

The effect of periodontal flap surgery on Matrix metalloproteinases (MMPs) and Tissue inhibitors of matrix metalloproteinase-1 (TIMP-1) levels in gingival crevicular fluids of periodontitis patients

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I. Introduction

As we all know, periodontal disease is characterized by inflammation that can lead to periodontal attachment loss and bone destruction and is largely initiated by bacterial plaque. It is now recognized that during active periodontitis, degradation of the collagenous matrices which support the skeletal framework of gingival complex is due in part to matrix metalloproteinase (MMPs) expressed *in situ* by inflammatory cells(monocytes, macrophages, lymphocytes, polymorphonuclear cells) and resident cells(fibroblasts, epithelial cells, and endothelial cells), and they are thought to be partly responsible for other kinds of diseases as well. MMPs belong to the matrixin family, which is composed of at least 23 related zinc- dependent endopeptidases that are

able to degrade extracellular matrix proteins at a pH close to neutral. And they can be divided into several major subgroups, such as the interstitial collagenases (MMP-1, -8 and -13), the gelatinases (MMP-2 and -9; also called type IV collagenase), the stromelysins(MMP-3, -10 and -11) and the membrane-bound group(MMP-14, -15, -16 and -17). Among them, several MMPs (like MMP-1, 2, 8, 9 and 13) as well as tissue inhibitors of matrix metalloproteinase-1(TIMP-1) are thought to be more strongly related to overwhelming periodontal disease, such as adult periodontitis (AP) and localized juvenile periodontitis(LJP) by previous classification.

The purpose of this study is to find out whether specific MMPs(-1, -8, -9, -13) or TIMP -1 are present in peri- odontitis patients and to determine if there are any significant differences between the amount

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of MMPs before periodontal surgery and after periodontal surgery. We got MMPs and TIMP-1 in gingival crevicular fluid (GCF) because GCF offers a unique opportunity to sample analytical quantities of interstitial fluid by minimally intrusive and non-surgical means, and we analyzed them by using ELISA.

We also tried to determine if those assays have some positive relationships with probing depth and GI scores (Löe and Silness), which represent clinical periodontal status easily.

II. Materials and Methods

1. Study population

Sixteen individuals, newly referred to Seoul National University Dental Hospital (SNUDH), Department of Periodontology with chronic periodontitis took part in this study; 2 subjects were dropped from the study because they did not attend recall examinations. The remaining 14 patients (5 women; 9 men) had a mean age of 42.9 years (range 25 to 58). All patients had moderate to advanced periodontitis, and the experimental tooth site for each patient had pocket depth more than 6mm, whereas the healthy control sites were selected which had a pocket depth under 3mm in the same patient.

All subjects were in good general health; no participants had a history of systemic conditions such as a heart disease, diabetes, uncontrolled hypertension, significant risk for infectious disease transmission, renal or liver disease, and other types of disorders which could influence the course of periodontal disease. They were also not on any medication that could affect the manifestations of periodontal disease, such as chronic antibiotic use, phenytoin, cyclosporine, anti-inflammatory drugs, or calcium

channel blockers. None of the women was postmenopausal.

2. Clinical measurements

The clinical evaluation of the experimental sites was based on the following indices: gingival index (GI) of Löe and Silness and probing depths (PD). Probing depths were measured with a Marquis probe calibrated in millimeters.

GI score and PD were measured before (Pre) and after (Post) periodontal flap surgery in experimental sites. All post-surgery measurements were obtained 6 weeks after periodontal flap operation.

3. Gingival biopsy

Gingival tissues were obtained from experimental sites in each participant during periodontal flap surgery, and immediately fixed in 10% formalin solution after excision.

Cells and tissue were stained with hematoxylin and eosin, and this staining technique revealed the structural integrity of the epithelia and the absence or presence of inflammatory infiltrate in the gingiva.

