

**Membrane proteins involved in epithelial-mesenchymal transition and tumor invasion;  
studies on TMPRSS4 and TM4SF5**

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Running title: EMT by TMPRSS4 or TM4SF5

## **1. Introduction**

Epithelial-mesenchymal transition (EMT) is one mechanism by which cells with mesenchymal features can be generated and is a fundamental event in morphogenesis. Recently, invasion and metastasis of cancer cells from the primary tumor are now thought to be initiated by the developmental process termed EMT, whereby epithelial cells lose cell polarity and cell-cell interactions, and gain mesenchymal phenotypes with increased migratory and invasive properties. EMT is believed to be an important step in metastasis and implicated in cancer progression although the influence of EMT in clinical specimens has been debated. This review presents the recent results of two cell surface proteins, of which functions and their underlying mechanisms recently began to be demonstrated, as novel regulators of the molecular networks that induce EMT and cancer progression.

Key words: EMT; invasion; membrane protein; TM4SF5; TMPRSS4.

## **2. Epithelial-mesenchymal transition (EMT)**

Metastasis is the leading cause of cancer-related deaths in most cancer types. As an initial step in cancer metastasis, epithelial tumor cells in general disseminate from primary solid tumor mass and invade into the surrounding stromal tissues. Invasion is enhanced by tumor cell activation of EMT [1-4]. EMT is characterized by the loss of epithelial apicobasal polarity and cell-cell contacts, modulation of cell-matrix adhesion, enhanced proteolytic activity, cytoskeletal remodeling, and acquisition of the ability to migrate and invade extracellular matrix (ECM) [1, 3]. During EMT, epithelial cells undergo molecular changes; epithelial cells gradually lose their epithelial markers such as E-cadherin, ZO-1, and cytokeratins, and concomitantly acquiring mesenchymal markers such as vimentin, fibronectin, N-cadherin, and alpha smooth muscle actin [1, 3]. EMT plays a critical role in the formation of various tissues and organs such as the mesoderm, neural crest, heart, secondary palate, and peripheral nervous systems during embryonic development and wound healing in adult organism [2, 4]. Furthermore, EMT is implicated in pathological processes, such as tumor cell invasion and metastasis and organ fibrosis [2].

One of the hallmarks of EMT is the functional loss of E-cadherin, which is currently thought to be a metastasis suppressor [5]. Downregulation of E-cadherin is usually mediated by E-cadherin transcriptional repressors/EMT-inducing transcription factors, including the Snail superfamily of zinc-finger factors (Snail and Slug), the ZEB family (ZEB1 and ZEB2) and basic helix-loop-helix factors (Twist1 and E47), which have been associated with tumor

invasion and metastasis [5, 4]. These factors repress transcription of E-cadherin by interacting with proximal E-box elements in the E-cadherin promoter [5]. In addition, these E-cadherin repressors may be directly or indirectly involved in the upregulation of certain mesenchymal genes expression [5], although the precise mechanism of these regulations is largely unknown.

EMT is triggered by soluble growth factors, such as members of transforming growth factor- $\beta$  (TGF $\beta$ ) and fibroblast growth factor (FGF) families, epidermal growth factor (EGF) and hepatocyte growth factor (HGF) [3, 4]. Subsequent activation of receptor-mediated signaling triggers the activation of the intracellular effector molecules, such as members of the small GTPase family, leading to the changes in cytoskeletal organization, and also results in the activation of EMT-inducing transcription factors [3, 4]. In addition, components of the ECM, such as collagen, and activation of integrin co-receptors are also involved in EMT process [3]. Certain proteases are sufficient to induce EMT [2]; for example, MMP3 triggers EMT by increasing the cellular levels of reactive oxygen species, which in turn induces Snail expression [6].

