA). SNPs from each gene were chosen with standards of having minor allele frequency > 0.05% and having reported to have been researched by the NCBI.

The final volume of polymerase chain reaction (PCR) was 5 μ l, containing 10ng

of genomic DNA and 2.5ul TaqMan Universal PCR Master Mix, with 0.13ul of 40X Assay Mix (Assay ID for rs769218, rs769217, rs4880, rs2758331, and rs5746136, C_3102900_10, C_3102907_10 C_8709053_10, C_16288770_10, and C_29322854_10, respectively). Thermal cycle conditions were as follows: 50°C for 2 min to activate the uracil N-glycosylase and to prevent carry-over contamination, 95°C for 10 min to activate the DNA polymerase, followed by 45 cycles of 95°C for 15 s and 60°C for 1 min. All PCR were performed using 384-well plates by a Dual 384-Well GeneAmp PCR System 9700 (ABI, Foster City, CA, USA) and the endpoint fluorescent readings were performed on an ABI PRISM 7900 HT Sequence Detection System (ABI, Foster City, CA, USA). Negative controls were included to ensure accuracy of genotyping. For confirmation, five percent of the samples were randomly chosen and genotyped again, producing identical results.

When we tested for Hardy-Weinberg equilibrium (HWE) of *CAT*, *SOD2* and *MPO* gene polymorphisms, the study subjects were in HWE for all seven SNPs (all SNPs, $p > 0.05$ by χ^2 -test). Incomplete data missing at least one of two SNPs were excluded, and then individual haplotypes were estimated from genotype data using the PHASE program (ver. 2.0.2). Pairwise linkage disequilibrium (LD) between both polymorphic sites was estimated as relative disequilibrium (D').

5. Measurement of seasonal factors

Daily average outdoor temperature and dew point data measured at the Songwol-dong monitoring center in Seongbuk-gu, which is the nearest monitoring center to the subjects' residence, were obtained from the Korea Meteorological Administration (<http://www.kma.go.kr>). Data were matched to the date each subject took measurements for urinary phthalate metabolite levels and pulmonary function test.

6. Statistical Analysis

Baseline characteristics determined on subjects' first visits were compared by sex. Urinary concentrations and distributions of MEHHP, MEOHP and MnBP were calculated and the concentrations under limit of detection (LOD) were assigned as a default value of LOD by 2 in the biomarker analysis. For the analysis of DEHP and DBP, the metabolites of DEHP (MEHHP and MEOHP) were evaluated individually and as the sum of the two (represented as " Σ DEHP"). For each subject, the levels of MEHHP, MEOHP, Σ DEHP and MnBP that were averaged across repeatedly measured samples used to represent individual exposure to phthalates.

For association analyses, phthalate levels were log-transformed to improve normality. The effects of exposure to the phthalate metabolites, Σ DEHP and MnBP,

on pulmonary function were estimated using mixed model for repeated analysis with adjustment for age, sex, months past after each previous visit, body mass index, creatinine-adjusted cotinine level, mean temperature and mean dewpoint. We also estimated the association in separate sexes, and investigated on the interaction between sex and phthalate metabolites. Adjustment with months past after each previous visit was done to account for decline in lung function with aging [23].

To account for the non-random loss of follow-up due to different number of repeated measures [24], we adjusted for weight values yielded by inverse probability of attaining a follow-up response. With data from subjects with more than one visit, we performed logistic regression to predict follow-up probability (follow-up=1, missing=0), with covariates as previously measured age, sex, BMI, blood pressure, years of attending school, season of the year, and mean daily temperature [25, 26].

We also estimated changes of FEV_1/FVC and FEF_{25-75} by the increase of $\Sigma DEHP$ and $MnBP$ levels in different *CAT*, *SOD2* and *MPO* genotypes, and grouped their haplotypes based on effect estimates and p-values.

Because we used direct measurements of pulmonary function test instead of using answers to questionnaires on history of having lung disease, we performed sensitivity analysis to see differences upon including or excluding subjects with lung/bronchiolar diseases history. We also ran t-tests to check difference in levels of phthalate metabolites between subjects with and without record of pulmonary function tests.

To identify linkage disequilibrium, we calculated $|D'|$ values between each *CAT*, *SOD2* and *MPO* haplotypes from our data, and drew LD plots for each *CAT*, *SOD2* and *MPO* genotypes using Japanese+Chinese SNP genotype data from HapMap (available at <http://hapmap.ncbi.nlm.nih.gov>) to confirm our analysis.

In all statistical analyses, we used SAS version 9.3 (SAS Institute Inc., Cary, NC, USA). Haploview version 4.2 (available at <http://www.broadinstitute.org/haploview>) was used for drawing LD plots.

RESULTS

1. Baseline characteristics by sex

Average number of repeated visits made by each subject was about 2.39 times. Age, BMI, pulmonary function, history of smoking and history of bronchial or lung disease assessed on first visit day were compared by sex (Table 1). Of all 418 subjects, 109 (26.08%) were male and 309 (73.92%) were female. Mean age of study participants were 70.85 ± 4.18 for males and 69.84 ± 5.23 for females ($p=0.0437$), and their BMI was $24.43 \pm 2.71\text{kg/m}^3$ and $25.06 \pm 2.73\text{kg/m}^3$ respectively ($p=0.0371$). Mean cotinine levels also differed by sex, and males showed higher levels of pulmonary function than females. Smoking status also varied by sex, but no gender difference was found in bronchial or lung disease status. Genotype distributions of *CAT*, *SOD2* and *MPO* SNPs showed no significant difference by gender.

2. Distribution of biomarker levels

From 418 subjects, we obtained 999 urinary samples for analysis, with 2.39 average samples per subject. Urinary mean concentrations and distributions by percentiles of MEHHP, MEOHP, Σ DEHP and MnBP levels were calculated (Table 2). Mean concentration values for all phthalate metabolites were higher than their

median values, indicative of the high level in the upper quantiles. LOD values for MEHHP, MEOHP, and MnBP were 0.160 µg/L, 0.206 µg/L, and 0.256 µg/L, respectively.

3. Effects of urinary phthalate metabolites on lung function

On association analysis by repeated measure analysis (Table 3a & 3b), negative relationship was found between Σ DEHP levels and two parameters of pulmonary function test, FEV₁/FVC and FEF₂₅₋₇₅ (FEV₁/FVC: $\beta=-0.632$, P=0.0275; FEF₂₅₋₇₅: $\beta=-0.077$, P=0.002). As there were significant differences in the mean values of adjusted variables between males and females, we divided the subject by gender and performed same analysis by sex, and found that significance remained with female group only (P-values, for males: 0.3196 for FEV₁/FVC and 0.1365 FEF₂₅₋₇₅; for females: 0.0312 for FEV₁/FVC and 0.0023 FEF₂₅₋₇₅); however, on interaction analysis between sex and phthalate levels, significant interaction was not observed (p-values for interaction: 0.5179 for FEV₁/FVC, 0.5179 for FEF₂₅₋₇₅). No significant association was found between MnBP and any of the pulmonary function measurements, in all subjects (FEV₁/FVC: $\beta=-0.212$, P=0.4913; FEF₂₅₋₇₅: $\beta=-0.025$, P=0.3457) and in gender-divided groups.

4. Influence of *CAT*, *SOD2* and *MPO* polymorphisms

On estimating the effect of *CAT*, *SOD2* and *MPO* genotypes on the phthalate-pulmonary function (Table 4a & 4b), association between Σ DEHP levels and pulmonary function was significant only in subjects with *CAT* GG genotype of rs769218 (FEV₁/FVC: β =-1.331, P=0.0082; FEF₂₅₋₇₅: β =-0.094, P=0.0284) and CC genotype of rs769217 (FEV₁/FVC: β =-1.332, P=0.008; FEF₂₅₋₇₅: β =-0.094, P=0.0283), *SOD2* TT genotype of rs4880 (FEV₁/FVC: β =-0.803, P=0.329; FEF₂₅₋₇₅: β =-0.091, P=0.0014) and CC genotype of rs2758331 (FEV₁/FVC: β =-0.843, P=0.0105; FEF₂₅₋₇₅: β =-0.089, P=0.0017), *MPO* AA genotype in rs2071409 (FEV₁/FVC: β =-0.705, P=0.0225; FEF₂₅₋₇₅: β =-0.089, P=0.0007) and GG genotype in rs7208693 (FEV₁/FVC: β =-0.607, P=0.0644; FEF₂₅₋₇₅: β =-0.066, P=0.0232).

Effect estimates and the number of subjects were almost equal among the significant genotypes in each of the three genes, suggesting possible linkage. When we calculated $|D'|$ values among the two SNPs in *CAT* and *MPO*, and among three SNPs in *SOD2*, both SNPs in *CAT*, *MPO* and two SNPs in *SOD2* (rs4880 and rs2758331) were found to be in LD ($|D'|=1$, p-value <0.0001 in both genes). The third SNP in *SOD2*, rs5746136, which did not show different nor significant effect estimates among different genotypes, had $|D'|$ values of 0.77 and 0.86 with rs4880 and 2758331, respectively, and was excluded from the analysis in combining genotypes.