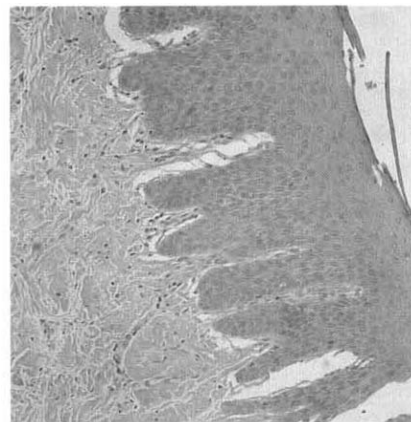


Figure 1. Gingival inflammation classified as mild, according to inflammatory cell infiltration.

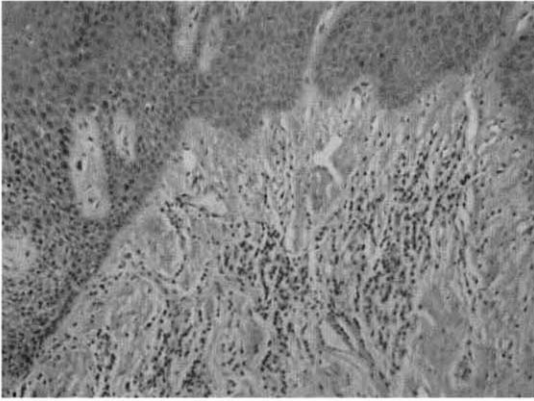


Figure 2. moderate inflammation

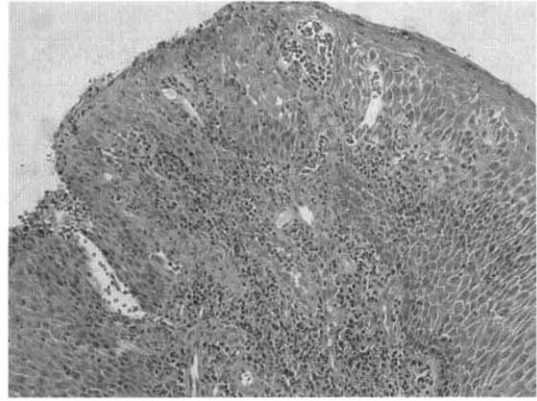


Figure 3. severe inflammation

They were assessed with severity of inflammation under electron microscopy, and compared them with GI scores to find out whether this clinical index is reliable. The severity of gingival inflammation was classified(B) as *normal, mild, moderate, and severe*, according to the degree of inflammatory cell infiltration, tissue destruction, vascular changes and scarring(Figure 1, 2, 3).

4. Gingival crevicular fluid (GCF) sampling and processing

GCF samples were collected using sterile paper strip(PROFLOW™, New York), and the area was carefully isolated and gently dried by an air syringe to prevent samples from being contaminated by saliva. The paper strips were inserted into the crevice until mild resistance was felt or in any event not more than 1mm, and left in place for 30 seconds. And they were immediately placed into a coded sterile microtube and stored at -70°C until analysis.

GCF samples were collected from control sites(C), pre-surgical experimental sites (Pre) and post-surgical experimental sites(Post), and paper strips were used for four times respectively in every site to obtain suf-

ficient amount of MMPs or TIMP from GCF.

For the quantification of MMPs(-1, -8, -9, -13) and TIMP-1, Human Biotrak ELISA kit(amersham pharmacia biotech, UK) were used.

5. Statistical analysis

Wilcoxon signed rank test was employed to evaluate if there were any statistical differences in levels of MMPs (-1, -8, -9, -13) or TIMP-1 between control, pre-surgical, and post-surgical groups, as the sample size was not enough to do ANOVA. The criterion for statistical significance was defined as a level of $P < 0.05$.

The correlations among clinical parameters(PD, GI) were analyzed using Spearman's correlation test. For the statistical analysis between clinical parameters and MMPs or TIMP-1 levels, Spearman's correlation coefficient(r) was also employed, and a P value < 0.05 was considered statistically significant.