Recently, microRNAs (miRs) have been identified as a novel class of EMT regulators; miRs to negatively regulate EMT include miR 153, 155, 194, 25, 212, and 200 family, and to positively regulate include miR 29a, 103/107, 150, and 221/22 [7]. miRs regulate invasion and metastasis by targeting the transcripts of various genes involved in EMT event, including EMT-inducing transcription factors. For example, members of the miR-200 family are negative regulators of EMT and essential for the maintenance of the epithelial status through the downregulation of ZEB1 and ZEB2. In turn, miR-200 members are transcriptionally repressed by ZEB1 and ZEB2 thus establishing a double-negative feedback loop [8].

EMT is recently shown to be linked to stemness, self-renewal capacity [9, 10]. In cases

of breast cancer stems, the linkage among EMT phenotype, stemness, and drug resistance has been well-studied [11]. Further, Epithelial-Mesenchymal Plasticity (EMP consisting of EMT and MET) is also described in circulating tumor cells (CTCs) [12-14]. CTCs with various degrees of EMT phenotypes are found during the breast cancer metastasis [15]. Therefore, CTCs may involve self-renewal capacity, which is linked to EMT, during cancer metastasis [16].

### **3. Transmembrane protease, serine 4 (TMPRSS4)**

#### **3.1. Introduction to TTSPs**

Dysregulation of proteases is a hallmark of cancer progression, thus proteases in general have been the subject of numerous cancer studies. Extracellular proteolytic enzymes, including matrix metalloproteinases (MMPs) and serine proteases, contribute to tumor cell invasion and metastasis through both direct proteolytic activity and the regulation of cellular signaling and functions [17-19]. Most members of the serine protease family are either secreted or sequestered in cytoplasmic organelles awaiting signal-regulated release. Recently, type II transmembrane serine proteases (TTSPs) have been recognized as a new subfamily of serine proteases that have in common an extracellular proteolytic domain, a single-pass transmembrane domain, a short intracellular domain and a variable-length stem region containing modular structural domains [20-24]. Enteropeptidase (also known as enterokinase) that has been identified over a century ago due to its pivotal role in food digestion is the first TTSP, which was revealed by the molecular cloning of the enteropeptidase cDNA two decades ago [25]. TMPRSS2, human airway trypsin-like protease (HAT), corin, and matriptase have been subsequently identified as cell surface-associated proteases [23, 24]. To

date, 20 TTSPs have been identified in mouse and humans due to the analysis of sequence data from the mammalian genome projects [23]. Analysis of the tissue distribution of the TTSPs and gene targeting in mice of certain TTSPs suggested that a significant number of TTSPs may have important functions in embryonic development and homeostasis of mammalian tissues such as heart, skin, inner ear, placenta, and digestive tract [23, 24].

Most TTSPs are overexpressed in a variety of tumors compared to normal tissues, implicating their potential as novel markers of tumor development and progression and possible molecular targets for anti-cancer therapeutics [26, 23]. Recently, a number of works have focused on the evaluation of the expression of individual TTSPs during tumor progression and on the investigation of the potential roles of these proteases in tumor cell proliferation, migration and invasion [27, 23].

### **3.2. TMPRSS4 in cancer**

TMPRSS4 (Gene ID, 56649; Chromosomal location, 11q23.3), initially referred to as TMPRSS3, was originally identified as a gene expressed in most pancreatic cancer tissues but not in the normal pancreas or chronic pancreatitis [28]. To date, 7 isoforms have been reported. The deduced sequence of 437 amino acids of the longest isoform (isoform 1) contains a serine protease domain with putative trypsin-like activity and a transmembrane domain [28]. In human, TMPRSS4 mRNA was detected in bladder, esophagus, stomach, small intestine, colon and kidney [28] although the physiological roles of TMPRSS4 remain unknown. Furthermore, TMPRSS4 expression was upregulated in malignant compared to benign thyroid neoplasm and was suggested as both a diagnostic and prognostic marker [29, 30]. TMPRSS4 was associated with poor prognosis in non-small cell lung cancer (NSCLC) with squamous cell histology [31], triple-negative breast cancer [32], cervical cancer [33],