We compared our results with the Hapmap data, and found that our data was not different from Japanese+Chinese data obtained from Hapmap based on frequency of each genotype and LDs of *CAT* SNPs rs769218-rs769217 and *SOD2* SNPs rs4880-rs2758331 ($|D'|=1$, p-value <0.0001 in both genes for Hapmap) (Fig.1). Both r-square values for the two selected SNPs in *CAT* and *SOD2* in HapMap were calculated to be 1.0, confirming complete linkage. For *MPO*, we were not able to extract Hapmap genotype data for rs7208693, but LD plotting with nearest available SNP (rs8082134) showed $|D'|=1$ and $r^2=0.030$. Because number of subjects and estimate size was similar among genotypes for rs2071409 and rs7208693, we assumed that the two SNPs in our data may also be closely linked.

Therefore, we interpreted that significant effect estimate in one genotype of a SNP from each gene in our data could represent the significant genotype in the other linked SNP. Using the linked structure, we paired the haplotypes producing risky diplotypes, i.e. GC-GC in *CAT*, TC-TC in *SOD2* and AG-AG in *MPO* (Table 5). We investigated if there is a dose-response relationship between number of risky diplotypes and effect estimates, and found that having all three significant diplotypes against other combinations (i.e. having none, one or two of the three significant diplotypes) showed marked difference in size of effect estimate and significance (Table 6a, 6b & 6c). This association also showed significant and borderline-significant p-values for interaction between the diplotype combination and phthalates ($p=0.0176$ for \sum DEHP and FEV₁/FVC, 0.0663 for \sum DEHP and FEF₂₅₋₇₅).

5. Comparison of association by smoking status and bronchial / lung disease history

We found that FEV₁/FVC measurements differed between current smokers, ex-smokers and never-smokers ($p < 0.0001$). Never-smokers had highest mean percentage of FEV₁/FVC (85.61%), followed by current smokers (82.42%) and ex-smokers (77.9%). No significant difference was observed between the groups for FEF₂₅₋₇₅ measurements (Table 8a). In sensitivity analysis upon subject groups divided by smoking status, inverse relationship was found between Σ DEHP and pulmonary function parameters in never-smokers, and no significant association was found in current smokers and ex-smokers for both Σ DEHP and MnBP with FEV₁/FVC nor FEF₂₅₋₇₅ (Table 8b).

We also investigated on any PTF measurement differences between those with no or already-cured history of bronchial or lung disease, with those currently diagnosed or in treatment. In all measurements summed, FEF₂₅₋₇₅ measures were different among the two groups (none/cured: 2.12 L/s, diagnosed/in treatment: 1.77 L/s; p -value=0.0186) (Table 9a). Upon including or excluding subjects currently diagnosed or in treatment with lung/bronchiolar disease history, we found no difference in size, direction or significance of effect estimate (Table 9b).

DISCUSSION

1. Mean levels of phthalate metabolites

Mean levels of phthalate metabolites, MEHHP, MEOHP and MnBP in our study is 25.59 μ g/L, 21.51 μ g/L, and 59.27 μ g/L, respectively, and is in range with the previous reports in Europe, USA, Japan and Taiwan [27].

2. Association between phthalate exposure and pulmonary function, and influence by genetic polymorphisms of oxidative stress marker genes

Results from our finding shows that phthalate metabolite levels, namely Σ DEHP (sum of MEHHP and MEOHP), is in association with pulmonary function parameters, FEV₁/FVC and FEF₂₅₋₇₅ in the repeated measure analysis in Korean elderly. This association was found more clearly with certain *CAT*, *SOD2* and *MPO* genotype groups, where two of the SNPs in each groups are in complete or near-complete linkage disequilibrium.

To our knowledge, there has been only a single reported epidemiologic study that investigated on gene-environment interaction, where authors attempted to find association between phthalate exposure and glutathione S-transferase M1 polymorphism in estrogen-dependent diseases, and found some significant interaction [28]. We also investigated on possible effect modification of genetic

polymorphism on phthalate-pulmonary function association, focusing on relevant polymorphisms for antioxidant markers. We have also found significant effect of genetic polymorphism, and p-values for gene-phthalate interactions were significant.

3. Possible mechanism for phthalate-pulmonary function association

DEHP is a high molecular weight phthalate that is first metabolized to hydrolytic monoesters in phase I biotransformation, followed by enzymatic oxidation of the alkyl chain, then transformed to more hydrophilic and oxidative metabolites. These products mostly undergo phase II biotransformation and produce glucuronide conjugates with increased water solubility and therefore increased urinary excretion [3]. Therefore, we assume that the notable effect of oxidative stress-related genetic polymorphisms on the significant association between Σ DEHP and pulmonary function is acceptable, based on the metabolic background.

Our result does not support for significant relationship between MnBP and pulmonary function measures. We assumed that the difference in the results may be due to the different metabolizing processes the short- and long-chain branched phthalates undertake; it is a well-documented fact that MEHHP and MEOHP, long-chain branched phthalates, continue further hydroxylation and oxidation, while short-chain branched MnBP does not [29, 30]. This assumption may be further supported by our extended gene-environment analysis, which showed influence of oxidative stress marker genes only in the Σ DEHP-pulmonary function association.

Additional epidemiological researches in various populations and settings may be needed to confirm the different association in short- and long-chain branched phthalates with lung function.

4. Possible mechanisms for influence of oxidative stress marker genes in the phthalate-pulmonary function association

Of the seven SNPs in our study, *SOD2* rs4880 and *MPO* rs7208693 were polymorphisms with missense mutation. By NCBI search, *SOD2* rs4880 (Ala16Val) was the only SNP with available information on clinical significance, categorized as “probable-pathogenic” based on 45 human researches (Table 11).

Researchers reported on influence of rs4880 in relation to susceptibility to cardiomyopathy and microvascular complications of diabetes mellitus, and suggested that the Val16 allele disrupts the alpha-helix structure of SOD2 and causes the protein to be retained at the level of the mitochondrial inner membrane, and that the mutant protein has lower activity and increases susceptibility to oxidative stress. Although inconsistent, influence of rs4880 was also reported in relation to response to chemotherapy in breast, colorectal and prostate cancer patients, aging and longevity [31]. In regard to pulmonary function, a single study reported association between increasing number of genotypes coding for high antioxidative enzyme activity, including rs4880 Val/Val, and FEV₁ % predicted in young adult smokers [32].

To summarize, existing reports sufficiently support our findings on the influence of rs4880 in the association between phthalate exposure and pulmonary function parameters. Only a single report was found for *MPO* rs7208693 that suggested its effect on susceptibility to benzene poisoning, but did not have adequate information [33]. Further investigation of clinical significance on this non-synonymous mutation would be encouraging.

No report was available for the other SNPs used in our analysis regarding associations with toxicological or epidemiological findings

5. Comparison of exposure effect by smoking versus phthalates on pulmonary function

Although our sensitivity analysis does not show significant difference in the phthalate-pulmonary function association between ex- and never-smokers, this result cannot be confirmative as sample sizes for current and ex-smokers are small in our study. Also, we were not able to take into account the duration of smoking in our analysis and are not exempt from possible measurement bias.

Smoking is nonetheless a major exogenous influential factor to pulmonary function decline [34, 35], and it is expected to produce more severe results in the elderly individual, and even more pronounced in those with chronic respiratory disease [36]. It has been reported that rate of decline in forced vital capacity by age is about 21 to 34 mL/year in men and 19 to 29 mL/year in women [37], and another

study also reported that decline in pulmonary function measures by smoking would be as much as up to 40% of the effect of aging [38]. Thus we can surmise that effect of smoking on pulmonary function in elderly to about 29 to 48 mL/year in men and 27 to 41 mL/year in women.

Our study shows no significant association results between phthalate exposure and FEV₁ and FVC, which are parameters for comparability with the smoking effect, and the effect estimates are in much larger scale (e.g. change in FEV₁ per IQR increase in Σ DEHP is -0.51 L (95% confidence limits: -1.3, 0.28)). Significant association results, however, shows some degree of phthalate effect on the pulmonary function; change in FEV₁/FVC per IQR increase in Σ DEHP is as much as -21.67% (-40.5, -2.84), and change in FEF₂₅₋₇₅ per IQR increase in Σ DEHP is about -2.64 L/sec (-4.29, -0.99) (Table 3b).

6. Strength and limitation of the study

Using direct measurement of representative phthalate metabolites and pulmonary function over questionnaires on exposure or outcome history add to strength of our study, as our hypothesis was based on association between acute exposure and short-term outcome. Our sensitivity analysis result, having no difference in association with inclusion or exclusion of subjects with lung/bronchiolar diseases history, i.e. long-term outcomes, may add support to this assumption.