III. Results

1. Level changes of MMPs and TIMP-1 after surgery

The results showed that all the MMPs(MMP-1, 8,

Table 1. Patients' whole data except MMPs and TIMP levels

pt	sex	C	E	PD(pre)	GI(pre)	PD(post)	GI(post)	B
1	M	#11(M)	#45(DL)	6	2	3	1	mild
2	M	#33(M)	#36(L)	6	1	3	0	mild
3	M	#33(DB)	#34(DB)	6	2	3	1	moderate
4	M	#14(MB)	#16(MB)	7	2	4	1	severe
5	M	#14(MB)	#32(DB)	7	3	3	1	severe
6	F	#17(MB)	#12(L)	7	3	4	1	severe
7	F	#27(BF)	#23(MB)	7	2	4	0	severe
8	M	#36(B)	#26(BF)	9	3	4	1	severe
9	F	#36(BF)	#33(MB)	6	2	3	0	moderate
10	M	#16(MP)	#15(MP)	6	2	2	0	moderate
11	M	#43(DB)	#46(DB)	6	2	3	0	moderate
12	F	#24(DP)	#26(P)	6	2	3	0	moderate
13	M	#23(DB)	#26(P)	8	3	4	1	severe
14	F	#23(M)	#36(BF)	8	3	3	0	severe

(C: Control site, E: Experimental site, pre: pre-operative data, post: post-operative data, B: severity of inflammation from biopsy)

Table 2. mean levels of MMPs and TIMP in each group (ng/ml)

group	MMP-1			MMP-8			MMP-9			MMP-13			TIMP-1		
	cont	pre	post	cont	pre	post	cont	pre	post	cont	pre	post	cont	pre	post
mean	0,04	0,08	0,02	4,13	5,85	4,20	20,20	54,17	27,30	0,05	0,19	0,04	3,77	3,64	3,30
SD	0,05	0,13	0,03	2,25	3,50	2,34	15,15	46,14	43,06	0,05	0,17	0,06	3,53	5,40	4,77

(cont: control site, pre: pre-operative data, post: post-operative data)

Table 3. p-values by Wilcoxon Signed Rank Test between control (C), pre-operative (Pre) and post-operative (Post) in MMPs and TIMP-1 (p <0.05)

	MMP-1	MMP-8	MMP-9	MMP-13	TIMP-1
C vs. Pre	0,114	0,177	0,011	0,026	0,875
C vs. Post	0,212	0,875	0,638	0,861	0,363
Pre vs. Post	0,025	0,158	0,016	0,009	0,551

9, 13) levels in GCF were markedly decreased after periodontal flap surgery. But we found statistically significant difference in MMP-1(p=0,025), MMP-9(p=0,016) and MMP-13(p=0,009) levels between pre-surgical(Pre) and post- surgical(Post) evaluation(Table 2, 3, Figure 4-8). Absolutely elevated level of MMPs was found in diseased experimental sites compared to healthy control sites, but we could find statistically significant difference only in MMP-9(p=0,011) and MMP-13(p=0,026). Level of MMP-9

showed greatest amount among all other MMPs and TIMP levels, and MMP-9 level was even 500 to 1500 fold greater than MMP-1 level.

TIMP-1 levels were decreased after periodontal flap surgery, and showed highest levels in healthy control sites even though not statistically significant.