and gastric cancer patients [34]. Kim *et al.* reported that TMPRSS4 mRNA levels were upregulated in colorectal cancer tissues than in adjacent normal mucosa [35]. The authors also reported that TMPRSS4 protein expression was significantly higher in human colorectal cancer tissues from advanced stages (52.5 and 50.0% of stages III and IV, respectively) than in that of early stage (6.3% in stage I), suggesting that TMPRSS4 may play a role in the progression of non-invasive tumors to invasive malignancies [35]. Jia *et al.* showed that the inhibitory tripeptide, tyrosyleutide, led to a downregulation of TMPRSS4 in hepatocellular carcinoma (HCC), thereby reducing the invasion and metastasis of HCC induced by irradiation [36]. Taken together, TMPRSS4 may be a novel biomarker for the prognosis of certain types of cancers and could be employed for diagnostics and therapeutics.

On the other hand, the mechanism by which TMPRSS4 expression is modulated has not been well characterized. Recently, Nguyen *et al.* reported that TMPRSS4 was increased in NSCLC cells under hypoxic conditions, suggesting that hypoxia within the tumor microenvironment may upregulate TMPRSS4 expression [37].

### **3.3. Function of TMPRSS4 in the regulation of EMT and invasion**

In colon cancer cells, TMPRSS4 induced downregulation of E-cadherin and leads to EMT events, accompanying morphological changes and actin reorganization [38]. Suppression of TMPRSS4 by siRNA reduced cell invasion in colon and lung cancer cells, while overexpression TMPRSS4 induces migration, invasion and metastasis [38]. Attachment and spreading of cells on the extracellular matrix, with concomitant formation of stress fibers and focal adhesions, is prerequisite for cell migration. TMPRSS4 also modulates cell-matrix adhesion and cell spreading mainly through modulation of integrins such as  $\alpha 5\beta 1$  that has been centrally implicated in EMT and cell motility [39, 40], which probably contributes to

enhanced motility and invasiveness. One of the molecular mechanisms by which TMPRSS4 mediates EMT and invasiveness in tumor cells is that TMPRSS4 mediates focal adhesion kinase (FAK) signaling pathway activation and extracellular signal-regulated kinase (ERK) activation mainly through integrin  $\alpha 5$  upregulation, leading to EMT and invasiveness. Furthermore, TMPRSS4 overexpression in human colorectal cancer tissues positively correlated with enhanced expression of integrin  $\alpha 5$  and inversely correlated with E-cadherin expression, confirming that TMPRSS4 modulated expression of EMT markers. Recently, Larzabal *et al.* reported that miR-205 is involved in TMPRSS4-induced integrin  $\alpha 5$  expression in NSCLC cells [41]. To further implicate TMPRSS4 in EMT, Cheng *et al.* suggested that interactions between hepatocyte growth factor activator inhibitor (HAI-1) and TMPRSS4 contribute to EMT events including E-cadherin reduction and morphological changes in pancreatic cancer cells [42]. In addition, TMPRSS4-induced E-cadherin reduction and EMT plays a critical role in radiation-induced long-term metastasis of residual hepatocellular carcinoma in nude mice [43].

Interaction of TMPRSS4 and integrin  $\alpha 5$  based on the observation that TMPRSS4 partially interacted with integrin  $\alpha 5$  under certain coimmunoprecipitation conditions in a cell line-dependent manner [35] (S. Kim, unpublished observation) suggests the possibility that TMPRSS4 may modulate or participate in the interaction of integrin and other cell surface proteins (for example, tetraspanin, receptor tyrosine kinases, etc), leading to subsequent signaling transduction activation. In fact, TMPRSS4 can interact with uPAR (CD87) [44] that can induce EMT in hypoxic breast cancer cells [45], although it is not clear whether TMPRSS4 interacts with uPAR directly or via integrin(s).