It has been reported that time of day urine samples are collected significantly influenced measured concentration of phthalate metabolites [39]. In the KEEP Study, provision and collection of urine sample containers was done only between 1000-1200 hours, thus is in exception to this information error.

Major limitations to our study are small sample size and cross-sectional nature of the analysis. Availability of greater number of subjects would have improved external validity on the effect of genetic polymorphisms on the exposure-outcome association. Also, although repeated measure analysis increased power to our finding, we cannot deduce causal relationship.

Study subjects were mostly female non-smokers without any lung disease, and although our research aimed at finding effect of environmental phthalate exposure on susceptible group such as the elderly, we cannot imply the finding on other pulmonary-function related susceptible groups such as smokers and lung disease patients.

Of the pulmonary function parameters, our results showed associations with FEV_1/FVC and FEF_{25-75} , but not with FEV_1 and FVC . FEV_1 and FVC are indices for obstructive and restrictive lung diseases, and FEV_1/FVC represents severity of either obstructive or restrictive obstruction. The less-clinically used index, FEF_{25-75} , is defined by flow (or speed) of air coming out of the lung during the middle portion of a forced expiration. It is known to be most sensitive to early diagnosis of small airway disease, but shows much variability even in normal persons, with its range depending largely on sex and age [40]. Although not directly

implicational, reports on correlation of FEF_{25-75} with bronchial hyperactivity in children with asthma or allergic rhinitis, even in normal FEV_1 , could be related to our significant association finding with FEF_{25-75} [41, 42].

CONCLUSION

In conclusion, we found urinary phthalate metabolites levels associated with decreasing pulmonary function in elderly Koreans, and suggested the effect modification of certain CAT, SOD2 and MPO polymorphisms on the phthalates-lung function association.

Table 1. Baseline characteristics of study subjects by gender

		All (n=418)	Male (n=109)	Female (n=309)	P-value
		mean±SD / n (%)	mean±SD / n (%)	mean±SD / n (%)	
Age		70.11 ± 4.99	70.85 ± 4.18	69.84 ± 5.23	0.0437
BMI (kg/m ²)		24.89 ± 2.71	24.43 ± 2.60	25.06 ± 2.73	0.0371
Cotinine (mg /L)		316.32±1366.79	838.5±2114.4	95.48±787.1	0.0022
FEV ₁ (L)		1.92 ± 0.44	2.34 ± 0.39	1.77 ± 0.36	<.0001
FVC (L)		2.27 ± 0.57	2.89 ± 0.47	2.05 ± 0.42	<.0001
FEV ₁ /FVC (%)		85.21 ± 7.00	81.49 ± 7.41	86.53 ± 6.36	<.0001
FEF ₂₅₋₇₅ (L/sec)		2.17 ± 0.72	2.37 ± 0.78	2.10 ± 0.69	0.0014
Smoking	Current smoker	26 (6.65)	25 (22.94)	1 (0.32)	
	Ex-smoker	26 (6.65)	23 (21.1)	3 (0.97)	<.0001
	Never-smoker	366 (93.35)	61 (55.96)	305 (98.71)	
Bronchial or lung disease	None / cured	409 (97.8)	108 (99.1)	301 (97.4)	
	Diagnosed / in treatment	9 (2.2)	1 (0.9)	8 (2.6)	0.4565
rs769218	GG	120 (28.78)	31 (28.44)	89 (28.9)	
	GA	225 (53.96)	57 (52.29)	168 (54.55)	0.6716
	AA	72 (17.27)	21 (19.27)	51 (16.56)	
rs769217	TT	69 (16.79)	21 (19.44)	48 (15.84)	
	CT	221 (53.77)	56 (51.85)	165 (54.46)	0.5392
	CC	121 (29.44)	31 (28.7)	90 (29.7)	
rs4880	TT	324 (77.88)	87 (79.82)	237 (77.2)	
	CT	86 (20.67)	21 (19.27)	65 (21.17)	0.5141
	CC	6 (1.44)	1 (0.92)	5 (1.63)	
rs2758331	CC	327 (78.61)	87 (79.82)	240 (78.18)	
	CA	84 (20.19)	21 (19.27)	63 (20.52)	0.6841
	AA	5 (1.2)	1 (0.92)	4 (1.3)	
rs5746136	GG	142 (34.22)	39 (35.78)	103 (33.66)	
	GA	207 (49.88)	54 (49.54)	153 (50)	0.6206
	AA	66 (15.9)	16 (14.68)	50 (16.34)	
rs2071409	AA	361 (86.99)	91 (84.26)	270 (87.95)	
	AC	52 (12.53)	17 (15.74)	35 (11.4)	0.4456
	CC	2 (0.48)	0 (0)	2 (0.65)	
rs7208693	GG	330 (79.52)	88 (80.73)	242 (79.08)	
	GT	77 (18.55)	20 (18.35)	57 (18.63)	0.5575
	TT	8 (1.93)	1 (0.92)	7 (2.29)	

FVC: forced vital capacity, FEV₁: forced expiratory volume in one second, FEF₂₅₋₇₅: forced expiratory flow between 25% and 75% of forced vital capacity.

Table 2. Distribution of urinary phthalate metabolite levels

Biomarker	No. of observations	Mean (SD)	LOD	n < LOD (%)	Selected percentiles					
					10th	25th	50th	75th	90th	95th
MEHHP (µg/L)	999	25.59 (41.8)	0.16	1(0.10)	5.4	9.15	16.15	28.85	50.3	75.05
MEOHP (µg/L)	999	21.51 (39.8)	0.206	17(1.70)	5.32	7.81	13.1	23.74	42.03	61.64
∑DEHP (µg/L)	999	49.1 (75.8)	-	-	11.22	17.37	29.38	51.66	96.65	131
MnBP (µg/L)	999	59.27 (79.57)	0.256	1(0.10)	14.52	22.66	38.9	65.38	114.81	162.7

∑DEHP: sum of two DEHP metabolites, MEHHP and MEOHP

Table 3a. Association of urinary phthalate metabolite levels and pulmonary function measurements, by repeated measure analysis^{1,2}

		All (n=418)		Female (n=309)		Male (n=109)	
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value
Σ DEHP	FEV ₁	-0.015 (0.012)	0.187	-0.011 (0.012)	0.3695	-0.034 (0.031)	0.2801
	FVC	-0.005 (0.015)	0.7163	0.002 (0.016)	0.8929	-0.021 (0.038)	0.573
	FEV ₁ /FVC	-0.632 (0.286)	0.0275	-0.646 (0.299)	0.0312	-0.759 (0.76)	0.3196
	FEF ₂₅₋₇₅	-0.077 (0.025)	0.002	-0.081 (0.026)	0.0023	-0.093 (0.062)	0.1365
MnBP	FEV ₁	0.001 (0.013)	0.9249	0.005 (0.013)	0.708	-0.006 (0.031)	0.8476
	FVC	0.007 (0.016)	0.6498	0.019 (0.018)	0.2824	-0.016 (0.037)	0.6748
	FEV ₁ /FVC	-0.212 (0.308)	0.4913	-0.478 (0.328)	0.1459	0.446 (0.758)	0.5575
	FEF ₂₅₋₇₅	-0.025 (0.027)	0.3457	-0.033 (0.029)	0.2522	-0.015 (0.062)	0.8098

¹ adjusted for age, sex (in analysis for both sex), months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dewpoint

² all *p*-interaction values for Σ DEHP and sex are more than 0.1.

Table 3b. Association of urinary phthalate metabolite levels (in interquartile range) and pulmonary function measurements, by repeated measure analysis ^{1,2}

		All (n=418)		Female (n=309)		Male (n=109)	
		change in PF (95% CI)	p-value	change in PF (95% CI)	p-value	change in PF (95% CI)	p-value
ΣDEHP (IQR)	FEV ₁	-0.51 (-1.3, 0.28)	0.187	-0.38 (-1.17, 0.41)	0.3695	-1.17 (-3.21, 0.88)	0.2801
	FVC	-0.17 (-1.16, 0.82)	0.7163	0.07 (-0.98, 1.12)	0.8929	-0.72 (-3.22, 1.78)	0.573
	FEV ₁ /FVC	-21.67 (-40.5, -2.84)	0.0275	-22.15 (-41.84, -2.47)	0.0312	-26.03 (-76.06, 24.01)	0.3196
	FEF ₂₅₋₇₅	-2.64 (-4.29, -0.99)	0.002	-2.78 (-4.49, -1.07)	0.0023	-3.19 (-7.27, 0.89)	0.1365
MnBP (IQR)	FEV ₁	0.04 (-1.05, 1.13)	0.9249	0.21 (-0.87, 1.3)	0.708	-0.26 (-2.85, 2.34)	0.8476
	FVC	0.3 (-1.04, 1.64)	0.6498	0.81 (-0.7, 2.32)	0.2824	-0.68 (-3.78, 2.41)	0.6748
	FEV ₁ /FVC	-9.06 (-34.85, 16.73)	0.4913	-20.42 (-47.88, 7.04)	0.1459	19.05 (-44.42, 82.52)	0.5575
	FEF ₂₅₋₇₅	-1.07 (-3.33, 1.19)	0.3457	-1.41 (-3.84, 1.02)	0.2522	-0.64 (-5.83, 4.55)	0.8098

¹ adjusted for age, sex (in analysis for both sex), months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dewpoint

² all p-interaction values for ΣDEHP and sex are more than 0.1.