2. Relationships between clinical parameters

After surgical treatment, there was absolute

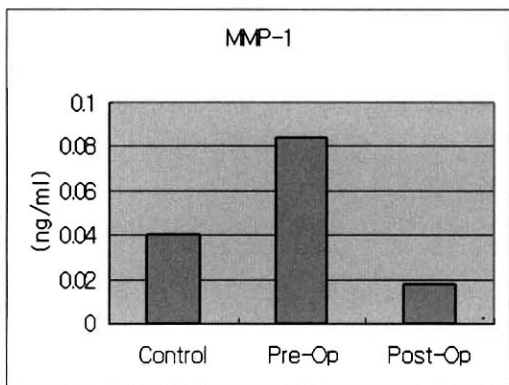


Figure 4, mean levels of MMP-1

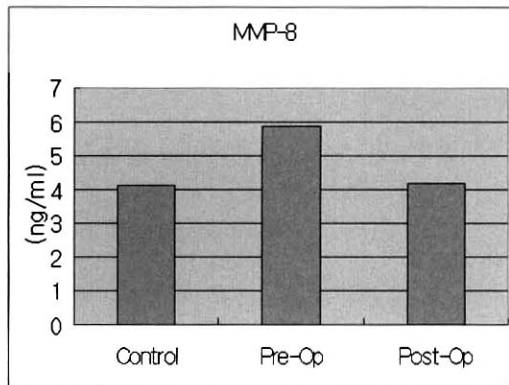


Figure 5, mean levels of MMP

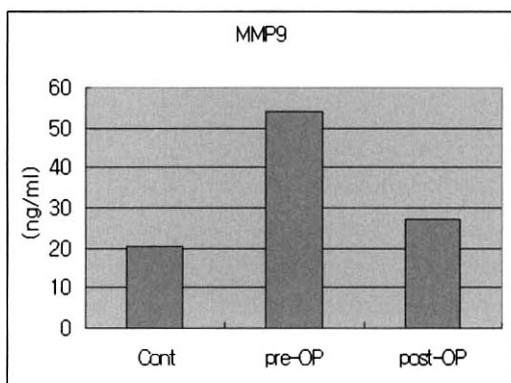


Figure 6, mean levels of MMP-9

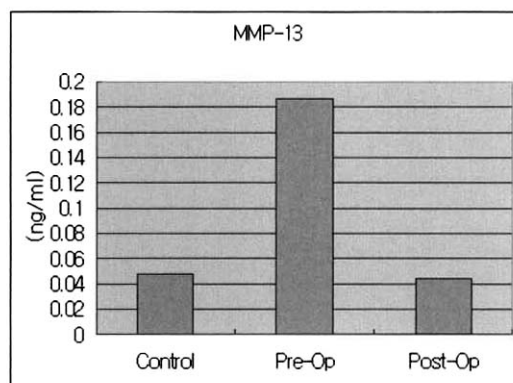


Figure 7, mean levels of MMP-13

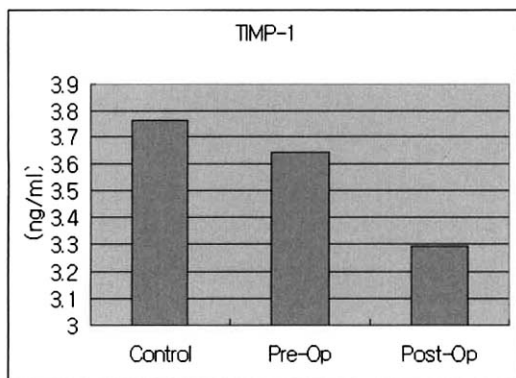


Figure 8, mean levels of TIMP-1

decrease in pocket depth(PD) and gingival index(GI). But when we took them into considerations with statistical view, only pocket depths

showed significant difference($p=0.005$). Maybe it can be easily expected, there was strong relationship between pocket depth and gingival index at baseline, and they were also statistically significant($p=0.000$, $r=0.808$).

With the biopsy procedure, the severity of gingival inflammation was classified as *normal*, *mild*, *moderate*, and *severe*, according to the degree of inflammatory cell infiltration, and compared the results with gingival index(GI) whether the index is clinically reliable. Fortunately, we could find that there were strong positive relationships between them and surely they were statistically significant ($p=0.001$, $r=0.773$) even though study pool was small.

Table 4. Spearman' s coefficient (r) of MMPs and TIMP-1 with PD (p <0.05)

	MMP-1	MMP-8	MMP-9	MMP-13	TIMP-1
r	0,000	-0,092	0,485	0,248	-0,433
p-value	1,000	0,968	0,072	0,394	0,122

Table 5. Spearman' s coefficient (r) of MMPs and TIMP-1 with GI (p <0.05)

	MMP-1	MMP-8	MMP-9	MMP-13	TIMP-1
r	-0,074	-0,243	0,498	-0,440	-0,268
p-value	0,801	0,403	0,070	0,115	0,355

When we compared the MMPs and TIMP-1 levels with pre-operative pocket depth and gingival index, all the data showed no statistically significant difference, but only MMP-9 showed some positive relationship with both baseline pocket depth and baseline gingival index(Table 4, 5).