Loss or reduction of E-cadherin expression is a well-known hallmark of EMT and

correlates positively with tumor cell invasion and metastasis [3]. E-cadherin expression is transcriptionally downregulated by several transcription factors including Snail family members (Snail and Slug) and ZEB family members (ZEB1 and ZEB2) [5]. TMPRSS4 appeared to modulate SIP1/ZEB2 expression based on the observation that SIP1 mRNA was upregulated in TMPRSS4-overexpressing colon cancer cells although induction of SIP1 at the protein level remains to be determined. Therefore, it is possible that SIP1 mediated TMPRSS4-induced EMT events including E-cadherin reduction.

Several studies have shown that suppression of high endogenous E-cadherin expression renders non-invasive cells partially invasive [46], whereas reconstitution of E-cadherin results in a tumor cell reversion from an invasive mesenchymal phenotype to a benign epithelial phenotype [47, 46]. In contrast, other studies have shown that ectopic expression of E-cadherin could not reverse EMT phenotypes induced by the transcription factor Twist1 [10]. On the other hand, downregulation of E-cadherin was required for TMPRSS4-mediated EMT and invasion in colon cancer cells but was not sufficient for induction of these phenotypes [35], suggesting that downregulation of E-cadherin is not the sole contributor to TMPRSS4-mediated phenotypes. In this respect, upregulation of specific mesenchymal marker such as integrin  $\alpha 5$ , besides the downregulation of E-cadherin by TMPRSS4 may be required for full invasiveness during colon cancer progression (Fig. 1).

#### **3.4. Molecular mechanisms and signals regulated by TMPRSS4**

Numerous studies focused on the investigation of *in vivo* substrates of TTSPs. However, few studies have conclusively addressed the *in vivo* molecular targets and function of TTSPs during tumor progression. *In vitro*, several TTSPs including matriptase were shown to activate pro-urokinase plasminogen activator (pro-uPA), pro-macrophage stimulating protein-

1 (MSP-1), and pro-HGF, which are implicated in proliferation, migration and invasion of various cancer cell types [23].

Like most of the members of the TTSP family, TMPRSS4 can activate epithelial sodium channel (ENaC) *in vitro* through its proteolytic activity, possibly regulating the sodium and water flux across high-resistant epithelia [48, 49]. TMPRSS4 induced cancer cell invasion in a manner that is dependent serine proteolytic activity [38], and inhibitory compounds against TMPRSS4 serine protease activity were reported to reduce colon cancer cell invasion [50]. However, it remains unknown which precursor substrates are cleaved by TMPRSS4 to contribute to tumor progression. On the other hand, it has recently been reported that TMPRSS4 induced urokinase-type plasminogen activator (uPA) gene expression through activation of transcription factors AP-1, Sp1 and Sp3 in mainly a JNK-dependent manner in prostate and lung cancer cells, but not in colon cancer cells [44]. uPA is a well-known serine protease involved in invasion and metastasis and correlates with poor prognosis in breast, lung, stomach, bladder, colon, prostate and ovarian cancers [51], and TMPRSS4 expression significantly correlated with uPA expression in human lung and prostate adenocarcinomas [44]. In addition, TMPRSS4-mediated uPA expression contributed to prostate cancer cell invasion [44] (Fig. 1). It is intriguing that TMPRSS4 activated JNK signaling pathways possibly through its association with uPAR, leading to uPA expression. uPAR can induce EMT and stem cell-like properties in breast cancer cells by activating diverse cell signaling pathways, including ERK, PI3K-Akt, and Rac1 [52, 45]. Therefore, the association of TMPRSS4 and uPAR and subsequent cell signaling modulation may be a novel mechanism for the control of invasion and EMT.

The observations that TMPRSS4 modulated cell signaling and subsequently activated

both AP-1 and Sp1/3 transcriptional activities [44], which have been reported to be involved in the transcriptional regulation of EMT and invasion [53], suggest that TMPRSS4 could modulate the expression of various genes, which may be associated with invasion and metastasis.