(Interquartile range (IQR): ΣDEHP 34.29 μg/L, MnBP 42.72μg/L. Pulmonary function (PF) parameters: FEV₁ and FVC in liters (L), FEV₁/FVC in percentage (%), FEF₂₅₋₇₅ in liters per second (L/s).

Table 4a. Effect of *CAT*, *SOD2* genotypes on phthalate-pulmonary function association ¹

<i>CAT</i> (rs769218)								
		GG (n=120)		GA (n=225)		AA (n=72)		P-int
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	
Σ DEHP	FEV ₁ /FVC	-1.331 (0.496)	0.008 2	-0.51 (0.436)	0.242 6	0.484 (0.561)	0.390 6	0.071 8
	FEF ₂₅₋₇₅	-0.094 (0.042)	0.028 4	-0.06 (0.037)	0.109 3	-0.089 (0.053)	0.097 9	0.837 1
	MnBP	FEV ₁ /FVC	-0.519 (0.598)	0.387 1	-0.212 (0.435)	0.627 2	0.271 (0.603)	0.653 7
	FEF ₂₅₋₇₅	-0.031 (0.051)	0.551 1	-0.005 (0.037)	0.894	-0.059 (0.056)	0.294 2	0.669 8
<i>CAT</i> (rs769217)								
		TT (n=69)		CT (n=221)		CC (n=121)		P-int
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	
Σ DEHP	FEV ₁ /FVC	0.515 (0.591)	0.385 7	-0.389 (0.436)	0.372 1	-1.332 (0.496)	0.008	0.065 6
	FEF ₂₅₋₇₅	-0.069 (0.054)	0.207 3	-0.049 (0.038)	0.197 4	-0.094 (0.042)	0.028 3	0.648 5
	MnBP	FEV ₁ /FVC	0.293 (0.621)	0.638 3	-0.175 (0.432)	0.686 5	-0.499 (0.597)	0.404 8
	FEF ₂₅₋₇₅	-0.07 (0.057)	0.223 6	-0.002 (0.038)	0.960 1	-0.03 (0.051)	0.555 5	0.556 1
<i>SOD2</i> (rs4880)								
		TT (n=324)		CT (n=86)		CC (n=6)		P-int
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	
Σ DEHP	FEV ₁ /FVC	-0.803 (0.329)	0.015 2	-0.234 (0.609)	0.702 5	n/a		0.383 1
	FEF ₂₅₋₇₅	-0.091 (0.028)	0.001 4	-0.057 (0.054)	0.294 4	0.014 (0.169)	0.939 4	0.178 7
	MnBP	FEV ₁ /FVC	-0.341 (0.353)	0.334	-0.151 (0.648)	0.815 8	n/a	
	FEF ₂₅₋₇₅	-0.039 (0.03)	0.199 7	-0.023 (0.057)	0.690 7	0.44 (0.159)	0.05	0.058 5
<i>SOD2</i> (rs2758331)								
		CC (n=327)		CA (n=84)		AA (n=5)		P-int
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	
Σ DEHP	FEV ₁ /FVC	-0.843 (0.328)	0.010 5	-0.18 (0.601)	0.765 2	n/a		0.383 1
	FEF ₂₅₋₇₅	-0.089 (0.028)	0.001 7	-0.06 (0.054)	0.265 5	n/a		0.178 7
	MnBP	FEV ₁ /FVC	-0.382 (0.352)	0.278	-0.072 (0.642)	0.911	4.861 (3.667)	0.316 2
	FEF ₂₅₋₇₅	-0.038 (0.03)	0.203 9	-0.027 (0.058)	0.647 1	0.363 (0.189)	0.195 6	0.058 5
<i>SOD2</i> (rs5746136)								

		GG (n=142)		GA (n=207)		AA (n=66)		P-int
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	
Σ DEHP	FEV ₁ /FVC	-0.653 (0.473)	0.169 4	-0.582 (0.382)	0.128 7	-0.767 (0.906)	0.400 1	0.914 9
	FEF ₂₅₋₇₅	-0.089 (0.042)	0.032 7	-0.062 (0.036)	0.084 5	-0.077 (0.062)	0.217 9	0.926 5
	MnBP	FEV ₁ /FVC	-0.358 (0.579)	0.537 7	-0.298 (0.373)	0.425 2	0.236 (1.07)	0.826 1
	FEF ₂₅₋₇₅	-0.064 (0.051)	0.209 9	-0.014 (0.035)	0.683 9	0.001 (0.072)	0.989 8	0.686 1
MPO (rs2071409)								
		AA (n=361)		AC (n=52)		CC (n=2)		P-int
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	
Σ DEHP	FEV ₁ /FVC	-0.705 (0.308)	0.022 5	-0.146 (0.763)	0.849 3	n/a		0.316 1
	FEF ₂₅₋₇₅	-0.089 (0.026)	0.000 7	-0.013 (0.078)	0.868 9	n/a		0.350 4
	MnBP	FEV ₁ /FVC	-0.135 (0.334)	0.685 8	-1.075 (0.821)	0.195 2	n/a	
	FEF ₂₅₋₇₅	-0.027 (0.028)	0.34	-0.068 (0.085)	0.430 8	n/a		0.715 5
MPO (rs7208693)								
		GG (n=330)		GT (n=77)		TT (n=8)		P-int
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	
Σ DEHP	FEV ₁ /FVC	-0.607 (0.327)	0.064 4	-0.777 (0.631)	0.221 7	-1.86 (1.513)	0.286 2	0.499 4
	FEF ₂₅₋₇₅	-0.066 (0.029)	0.023 2	-0.097 (0.049)	0.052 3	-0.538 (0.235)	0.083 7	0.773 3
	MnBP	FEV ₁ /FVC	-0.043 (0.343)	0.900 3	-0.654 (0.738)	0.377 9	-1.088 (1.58)	0.529
	FEF ₂₅₋₇₅	-0.003 (0.031)	0.920 3	-0.085 (0.057)	0.143 6	-0.342 (0.245)	0.235 6	0.324 3

¹ adjusted for age, sex, months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dewpoint

Table 4b. Effect of *CAT*, *SOD2* genotypes on relationship between interquartile increase of phthalate metabolites and pulmonary function parameters¹

		change in PF (95% CI)	<i>p</i> - value	change in PF (95% CI)	<i>p</i> - value	change in PF (95% CI)	<i>p</i> - value	p-int
CAT (rs769218)		GG (n=120)		GA (n=225)		AA (n=72)		
ΣDEHP	FEV ₁ /FVC	-45.64 (-78.98, - 12.3)	0.0082	-17.49 (-46.79, 11.81)	0.2426	16.6 (-21.11, 54.3)	0.3906	0.0718
	FEF ₂₅₋₇₅	-3.22 (-6.05, - 0.4)	0.0284	-2.06 (-4.54, 0.43)	0.1093	-3.05 (-6.61, 0.51)	0.0979	0.8371
MnBP	FEV ₁ /FVC	-22.21 (-72.24, 27.9)	0.3871	-9.07 (-45.48, 27.37)	0.6272	11.6 (-38.91, 62.07)	0.6537	0.6835
	FEF ₂₅₋₇₅	-1.33 (-5.59, 2.95)	0.5511	-0.21 (-3.31, 2.88)	0.894	-2.52 (-7.21, 2.17)	0.2942	0.6698
CAT (rs769217)		TT (n=69)		CT (n=221)		CC (n=121)		
ΣDEHP	FEV ₁ /FVC	17.66 (-22.06, 57.38)	0.3857	-13.34 (-42.64, 15.96)	0.3721	-45.67 (-79.01, - 12.34)	0.008	0.0656
	FEF ₂₅₋₇₅	-2.37 (-6, 1.26)	0.2073	-1.68 (-4.23, 0.87)	0.1974	-3.22 (-6.05, - 0.4)	0.0283	0.6485
MnBP	FEV ₁ /FVC	12.54 (-39.48, 64.51)	0.6383	-7.49 (-43.65, 28.7)	0.6865	-21.35 (-71.3, 28.67)	0.4048	0.6998
	FEF ₂₅₋₇₅	-3 (-7.76, 1.78)	0.2236	-0.09 (-3.27, 3.1)	0.9601	-1.28 (-5.55, 2.99)	0.5555	0.5561
SOD2 (rs4880)		TT (n=324)		CT (n=86)		CC (n=6)		
ΣDEHP	FEV ₁ /FVC	-27.53 (-49.65, - 5.42)	0.0152	-8.02 (-48.95, 32.91)	0.7025	n/a		0.3831
	FEF ₂₅₋₇₅	-3.12 (-5, -1.24)	0.0014	-1.95 (-5.58, 1.67)	0.2944	0.48 (-10.88, 11.84)	0.9394	0.1787
MnBP	FEV ₁ /FVC	-14.59 (-44.12, 14.99)	0.334	-6.46 (-60.71, 47.81)	0.8158	n/a		0.1661
	FEF ₂₅₋₇₅	-1.67 (-4.18, 0.85)	0.1997	-0.98 (-5.76, 3.79)	0.6907	18.83 (5.48, 32.11)	0.05	0.0585
SOD2 (rs2758331)		CC (n=327)		CA (n=84)		AA (n=5)		
ΣDEHP	FEV ₁ /FVC	-28.91 (-50.95, - 6.86)	0.0105	-6.17 (-46.56, 34.22)	0.7652	n/a		0.3831
	FEF ₂₅₋₇₅	-3.05 (-4.93, - 1.17)	0.0017	-2.06 (-5.69, 1.57)	0.2655	n/a		0.1787
MnBP	FEV ₁ /FVC	-16.35 (-45.79, 13.15)	0.278	-3.08 (-56.83, 50.68)	0.911	208 (-99.38, 514.7)	0.3162	0.1661