IV. Discussion

As mentioned above, it is well known that extracellular matrix degradation in periodontal disease is mainly due to the increased level of host and microbial-derived proteinases such as MMPs, which are secreted during the interaction of plaque bacteria with resident gingival cells(fibroblast, epithelial cells) or inflammatory cells such as PMN or monocyte/macrophage(Birkedal-Hansen et al 1993, Sorsa et al 1992). They usually secreted in latent proforms and have a pro-peptide containing a conserved cysteine in a region of homology. This cysteine is bonded to the Zn²⁺ of the active site in the latent form, and it is thought that dissociation of the cysteine thiol leads to exposure of the active site. Then the specific attack on matrix macromolecules take place, and this initial attack on the integrity of extracellular macromolecules facilitates further extracellular degradation.

There were many evidences that withstand the

fact that MMPs have intimate relationship with periodontal disease progression.

It was reported that elevated interstitial collagenase(MMP-1, MMP-8) activities have been detected in GCF of adult periodontitis(AP) and localized juvenile periodontitis(LJP), and these activities have been shown to decrease following periodontal treatment in both groups (Hakkarainen et al 1988). Ingman(1996) also reported that MMP-8 and MMP-9 are the main collagenases in gingival tissue and GCF of AP patients, whereas the predominant collagenase in GCF of LJP seems to be of the MMP-1 type. He emphasized the importance of doxycycline, a known inhibitor of MMP-8 and MMP-9, as a possible adjunctive drug in the treatment of AP. However, the presence of MMP-1 in LJP GCF, whose MMP-type has been shown to be rather resistant to doxycycline inhibition at therapeutic levels, may suggest that doxycycline may be a less useful drug in the control of periodontal tissue destruction in LJP than in AP.

On the other hand, Anne-Laure et al,(2003) revealed that MMP-9, MMP-13 and TIMP-1 increased significantly in the culture media using blot technique with severe inflammation. Furthermore, their results seemed to be correlated with those reported by Makela et al.(1994), who demonstrated that MMP-9 was the major gelatinase present in the gin-

gival crevicular fluid of periodontal patients.

In this study, we got results that have something in common with former reports in that all the MMPs from the inflamed experimental sites showed much elevated level compared to those from the healthy control sites, and PMN-type gelatinase (MMP-9) showed much higher level among other MMPs in GCF, from which we can conclude that MMP-9 is predominant matrix metalloproteinase in periodontitis patients.

After the flap surgery, all the MMPs (MMP-1,-8,-9,-13) levels in GCF were markedly decreased. But we found statistically significant difference only in MMP-1, MMP-9 and MMP-13 levels between pre-surgical (Pre) and post-surgical (Post) evaluation. There were several reports which help us to understand this result. After establishment of a supragingivally clean oral environment, a rapid decrease of the collagenase activity took place following scaling and root planing of the root surfaces within the periodontal pockets. Also, occlusal adjustment of the hypermobile teeth with deep pathological pockets reduced the protein content and collagenase activity in sulcular fluid (Hakkarainen et al 1988). And Glay Tüter (2002) reported that levels of MMP-1 in GCF decreased and total levels of TIMP-1 in GCF increased after phase-I periodontal therapy. Those findings strongly suggest that such pathogen-induced MMPs can be well regulated by our conventional periodontal treatment.