#### **4. Transmembrane 4 L six family member 5 (TM4SF5)**

##### **4.1 The tetraspanins**

Tetraspanins (TM4SFs) have four transmembrane protein domains with two extracellular loops and one intracellular loop and the N- and C- terminal tails [54]. They are expressed on the cell surface and/or intracellular vesicles and contain 33 members in mammals [55]. Tetraspanins or TM4SFs are suggested to locate at tetraspanin-enriched microdomain (TERM) [56], where they form protein-protein complexes in hemophilic or heterophilic manners with other TM4SFs, integrins, or growth factor receptors [57, 58]. The protein complexes are known to regulate dynamics of the complex components on the cell surface with regards to diffusion, trafficking, retention, and stability, in addition to influence to intracellular signal transductions [59, 60, 56].

##### **4.2 TM4SF5 in cancer**

*TM4SF5* (Gene ID, 9032) gene maps on chromosome 17, at 17p13.3 according to Entrez Gene. In AceView, it covers 11.34 kb, from 4621928 to 4633262 (NCBI 36, March 2006), on the direct strand containing 4 different gt-ag introns. Its transcription produces 2 alternatively spliced mRNAs via alternative polyadenylation sites, which putatively encode 2 different isoforms (197 and 132 amino acids), containing L6 membrane domain (<http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/av.cgi?db=35g&c=Gene&l=TM4SF5>).

TM4SF5 (20,823 Da) is a transmembrane glycoprotein as a family group related to the

tetraspanin family (transmembrane 4 L six family) including TM4SF1 (L6, L6-Ag), TM4SF4 (IL-TIMP), TM4SF518 (L6D), and TM4SF20 [61, 62]. TM4SF5 is highly expressed in diverse types of cancers, including liver, pancreatic, gastric, colon, ACTH (corticotropin)-negative bronchial carcinoid tumors, soft-tissue sarcoma, nonendocrine lung, and papilla vateri carcinoma [63-66]. Similar to tetraspanins (i.e., transmembrane 4 superfamily, TM4SFs), TM4SF5 has four transmembrane domains (TM1 ~ TM4), short cytoplasmic domains at their N- and C-termini, an intracellular loop (ICL) between TM2 and TM4, and two extracellular loops (EC), a smaller extracellular loop (SEL) between TM1 and TM2, and a larger extracellular loop (LEL) between TM3 and TM4 [61, 62]. Recent clinical studies separately report that *TM4SF5* is highly expressed in tumors from deceased breast cancer patients, compared to those from 10-year breast cancer survivors [67], and that postoperative 5 year overall survival of esophageal cancer patients negatively correlates with TM4SF5 expression [68]. These reports suggest that TM4SF5 overexpression correlates with poor prognosis of cancer patients.

### **4.3 TM4SF5-mediated regulation of signaling molecules**

TM4SF5 can appear to form tetraspanin-enriched microdomain (TERM) on cell surface, via formation of large protein-protein complexes with tetraspanins, integrins, and growth factor receptors [69, 61]. Therefore, by virtues of the protein complex formation, overexpressed TM4SF5 in cancer cells can influence or activate diverse intracellular signaling pathways for cell adhesion, proliferation, EMT, migration, and invasion for tumor progression and maintenance.

TM4SF5 is shown to associate with integrins  $\alpha 2$ ,  $\beta 1$  [70, 71],  $\alpha 5$  [72], and EGFR [73, 74], while cell migration [70, 71], angiogenesis [72], drug resistance [74], and fibrosis [73].

With association and retention of integrin  $\alpha 5$  on cell surface, TM4SF5 can activate intracellular signaling for FAK/c-Src activation leading to STAT3 activity for VEGF induction [72]. In addition, TM4SF5 directly interacts with FAK or c-Src to regulate migration [75] and invasive ECM-degradation [76]. In addition, TM4SF5 expression causes AKT activation, which in turn causes phosphorylation of p27<sup>Kip1</sup> Ser10 for its cytosolic translocation, where it can regulate RhoA activity for morphological change and migratory function [74].