	FEF ₂₅₋₇₅	-1.63 (-4.14, 0.89)	0.2039	-1.16 (-6.01, 3.7)	0.6471	15.53 (-0.32, 31.33)	0.1956	0.0585
SOD2 (rs5746136)		GG (n=142)		GA (n=207)		AA (n=66)		
∑DEHP	FEV ₁ /FVC	-22.39 (-54.18, 9.4)	0.1694	-19.96 (-45.63, 5.72)	0.1287	-26.3 (-87.19, 34.59)	0.4001	0.9149
	FEF ₂₅₋₇₅	-3.05 (-5.87, -0.23)	0.0327	-2.13 (-4.55, 0.29)	0.0845	-2.64 (-6.81, 1.53)	0.2179	0.9265
MnBP	FEV ₁ /FVC	-15.32 (-63.77, 33.19)	0.5377	-12.75 (-43.96, 18.5)	0.4252	10.1 (-79.51, 99.67)	0.8261	0.7838
	FEF ₂₅₋₇₅	-2.74 (-7, 1.54)	0.2099	-0.6 (-3.53, 2.33)	0.6839	0.04 (-5.99, 6.07)	0.9898	0.6861
MPO (rs2071409)		AA (n=361)		AC (n=52)		CC (n=2)		
∑DEHP	FEV ₁ /FVC	-24.17 (-44.87, -3.47)	0.0225	-5.01 (-56.29, 46.27)	0.8493	n/a		0.3161
	FEF ₂₅₋₇₅	-3.05 (-4.8, -1.3)	0.0007	-0.45 (-5.69, 4.8)	0.8689	n/a		0.3504
MnBP	FEV ₁ /FVC	-5.78 (-33.73, 22.2)	0.6858	-46 (-114.67, 22.82)	0.1952	n/a		0.9538
	FEF ₂₅₋₇₅	-1.16 (-3.5, 1.19)	0.34	-2.91 (-10.02, 4.21)	0.4308	n/a		0.7155
MPO (rs7208693)		GG (n=330)		GT (n=77)		TT (n=8)		
∑DEHP	FEV ₁ /FVC	-20.81 (-42.79, 1.16)	0.0644	-26.64 (-69.05, 15.77)	0.2217	-63.78 (-165.47, 37.91)	0.2862	0.4994
	FEF ₂₅₋₇₅	-2.26 (-4.21, -0.31)	0.0232	-3.33 (-6.62, -0.03)	0.0523	-18.45 (-34.24, -2.65)	0.0837	0.7733
MnBP	FEV ₁ /FVC	-1.84 (-30.56, 26.88)	0.9003	-27.98 (-89.73, 33.85)	0.3779	-46.56 (-178.77, 85.82)	0.529	0.816
	FEF ₂₅₋₇₅	-0.13 (-2.72, 2.47)	0.9203	-3.64 (-8.4, 1.14)	0.1436	-14.63 (-35.12, 5.9)	0.2356	0.3243

¹ adjusted for age, sex, months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dewpoint
(Interquartile range (IQR): ∑DEHP 34.29 µg/L, MnBP 42.72 µg/L. Pulmonary function (PF) parameters: FEV₁ and FVC in liters (L), FEV₁/FVC in percentage (%), FEF₂₅₋₇₅ in liters per second (L/s).

Table 5. Effect of *CAT*, *SOD2*, *MPO* haplotypes on phthalate-pulmonary function association ¹

<i>CAT</i> (rs769218-rs769217)						
		GC-GC (n=120)		GC-AT, AT-AT, AT-AC (n=290)		P-int
		β (SE)	p-value	β (SE)	p-value	
ΣDEHP	FEV ₁ /FVC	-1.331 (0.496)	0.0082	-0.094 (0.355)	0.7913	0.1053
	FEF ₂₅₋₇₅	-0.094 (0.042)	0.0284	-0.052 (0.031)	0.0931	0.3581
MnBP	FEV ₁ /FVC	-0.519 (0.598)	0.3871	-0.031 (0.358)	0.9301	0.9107
	FEF ₂₅₋₇₅	-0.031 (0.051)	0.5511	-0.022 (0.031)	0.4759	0.9794
<i>SOD2</i> (rs4880- rs2758331)						
		TC-TC (n=323)		TC-CA, CC-CC, CA-CA (n=91)		P-int
		β (SE)	p-value	β (SE)	p-value	
ΣDEHP	FEV ₁ /FVC	-0.809 (0.33)	0.0146	-0.314 (0.571)	0.583	0.3537
	FEF ₂₅₋₇₅	-0.091 (0.028)	0.0014	-0.044 (0.051)	0.3869	0.0967
MnBP	FEV ₁ /FVC	-0.345 (0.353)	0.3294	-0.045 (0.622)	0.9419	0.2837
	FEF ₂₅₋₇₅	-0.039 (0.03)	0.1983	-0.012 (0.056)	0.8337	0.0585
<i>MPO</i> (rs2071409-rs7208693)						
		GA-GA (n=278)		GA-GC, GA-TA, GC-TA, TA-TA (n=134)		P-int
		β (SE)	p-value	β (SE)	p-value	
ΣDEHP	FEV ₁ /FVC	-0.723 (0.357)	0.0438	-0.478 (0.476)	0.3168	0.3521
	FEF ₂₅₋₇₅	-0.077 (0.031)	0.0143	-0.081 (0.041)	0.0507	0.8819
MnBP	FEV ₁ /FVC	-0.033 (0.375)	0.929	-0.463 (0.543)	0.3958	0.9278
	FEF ₂₅₋₇₅	-0.01 (0.033)	0.7653	-0.053 (0.047)	0.2578	0.687

¹ adjusted for age, sex, months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dewpoint

Table 6a. Effect of grouped *CAT*, *SOD2*, *MPO* haplotypes on phthalate-pulmonary function association, by comparing having all 3 significant diplotypes versus having none, one or two of the 3 diplotypes ¹

		GC-GC & TC-TC & GA-GA (n=64)		others (n=340)		
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	P-int
Σ DEHP	FEV ₁ /FVC	-2.169 (0.711)	0.0032	-0.221 (0.314)	0.4827	0.0176
	FEF ₂₅₋₇₅	-0.155 (0.059)	0.0103	-0.049 (0.028)	0.0749	0.0663
MnBP	FEV ₁ /FVC	-0.382 (1.037)	0.7138	-0.24 (0.321)	0.4554	0.8347
	FEF ₂₅₋₇₅	-0.012 (0.086)	0.8915	-0.034 (0.028)	0.2233	0.9048

¹ adjusted for age, sex, months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dew point

Table 6b. Effect of grouped *CAT*, *SOD2*, *MPO* haplotypes on phthalate-pulmonary function association, by comparing each of the possible combinations of having significant diplotypes ¹

		Having all 3 diplotypes (GC-GC & TC-TC & GA-GA) (n=64)		Having 2 of the 3 diplotypes (n=195)		Having 1 of the 3 diplotypes (n=124)		Having none of the 3 diplotypes (n=21)		P-int
		β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	β (SE)	<i>p</i> -value	
Σ DEHP	FEV ₁ / FVC	-2.169 (0.711)	0.0032	-0.34 (0.425)	0.4238	-0.088 (0.51)	0.864	-0.9 (1.344)	0.5105	0.037 8
	FEF ₂₅₋₇₅	-0.155 (0.059)	0.0103	-0.04 (0.038)	0.297	-0.072 (0.042)	0.094	-0.015 (0.153)	0.9227	0.094 8
MnBP	FEV ₁ / FVC	-0.382 (1.037)	0.7138	-0.499 (0.41)	0.2248	0.336 (0.569)	0.5564	-2.097 (1.588)	0.2016	0.497 8
	FEF ₂₅₋₇₅	-0.012 (0.086)	0.8915	-0.055 (0.036)	0.1327	0.007 (0.048)	0.886	-0.101 (0.184)	0.5893	0.336 7