For the regulation of destructive cascade by MMPs, there exists glycoprotein which called tissue inhibitor of matrix metalloproteinase (TIMP) that is synthesized and secreted by most connective tissue cells as well as by macrophages. TIMPs share a common two-domain structure in which only the inhibitory N-domain is capable of inhibiting MMPs. The amino group of the N-terminal cysteine of TIMP co-ordinates with the active site Zn^{2+} of MMPs with

the adjacent residues of TIMP occupying the active site cleft of MMPs and contacting the surrounding surface of the catalytic domain of MMPs. They inactivate MMPs in that manner. But even though we clearly know the exact mechanism of how they work, whether TIMP level is higher or not in inflamed site than in healthy site is still controversial. Nomura (1993) reported that TIMP was higher in inflamed sites than in healthy sites. He explained this result that when there is stimulation by bacterial colonization, MMP expression by host cell increases, and subsequent self tissue destruction occurs. So the host cells recognize the ongoing tissue destruction and try to defend the host tissue by producing TIMPs. Similarly, with elevated level of PMN elastase in inflamed sites, concomitant elevated level of 1-proteinase inhibitor which regulates PMN elastase by forming an irreversible enzyme-inhibitor complex was detected in the GCF of periodontitis patients compared to controls (Ingman 1994).

On the contrary, Larivee et al, (1986) said that the activity of TIMP is relatively higher rather in healthy sites than in diseased sites. Moreover, Sorsa (1994) found that GCF of adult periodontitis patients contain PMN-derived MMP-8 and -9 but not recognizable amounts of TIMP-1. Thus it seems that PMNs do not contain and release TIMP-1 in amounts comparable to MMP-8 or -9. Glay Tüter (2002) found levels of TIMP-1 increase significantly after phase-I therapy compared to baseline, and explained this phenomenon as a reduction of MMP-1, which bind to free TIMP (however, the regulation of TIMP-1 may not be solely dependent on the MMP-1). Moreover, the decreased levels of TIMP-1 in periodontally diseased subjects may be due to the selective degradation of TIMP-1 by neutrophil elastase or the inactivation of TIMP-1 by neutrophils themselves following oxidant release. Phase I therapy may have reduced the number of neutrophils and the elastase

released due to resolved gingival inflammation and tissue healing. Haerian et al.(1996) also reported that the GCF levels of stromelysin and TIMP were reduced by periodontal treatment. Our results are consistent with these previous studies in some extent that TIMP-1 levels were decreased after periodontal flap surgery, and showed highest levels in healthy control sites even though not statistically significant.

If we take a look at clinical parameters, though only pocket depths(PD) showed significant difference($p=0.005$), there was absolute decrease in PD and GI after surgical treatment as expected. And strong correlations between PD and GI ($p=0.000$, $r=0.808$) was also found. Moreover, we could conclude that gingival index (GI) is pretty much reliable, when we compared them with the gingival biopsy specimens using electron microscopy. But there were no special correlations between MMPs or TIMP-1 levels and clinical parameters in the present study (only MMP-9 showed weak correlations). Haerian(1996) found a moderate positive and significant correlation between biochemical(GCF levels of stromelysin and TIMP) and clinical parameters when data from healthy, gingivitis and periodontitis sites were pooled. But Viella et al.(1987) reported a positive but rather weak correlation between PD and GI and collagenase activity while Gangbar et al.(1990) and Teng et al.(1992) failed to find any correlations between them. It seems that different GCF sampling methods and laboratory techniques as well as variations in reporting the results including different statistical analysis methods may influence the presence and extent of correlation between clinical and biochemical parameters.

In conclusion, when we think of the level changes of MMPs and TIMP-1 after periodontal flap surgery as well as the other results like correlation between clinical parameters or the actual high

amount we got, it is strongly suggested that MMP-9 and MMP-13 can be a possible marker in periodontal disease and controlled by conventional periodontal flap surgery. Other MMPs or TIMP-1 need to be examined for more information by further longitudinal study with more sample sizes.