#### **4.4 TM4SF5-mediated EMT in tumor progression**

TM4SF5 expression in hepatocytes or non-small cell lung cancer (NSCLC) leads to EMT phenotypes, which in turn cause loss of contact inhibition [74], enhance migration and invasion for metastasis [77], and render gefitinib resistance [78]. TM4SF5 expression causes morphological changes through abnormal regulation of RhoA and Rac1 in hepatocytes, together with loss of E-cadherin expression leading to an EMT induction [74] via an induction of Slug [79]. Inhibition of TM4SF5-mediated signaling event of a cytosolic enrichment of p27<sup>Kip1</sup> abolishes abnormal multilayer cell growth [74] and retards the G1 to S phase progression [80]. Further, inhibition of TM4SF5-mediated EMT by suppression of cytosolic p27<sup>Kip1</sup> expression leads for gefitinib-resistant NSCLC cells to become gefitinib-sensitive [78]. TM4SF5 is involved in activation of hepatic stellate cells via causing an EMT processes, leading to a correlation to development of liver fibrosis in CCl<sub>4</sub>-treated mouse models [81]. TM4SF5 expression is achieved by TGF $\beta$ 1-mediated Smads actions on the EGFR activation [73], such that the important roles of the multifunctional cytokine TGF $\beta$ 1 in activation of hepatic stellate cells and EMT are confirmed in a development of murine liver fibrosis. Since liver fibrosis can lead to eventually hepatocarcinoma at a high rate over 70%

[82], the roles of TM4SF5 both in development of fibrosis and tumorigenesis in livers can be reasonable.

Meanwhile, TM4SF5 expression enhances directional migration and invasion of hepatocytes. TM4SF5 in hepatocytes causes a directional migration at an enhanced speed and formation of more invadosome-like structures enriched with cortactin, actin, and actin-regulatory proteins like Arp2 and WASP [77]. TM4SF5-mediated directional migration involves a direct interaction and activation of FAK via the ICL domain of TM4SF5 and the F1 lobe of FAK FERM domain [75]. Further, TM4SF5-mediated invasive ECM degradation requires a direct interaction between the COOH-terminus of TM4SF5 and c-Src, which is linked to Tyr845 phosphorylation of EGFR to form more invasive protrusions [76]. TM4SF5-mediated multilayer growth [74], FAK activity, migration and invasion [75] are abolished by anti-TM4SF5 reagent, TSAHC (a synthetic compound), which appears to affect its *N*-glycosylation and at the same time blocks the TM4SF5-dependent EMT phenotype induction and multilayer growth [83]. Therefore, TM4SF5 also plays important roles in tumor initiation and progression, possibly being supported by an EMT process.

#### **4.5 TM4SF5-mediated other EMT-related biological processes**

EMT is well known to be related to also development [84] and stemness of self-renewal capacity [9]. We also observed that TM4SF5 can play roles in other EMT-mediated biological processes, like development of muscles and self-renewal capacity of cancer cells. In zebrafish, suppression of *tm4sf5* results in abnormal development of fishes with an aberrant trunk and morphology of muscle fibers, presumably via an alteration in expression and localization of integrin  $\alpha 5$  necessary for somite boundary maintenance (YJ Choi and JW Lee, unpublished observations). In addition, TM4SF5 expression in hepatocytes leads to spheroid formation in

a non-adhesive condition, which also causes xenograft tumor growth even with injections of cells at small numbers less than 5000 cells/mouse. The self-renewal capacity of the TM4SF5-positive cancer cells is abolished by treatment of anti-TM4SF5 small compound, TSAHC [83] (D Lee and JW Lee, unpublished observations). In addition to liver fibrosis and tumorigenesis, therefore, TM4SF5 expression is importantly involved in development of zebrafish muscles and acquirement of self-renewal property, which are known to be mediated by EMT.