¹ adjusted for age, sex, months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dew point

Table 6c. Effect of grouped *CAT*, *SOD2*, *MPO* haplotypes in diplotype combinations, on relationship between interquartile increase of phthalate metabolites and pulmonary function parameters¹

		Having all 3 diplotypes (GC-GC & TC-TC & GA-GA) (n=64)		Having 2 of the 3 diplotypes (n=195)		Having 1 of the 3 diplotypes (n=124)		Having none of the 3 diplotypes (n=21)		P-int
		change in PF (95% CI)	<i>p</i> -value	change in PF (95% CI)	<i>p</i> -value	change in PF (95% CI)	<i>p</i> -value	change in PF (95% CI)	<i>p</i> -value	
ΣDEHP	FEV ₁ /FVC	-74.38 (-122.16, -26.59)	0.0032	-11.66 (-40.22, 16.9)	0.4238	-3.02 (-37.29, 31.26)	0.864	-30.86 (-121.19, 59.47)	0.5105	0.0378
	FEF ₂₅₋₇₅	-5.31 (-9.28, -1.35)	0.0103	-1.37 (-3.93, 1.18)	0.297	-2.47 (-5.29, 0.35)	0.094	-0.51 (-10.8, 9.77)	0.9227	0.0948
MnBP	FEV ₁ /FVC	-16.32 (-103.15, 86.83)	0.7138	44.37 (86.08, 2.52)	0.2248	17.54 (-10.62, 45.65)	0.5564	24.35 (199.89, -151.28)	0.2016	0.4978
	FEF ₂₅₋₇₅	-0.51 (-7.71, 7.2)	0.8915	3.68 (8.28, -0.93)	0.1327	1.54 (0.95, 2.12)	0.886	2.05 (10.51, -6.41)	0.5893	0.3367

¹ adjusted for age, sex, months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dew point (Interquartile range (IQR): ΣDEHP 34.29 µg/L, MnBP 42.72 µg/L. Pulmonary function (PF) parameters: FEV₁ and FVC in liters (L), FEV₁/FVC in percentage (%), FEF₂₅₋₇₅ in liters per second (L/s).

Table 7. Mean levels of pulmonary function tests by genotypes.

		N	FEV ₁ /FVC		FEF ₂₅₋₇₅	
			mean (SD)	p-value	mean (SD)	p-value
CAT <i>(rs769218)</i>	GG	298	84.73 (8)	0.8628	2.09 (0.75)	0.5892
	GA	528	84.91 (7.64)		2.11 (0.7)	
	AA	172	85.12 (6.45)		2.16 (0.76)	
CAT <i>(rs769217)</i>	TT	166	85 (6.48)	0.8799	2.14 (0.76)	0.6686
	CT	518	85.02 (7.65)		2.13 (0.7)	
	CC	299	84.75 (8)		2.09 (0.75)	
SOD2 <i>(rs4880)</i>	TT	778	84.73 (7.67)	0.195	2.1 (0.73)	0.3894
	CT	199	85.8 (7.24)		2.18 (0.7)	
	CC	17	84.35 (4.74)		2.1 (0.53)	
SOD2 <i>(rs2758331)</i>	CC	787	84.78 (7.65)	0.2808	2.1 (0.73)	0.3075
	CA	193	85.67 (7.27)		2.19 (0.71)	
	AA	14	83.71 (4.87)		2.06 (0.57)	
SOD2 <i>(rs5746136)</i>	GG	344	84.56 (7.5)	0.0616	2.08 (0.72)	0.0044
	GA	495	85.43 (7.21)		2.18 (0.75)	
	AA	157	83.96 (8.64)		1.98 (0.65)	
MPO <i>(rs2071409)</i>	AA	361	84.84 (7.57)	0.9533	2.09 (0.72)	0.3095
	AC	52	85.06 (7.53)		2.2 (0.74)	
	CC	2	85 (4)		2.1 (0.73)	
MPO <i>(rs7208693)</i>	GG	330	84.82 (7.41)	0.5243	2.11 (0.72)	0.9751
	GT	77	85.26 (8.22)		2.12 (0.76)	
	TT	8	86.5 (7.12)		2.15 (0.63)	

Table 8a: Mean levels of pulmonary function indices by smoking status

		All visits			First visit			Second visit			Third visit		
		N	Mean (SD)	P-value	N	Mean (SD)	P-value	N	Mean (SD)	P-value	N	Mean (SD)	P-value
FEV ₁ / FVC	Current smoker	26	82.42 (8.39)	<0.0001	26	82.23 (8.41)	<0.0001	19	79.26 (8.54)	<0.0001	15	86.73 (6.56)	
	Ex-smoker	26	77.9 (9.79)		26	78.38 (7.16)		20	75.2 (12.33)		21	79.86 (9.79)	0.0004
	Never-smoker	366	85.61 (6.98)		366	85.91 (6.58)		268	83.86 (6.32)		238	87.12 (7.84)	
FEF ₂₅₋₇₅	Current smoker	26	2.34 (0.83)	0.0312	26	2.36 (0.79)	0.2629	19	2.16 (0.83)	0.5098	15	2.56 (0.89)	
	Ex-smoker	26	2.04 (0.81)		26	2.03 (0.68)		20	1.95 (0.88)		21	2.14 (0.92)	0.1259
	Never-smoker	366	2.1 (0.71)		366	2.17 (0.72)		268	1.97 (0.66)		238	2.15 (0.74)	

Table 8b: Association of urinary phthalate metabolite levels and pulmonary function measurements, by smoking status^a

		Current smoker (n=26)		Ex-smoker (n=26)		Never-smoker (n=366)		p- int
		β (SE)	p- value	β (SE)	p- value	β (SE)	p- value	
Σ DEHP	FEV ₁ /FVC	-0.208 (1.757)	0.9066	0.066 (1.641)	0.9683	-0.615 (0.288)	0.0335	0.4176
	FEF ₂₅₋₇₅	0.011 (0.159)	0.9433	-0.015 (0.114)	0.8956	-0.085 (0.026)	0.0009	0.5052
MnBP	FEV ₁ /FVC	3.462 (1.839)	0.0714	1.839 (2.204)	0.4103	-0.426 (0.307)	0.1653	0.1197
	FEF ₂₅₋₇₅	0.228 (0.168)	0.1875	0.149 (0.151)	0.332	-0.04 (0.027)	0.1382	0.3399

^a adjusted for age, sex, months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dew point

Table 9a: Mean levels of pulmonary function indices by history of bronchial or lung disease

		All visits			First visit			Second visit			Third visit		
		N	Mean (SD)	P-value	N	Mean (SD)	P-value	N	Mean (SD)	P-value	N	Mean (SD)	P-value
FEV ₁ /FVC	None / cured	409	84.92 (7.54)	0.5467	409	85.23 (6.98)	0.7038	299	83.15 (7.34)	0.0452	266	86.44 (8.2)	0.2584
	Diagnosed / in treatment	9	84 (8.22)		9	84.33 (8.54)		8	77.88 (6.56)		8	89.75 (4.95)	
FEF ₂₅₋₇₅	None / cured	409	2.12 (0.73)	0.0186	409	2.17 (0.72)	0.3182	299	2 (0.68)	0.0087	266	2.18 (0.76)	0.5623
	Diagnosed / in treatment	9	1.77 (0.7)		9	1.93 (0.73)		8	1.35 (0.55)		8	2.02 (0.69)	

Table 9b: Association of urinary phthalate metabolite levels and pulmonary function measurements, on including or excluding subjects diagnosed or in current treatment of bronchial / lung disease^a

		All subjects (n=418)		Excluding bronchial / lung disease history (n=409)	
		β (SE)	p- value	β (SE)	p- value
Σ DEHP	FEV ₁ /FVC	-0.632 (0.286)	0.0275	-0.581 (0.29)	0.0456
	FEF ₂₅₋₇₅	-0.077 (0.025)	0.002	-0.079 (0.025)	0.0017
MnBP	FEV ₁ /FVC	-0.212 (0.308)	0.4913	-0.166 (0.315)	0.5974
	FEF ₂₅₋₇₅	-0.025 (0.027)	0.3457	-0.023 (0.027)	0.3886