V. References

1. Uitto VJ, Airola K, Vaalamo M, et al. Collagenase-3(MMP-13) expression is induced in oral mucosal epithelium during chronic inflammation. *Am J Pathol* 1998;152:1489-1499.
2. Segulier S, Godeau G, Brousse N. Collagen fibers and inflammatory cells in healthy and diseased human gingival tissues: A comparative and quantitative study by immunohistochemistry and automated image analysis. *J Periodontol* 2000;71:1079-1085.
3. Makela M, Salo T, Uitto VJ, Larjava H. Matrix metalloproteinases(MMP-2 and MMP-9) of the oral cavity: Cellular origin and relationship to periodontal status. *J Dent Res* 1994;73:1397-1406.
4. Golub LM, Sorsa T, Lee H-M, et al. Doxycycline inhibits neutrophil (PMN)- type matrix metalloproteinases in human adult periodontitis gingiva. *J Clin Periodontol* 1995;22:100-109.
5. Anne-Laure Ejeil, Sylvie Igondjo-Tchen, Sabah Ghomrasseni, Bernard Pellat, Gaston Godeau, and Bruno Gogly. Expression of MMPs and tissue inhibitors of matrix metalloproteinases (TIMPs) in healthy and diseased human gingiva. *J Periodontol* 2003;73:188-195.
6. Birkedal-Hansen, H. Role of matrix metalloproteinases in human periodontal diseases. *J Periodontol* 1993;64: 474-484.
7. Vilella B, Cogen RB, Bartolucci AA, Birkedal-Hansen, H. Collagenolytic activity in crevicular

- fluid from patients with chronic adult periodontitis, localized juvenile periodontitis and gingivitis, and from healthy control subjects. *J Periodont Res* 1987;22:381-389.
8. Glay Tüter, Blent Kurtis, Muhittin Serdar. Effects of Phase I Periodontal treatment on gingival crevicular fluid levels of matrix metalloproteinase-1 and tissue inhibitor of metalloproteinase-1. *J Periodontol* 2002;73:487-493.
 9. Hakkarainen K, Uitto V-J, Ainamo J. Collagenase activity and protein content of sulcular fluid after scaling and occlusal adjustment of teeth with deep periodontal pockets. *J Periodont Res* 1988;23:204-210.
 10. Denis E Kinane. Regulators of tissue destruction and homeostasis as diagnostic aids in periodontology. *Periodontology* 2000, vol24, 2000215-225.
 11. John J. Reynolds, Murray C. Meikle. Mechanisms of connective tissue matrix destruction in periodontitis. *Periodontology* 2000, vol 14, 1997; 144-157.
 12. Ingman T, Sorsa T, Suomalainen K et al. Tetracycline inhibition and the cellular source of collagenase in gingival crevicular fluid in different periodontal diseases. A review article. *J Periodontol* 1993;64:82-88
 13. Ingman T, Sorsa T, Kangaspunta P et al. Elastase and 1-proteinase inhibitor in gingival crevicular fluid and gingival tissue in adult and juvenile periodontitis. *J Periodontol* 1994;65:702-709.
 14. Ingman T, Tervahartiala T, Ding Y et al. Matrix metalloproteinases and their inhibitors in gingival crevicular fluid and saliva of periodontitis patients. *J Clin Periodontol* 1996;23: 1127-1132.
 15. Uitto VJ, Overall CM, McCulloch C. Proteolytic host cell enzymes in gingival crevice fluid. *Periodontology* 2000, vol 31, 2003;77-104.
 16. Sorsa T, Uitto VJ, Suomalainen K et al. Comparison of interstitial collagenases from human gingiva, sulcular fluid and polymorphonuclear leukocytes. *J Periodont Res* 1988; 23:386-393.
 17. Meikle MC, Hembry RM, Holley J et al. Immunolocalization of matrix metalloproteinases and TIMP-1(tissue inhibitors of metalloproteinases) in human gingival tissues from periodontitis patients. *J Periodont Res* 1994;29:118-126.
 18. Teng YT, Sodek J, McCulloch C. Gingival crevicular fluid gelatinase and its relationship to periodontal disease in human subjects. *J Periodont Res* 1992;27:544-552.
 19. Nomura T, Takahashi T, Hara K. Expression of TIMP-1, TIMP-2 and collagenase mRNA in periodontitis affected human gingival tissue. *J Periodont Res* 1993;28:354-360.
 20. Haerian A, Adonogianaki E, Mooney J et al. Gingival crevicular stromelysin, collagenase and tissue inhibitor of metalloproteinases level in healthy and diseased sites. *J Clin Periodontol* 1995;22:505-509.
 21. Larvee et al. Collagenase and collagenase inhibitor activities in crevicular fluid of patients receiving treatment for localized juvenile periodontitis. *J Periodont Res* 1986; 21:701-715.
 22. Haerian A, Adonogianaki E, Mooney J, Manos A, Kinane DF. Effects of treatment on gingival crevicular collagenase, stromelysin and tissue inhibitor of metalloproteinases and their ability to predict response to treatment. *J Clin Periodontol* 1996; 23:83-91.
 23. Gangbar S, Overall CM, McCulloch CAG, Sodek J. Identification of polymorphonuclear leukocyte collagenase and gelatinase activities in mouthrinse samples: Correlation with periodontal disease activity in adult and juvenile periodontitis. *J Periodont Res* 1990; 25:257-267.