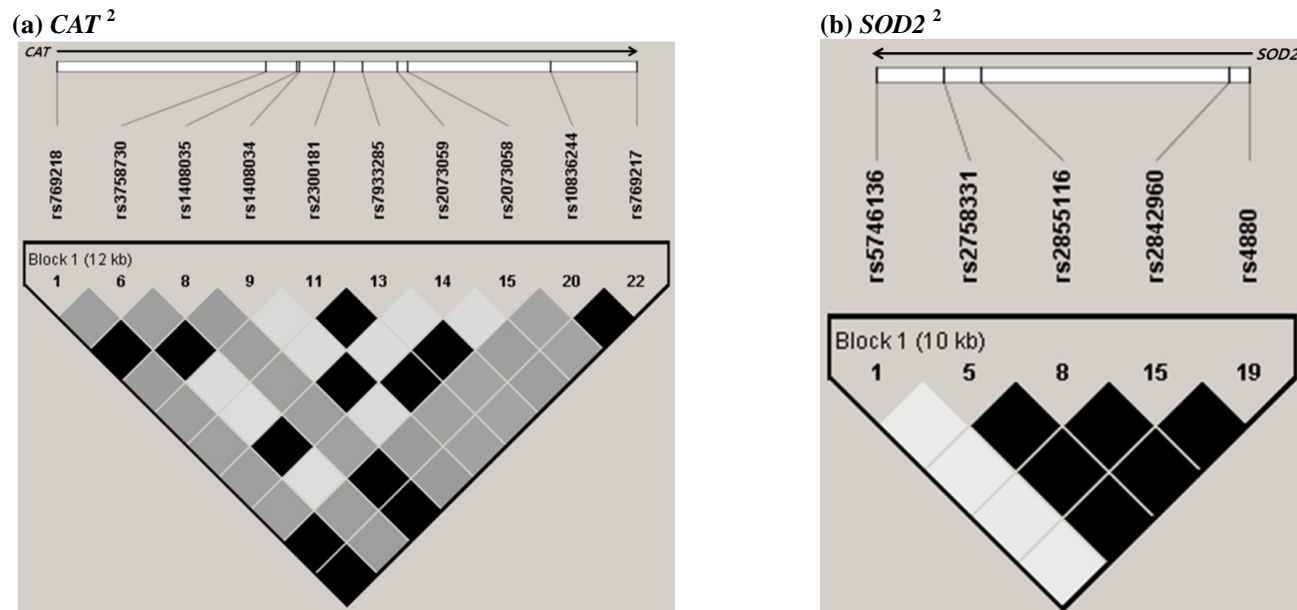
^a adjusted for age, sex, months past after each previous visit, BMI, creatinine-adjusted cotinine levels, mean temperature and mean dew point

Table 10. Genetic information on selected *CAT*, *SOD2*, *MPO* SNPs

Gene (ID)	SNP no.	Chromosome no.	Allele	Position	Function	SNP to mRNA	Allele change, at mRNA level	Residue change, at protein level	Clinical Significance *
CAT (847)	rs769218	11	GA	intron					n/a
	rs769217	11	CT	codon389	cds-synon	forward	GAC → GAT	D[Aspt]→D[Asp]	n/a
SOD2 (6648)	rs4880	16	CT	codon16	missense	forward	GTT → GCT	V[Val]→A[Ala]	Probable-pathogenic
	rs2758331	16	AC	intron					n/a
	rs5746136	16	CT	downstream	UTR-3	forward	n/a	n/a	n/a
MPO (4353)	rs7208693	17	GT	codon53	missense	reverse	GTC → TTC	V[Val]→F[Phe]	n/a
	rs2071409	17	GT	intron					n/a

* supported values of clinical significance are categorized into ‘unknown’, ‘untested’, ‘non-pathogenic’, ‘probable-non-pathogenic’, ‘probable-pathogenic’, ‘pathogenic’, ‘drug-response’, ‘histocompatibility’, and ‘other’.

Fig.1. Linkage disequilibrium plots for selected *CAT* (rs769218-rs769217) and *SOD2* (rs4880-rs2758331-rs5746136) SNPs using Japanese+Chinese SNP genotypes from HapMap data ¹



¹ LD plots are displayed in r^2 color scheme (black: $r^2=1$, shades of gray: $0 < r^2 < 1$, white: $r^2=0$), and SNPs with only $|D'|=1$ are presented. Arrows indicate direction of transcription.

² Ranges for r^2 in dark and light-grey colors are 0.356-0.397 and 0.135-0.149 respectively, and all LOD scores are greater than 3. Four SNPs (rs2076556, rs2268063, rs710395, rs1055981), although with minor allele frequency (MAF) >0.001 , were excluded from plotting as SNP pairing with these SNPs produced $|D'| < 1$.

³ Range for r^2 in light-grey colors is 0.081-0.085. LOD scores are greater than 3 in black colors, and 1.54-1.58 in light-grey colors

REFERENCES

1. Rudel RA, Perovich LJ. Endocrine disrupting chemicals in indoor and outdoor air. *Atmos Environ.* 2009 Jan 1;43(1):170-81.
2. Rakkestad KE, Dye CJ, Yttri KE, Holme JA, Hongslo JK, Schwarze PE et al. Phthalate levels in Norwegian indoor air related to particle size fraction. *J Environ Monit.* 2007 Dec;9(12):1419-25.
3. Hauser R, Calafat AM. Phthalates and human health. *Occup Environ Med.* 2005 Nov;62(11):806-18. Review.
4. Jaakkola JJ, Oie L, Nafstad P, Botten G, Samuelsen SO, Magnus P. Interior surface materials in the home and the development of bronchial obstruction in young children in Oslo, Norway. *Am J Public Health.* 1999 Feb;89(2):188-92.
5. Jaakkola JJ, Verkasalo PK, Jaakkola N. Plastic wall materials in the home and respiratory health in young children. *Am J Public Health.* 2000 May;90(5):797-9.
6. Ponsonby AL, Dwyer T, Kemp A, Cochrane J, Couper D, Carmichael A.

- Synthetic bedding and wheeze in childhood. *Epidemiology*. 2003
Jan;14(1):37-44.
7. Oie L, Nafstad P, Botten G, Magnus P, Jaakkola JK. Ventilation in homes and bronchial obstruction in young children. *Epidemiology*. 1999
May;10(3):294-9.
 8. Hoppin JA, Ulmer R, London SJ. Phthalate exposure and pulmonary function. *Environ Health Perspect*. 2004 Apr;112(5):571-4.
 9. Bornehag CG, Sundell J, Weschler CJ, Sigsgaard T, Lundgren B, Hasselgren M et al. The association between asthma and allergic symptoms in children and phthalates in house dust: a nested case-control study. *Environ Health Perspect*. 2004 Oct;112(14):1393-7.
 10. Risher JF, Todd GD, Meyer D, Zunker CL. The elderly as a sensitive population in environmental exposures: making the case. *Rev Environ Contam Toxicol*. 2010;207:95-157. Review.
 11. Hong YC, Park EY, Park MS, Ko JA, Oh SY, Kim H et al. Community level exposure to chemicals and oxidative stress in adult population. *Toxicol Lett*. 2009 Jan 30;184(2):139-44.
 12. Chitra KC, Latchoumycandane C, Mathur PP. Induction of oxidative stress

- by bisphenol A in the epididymal sperm of rats. *Toxicology*. 2003 Mar 14;185(1-2):119-27.
13. Bindhumol V, Chitra KC, Mathur PP. Bisphenol A induces reactive oxygen species generation in the liver of male rats. *Toxicology*. 2003 Jun 30;188(2-3):117-24.
 14. Singh VK, Patel DK, Jyoti, Ram S, Mathur N, Siddiqui MK. Blood levels of polycyclic aromatic hydrocarbons in children and their association with oxidative stress indices: an Indian perspective. *Clin Biochem*. 2008 Feb;41(3):152-61.
 15. Drost EM, Skwarski KM, Sauleda J, Soler N, Roca J, Agusti A et al. Oxidative stress and airway inflammation in severe exacerbations of COPD. *Thorax*. 2005 Apr;60(4):293-300.
 16. Seagrave J, Campen MJ, McDonald JD, Mauderly JL, Rohr AC. Oxidative stress, inflammation, and pulmonary function assessment in rats exposed to laboratory-generated pollutant mixtures. *J Toxicol Environ Health A*. 2008;71(20):1352-62.
 17. Cerutti P, Ghosh R, Oya Y, Amstad P. The role of the cellular antioxidant defense in oxidant carcinogenesis. *Environ Health Perspect*. 1994 Dec;102

Suppl 10:123-9. Review.