치주 수술이 치주염 환자의 치은 열구액 내의 MMPs와 TIMP-1에 미치는 영향

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중등도 이상의 치주염 환자에서 치은 열구액내의 MMPs 및 TIMP-1과 치주염과의 연관성을 규명하고, 치주 수술이 MMPs 및 TIMP-1의 정량에 미치는 영향을 연구하고자 하였다.

총 14명의 치주낭 깊이 6mm 이상의 중등도 이상의 치주 질환 이환자에서 치아를 선정하여, 치주낭 심도, 치은지수(gingival index)를 측정하고, 치은의 조직학적 염증의 정도를 측정하기 위해, 해당 치아의 치주낭 연조직을 절취하여 H-E염색을 하고, 치은 절편에서 염증세포 침윤의 정도 및 분포를 비교하였다. Perio-paper를 이용하여 치은열구액을 얻고, pyrogen-free water에서 추출하였다. 채취한 치은 열구액에서 ELISA-kit를 이용하여 MMP-1, 8, 9, 13과 TIMP-1을 측정하여 수술 전과, 수술 후, 그리고 건강한 조직인 대조군을 비교하였으며, 통계 처리는 Wilcoxon 검정을 사용하였다. 또한 MMPs 혹은 TIMP-1이 치주낭 심도나 치은지수등의 임상적 지표와 가지는 연관성을 Spearman's correlation coefficient를 이용하여 알아보았다.

TIMP-1을 제외한 MMP-1, 8, 9, 13 에서 수술 전보다 수술 후에 치은 열구액 내의 양이 현저하게 줄어든 것을 관찰할 수 있었으나, MMP-1($p=0.025$), MMP-9($p=0.016$) 와 MMP-13($p=0.009$) 에서만 통계적으로 유의성있는 차이를 보였다. 한편 MMP-9 ($p=0.011$) 나 MMP-13($p=0.026$) 은 건강한 대조군과 수술 전 사이에도 유의성있는 차이를 보였다. 연조직의 조직학적 관찰을 통하여 치은지수의 임상적 신뢰도를 평가한 결과 통계학적으로 유의한 결과를 얻을 수 있었으며, 치주 치료 전의 치주낭 심도와 치은지수와의 관계나, 수술 전과 수술 후의 치주낭 심도등의 변화도 통계적으로 유의성있는 결과를 보였다. 하지만 치주낭 심도나 치은지수등의 임상적 지표는 MMPs나 TIMP의 정량과는 별다른 연관성을 보이지 않았다.

이 실험의 결과로 보아 MMP-1, MMP-9 나 MMP-13을 치주 수술 전과 수술 후의 치주염의 심도 변화를 반영할 수 있는 지표로 생각할 수 있으며, 특히 MMP-9와 MMP-13가 치주염과 가지는 연관성은 크다고 할 수 있겠다.

주요어: matrix metalloproteinase(MMP), tissue inhibitor of matrix metalloproteinase (TIMP), 치주낭 심도, 치은지수, 치주 수술, 치은 열구액