18. Roberts AM. Central role of oxidative stress and its signaling pathways in causing and preventing acute lung injury. *Crit Care Med*. 2011 Dec;39(12):2776-7.
19. Obeidat M, Wain LV, Shrine N, Kalsheker N, Soler Artigas M, Repapi E et al; SpiroMeta Consortium. A comprehensive evaluation of potential lung function associated genes in the SpiroMeta general population sample. *PLoS One*. 2011;6(5):e19382.
20. Masuko H, Sakamoto T, Kaneko Y, Iijima H, Naito T, Noguchi E et al. An interaction between Nrf2 polymorphisms and smoking status affects annual decline in FEV1: a longitudinal retrospective cohort study. *BMC Med Genet*. 2011 Jul 20;12:97.
21. Repapi E, Sayers I, Wain LV, Burton PR, Johnson T, Obeidat M et al. Genome-wide association study identifies five loci associated with lung function. *Nat Genet*. 2010 Jan;42(1):36-44.
22. Hancock DB, Eijgelsheim M, Wilk JB, Gharib SA, Loehr LR, Marcianti KD et al. Meta-analyses of genome-wide association studies identify multiple loci associated with pulmonary function. *Nat Genet*. 2010

Jan;42(1):45-52.

23. Ware JH, Dockery DW, Louis TA, Xu XP, Ferris BG Jr, Speizer FE. Longitudinal and cross-sectional estimates of pulmonary function decline in never-smoking adults. *Am J Epidemiol.* 1990 Oct;132(4):685-700.
24. Rubin DB. Inference and missing data. *Biometrika.* 1976 Dec;63(3):581-92.
25. Robins JM, Rotnitzky A, Zhao LP. Analysis of Semiparametric Regression Models for Repeated Outcomes in the Presence of Missing Data. *J Am Stat Assoc.* 1995;90(429):106-21.
26. McCracken J, Baccarelli A, Hoxha M, Dioni L, Melly S, Coull B et al. Annual ambient black carbon associated with shorter telomeres in elderly men: Veterans Affairs Normative Aging Study. *Environ Health Perspect.* 2010 Nov;118(11):1564-70.
27. Wittassek M, Koch HM, Angerer J, Brüning T. Assessing exposure to phthalates - the human biomonitoring approach. *Mol Nutr Food Res.* 2011 Jan;55(1):7-31. Review.
28. Huang PC, Tsai EM, Li WF, Liao PC, Chung MC, Wang YH et al. Association between phthalate exposure and glutathione S-transferase M1

polymorphism in adenomyosis, leiomyoma and endometriosis. Hum
Reprod. 2010 Apr;25(4):986-94.

29. Frederiksen H, Skakkebaek NE, Andersson AM. Metabolism of phthalates in humans. Mol Nutr Food Res. 2007 Jul;51(7):899-911. Review.
30. Wittassek M, Angerer J. Phthalates: metabolism and exposure. Int J Androl. 2008 Apr;31(2):131-8. Epub 2007 Dec 7. Review.
31. Online Mendelian Inheritance in Man, OMIM® . Johns Hopkins University, Baltimore, MD. MIM Number:147460:2011 Apr. Available from:
<http://omim.org>
32. Malling TH, Sigsgaard T, Brasch-Andersen C, Frischknecht L, Andersen HR, Kruse TA et al. Genetic polymorphisms in antioxidative enzymes are associated to forced expiratory volume in 1 s (FEV1) in smokers independently of asthma. Clin Respir J. 2012 Jan;6(1):46-55.
33. Sun P, Zhang Z, Wan J, Shao M. [Genetic polymorphisms of MPO, NQO1, GSTP1, UGT1A6 associated with susceptibility of chronic benzene poisoning]. Wei Sheng Yan Jiu. 2007 Jan;36(1):11-5. Chinese.
34. Anthonisen NR, Connett JE, Murray RP. Smoking and lung function of

- Lung Health Study participants after 11 years. *Am J Respir Crit Care Med*. 2002 Sep;166(5):675-9.
35. Beck GJ, Doyle CA, Schachter EN. Smoking and lung function. *Am Rev Respir Dis*. 1981 Feb;123(2):149-55.
36. Kradjan WA, Driesner NK, Abuan TH, Emmick G, Schoene RB. Effect of age on bronchodilator response. *Chest*. 1992 Jun;101(6):1545-51.
37. Ware JH, Dockery DW, Louis TA, Xu XP, Ferris BG Jr, Speizer FE. Longitudinal and cross-sectional estimates of pulmonary function decline in never-smoking adults. *Am J Epidemiol*. 1990 Oct;132(4):685-700.
38. Miller A, Thornton JC, Warshaw R, Bernstein J, Selikoff IJ, Teirstein AS. Mean and instantaneous expiratory flows, FVC and FEV1: prediction equations from a probability sample of Michigan, a large industrial state. *Bull Eur Physiopathol Respir*. 1986 Nov-Dec;22(6):589-97.
39. Meeker JD, Calafat AM, Hauser R. Urinary phthalate metabolites and their biotransformation products: predictors and temporal variability among men and women. *J Expo Sci Environ Epidemiol*. 2012 Jul;22(4):376-85.
40. Harrison's Principles of Internal Medicine, 18th ed. Chapter 10. Disorders

of the Respiratory System 2011: McGraw-Hill.

41. Simon MR, Chinchilli VM, Phillips BR, Sorkness CA, Lemanske RF Jr, Szeffler SJ et al; Childhood Asthma Research and Education Network of the National Heart, Lung, and Blood Institute. Forced expiratory flow between 25% and 75% of vital capacity and FEV1/forced vital capacity ratio in relation to clinical and physiological parameters in asthmatic children with normal FEV1 values. *J Allergy Clin Immunol.* 2010 Sep;126(3):527-34.e1-8.

42. Ciprandi G, Tosca MA, Capasso M. Forced expiratory flow between 25 and 75% of vital capacity might be a predictive factor for bronchial hyperreactivity in children with allergic rhinitis, asthma, or both. *Allergy Asthma Proc.* 2011 Sep-Oct;32(5):e22-8.

국문 초록

서론: 프탈레이트는 흔한 환경 노출과 이에 따른 폐기능 감소 등의 건강 영향에 대해 우려를 일으키는 화학물질이며, 노령 인구는 이러한 영향에 대해 취약한 군이므로 이들 집단에서 프탈레이트 노출에 의한 폐기능 감소 또한 중요한 건강문제로 대두되고 있다. 국내 노령인구에서 프탈레이트 대사물질의 수치와 폐기능 지표 간 관련성을 조사하고, 이러한 관련성에 대한 산화손상 관련 유전자의 (*CAT*, *SOD2*, *MPO*) 가능한 효과 조절을 평가한다.

방법: 서울시에 거주하는 60 세 이상의 노인 418 명에 대상자에 대해 패널 연구를 수행하여, 요중 프탈레이트 대사물질 측정과 폐기능 검사를 반복하였고, *CAT* (rs769218, rs769217), *SOD2* (rs4880, rs2758331, rs5746136), *MPO* (rs2071409, rs7208693) 유전자의 유전자 다형성을 조사하였다. 혼합효과모형을 적용하여 프탈레이트 대사물질 수치와 폐기능 지표 간 관련성 및 이러한 관련성에 대한 *CAT*, *SOD2*, *MPO* 다형성의 영향을 분석하였다.

결과: Mono-(2-ethyl-5-hydroxyhexyl) phthalate, mono-(2-

ethyl-5-oxohexyl) phthalate를 합한 수치와 (Σ DEHP) FEV₁/FVC 및 FEF₂₅₋₇₅의 관련성은 연령, 이전 측정 후 지난 개월 수, 성별, 체질량지수, 코티닌 수치, 평균 기온 및 이슬점을 보정한 상태에서 유의한 역의 상관관계를 나타냈다 (P-values: FEV₁/FVC 0.0275, FEF₂₅₋₇₅ 0.002). 프탈레이트 노출의 폐기능에 대한 영향은 *CAT* rs769218 중 GG 유전형, *CAT* rs769217의 CC 유전형, *SOD2* rs4880의 TT 유전형, *SOD2* rs2758331의 CC 유전형, *MPO* rs2071409의 AA 유전형, *MPO* rs7208693의 GG 유전형에서만 유의하게 나타났다. 세 가지 유전자에서 세 개의 유의한 이배체를 모두가 가진 경우 그렇지 않은 경우에 비해 프탈레이트-폐기능 관련성에 있어서 차이를 나타내었다 (P-values for interaction: Σ DEHP-FEV₁/FVC 0.0176, Σ DEHP-FEF₂₅₋₇₅ 0.0663)

결론: 국내 노령인구에서 요중 프탈레이트 대사물질 수치의 증가와 폐기능의 감소는 유의한 관련성을 나타내었으며, 이러한 관련성에 대한 몇몇 *CAT*, *SOD2*, *MPO* 유전자 다형성의 취약성 영향을 확인할 수 있었다. 폐기능의 감소는 유의한 관련성을 나타내었으며, 이러한 관련성에 대한 몇몇 *CAT*, *SOD2*, *MPO* 유전자 다형성의 취약성 영향을 확인할 수 있었다.

주요어 : 프탈레이트, 폐기능, 유전자 다형성, 노령인구

학 번 : 2010-23717