Presumably, these diverse cellular effects by TM4SF5 expression might be possible due to the characteristic of TM4SF5, similar to the tetraspanins, which forms large protein networks via heterophilic or homophilic interactions between tetraspanins, integrins, and growth factor receptors. TM4SF5 is shown to bind integrin  $\alpha 2$ ,  $\beta 1$  [70, 71],  $\alpha 5$  [72], EGFR [73], CD151 (M Kang and JW Lee, unpublished observations), IL6R (J Ryu and JW Lee, unpublished observations), and so on. Although its ligand has not been identified, interaction(s) to (an)other membrane protein or receptor can recapitulate the liganding-based activation. Therefore, TM4SF5 can transduce signaling activities for diverse cellular functions including EMT and EMT-mediated different phenotypes. Although diverse miRs are known to regulate EMT [7], miRs targeting TM4SF5 are being studied.

We have observations showing that TM4SF5 expression can be related to stemness (Lee D and Lee JW, unpublished observations). Since TM4SF5 is important for EMT [74] and drug resistance [78], and EMT is also linked to stemness [10], it is likely that TM4SF5 can be linked to stemness property.

#### **4.6 TM4SF5-mediated gene regulation**

Comparison in protein expression patterns between TM4SF5-null and -expressing cells

shows a negative correlation between TM4SF5 and cell-cell adhesion-related molecules of epithelial markers including E-cadherin [74], and a positive correlation between TM4SF5 and mesenchymal markers including Slug [79] or Twist (D Lee and JW Lee, unpublished observations), supporting for TM4SF5-mediated EMT. RT-PCR analyses of them have been the cases, so that their expression regulation by TM4SF5 occurs at transcriptional levels (Lee JW, unpublished observation). However, the signaling pathways underlying for the expression regulation are not determined yet.

In addition, TM4SF5 expression correlates also with cytosolic p27<sup>Kip1</sup> [74]. Although p27<sup>Kip1</sup> in nucleus is inhibitory to cyclin-dependent kinases (CDKs) to suppress cell cycle and proliferation, its localization in the cytosol can lead to tumorigenic functions [85]. Cytosolic p27<sup>Kip1</sup> has been reported in different clinical reports, where different cancer types show enriched cytosolic localizations of p27<sup>Kip1</sup> [86-88], suggesting that cytosolic p27<sup>Kip1</sup> can be tumorigenic [89]. p27<sup>Kip1</sup> can be phosphorylated by Akt, KIS, or JNK [90-92], resulting in translocation and stabilization in the cytosol, where it binds to and inactivates RhoA GTPase leading to alteration in actin organization and motility regulation [93]. TM4SF5 expression also causes overexpression of p27<sup>Kip1</sup>, although how it occurs is unknown yet; TM4SF5 causes Akt-mediated Ser10 phosphorylation of p27<sup>Kip1</sup>, leading to its stabilization and RhoA activity changes, and eventually morphological elongation for EMT and contact inhibition loss [74]. JNK-mediated p27<sup>Kip1</sup> phosphorylation in a TM4SF5-dependent manner also results in localization of p27<sup>Kip1</sup> at cell-cell contacts [91], possibly leading to altered actin organization at the cell-cell contacts. In addition, proteasome inhibition in terms of proteasome activity and proteasome subunit expression also depending on TM4SF5 expression results in morphological changes and EMT, suggesting another novel mechanism

for TM4SF5-mediated EMT [79].

Meanwhile, TM4SF5 causes activation of FAK/c-Src signaling pathways leading to STAT3 phosphorylation at Tyr705 for induction and secretion of VEGF, which can stimulate neighboring endothelial cells for enhanced (tumor) angiogenesis [72]. During modeling of tumor microenvironment, cancer cells overexpressing TM4SF5 appear to negatively regulate expression of cytokine IL6, and exogenous IL6 treatment leads to a less STAT3 signaling activation in TM4SF5-positive cancer cells (J. Ryu and JW Lee, unpublished observations), so that a TM4SF5-dependent suppression of IL6 can be a strategy for the TM4SF5-positive tumor cells to avoid pro-immunological actions by IL6 secreted by neighboring immune cells. In addition, we also observe that TM4SF5 expression induces mRNA and protein of CD151, another tumorigenic tetraspanin, but suppresses those of CD63, a tumor-suppressive tetraspanin, which eventually enhance aggressive migration and invasion (M. Kang and JW Lee, unpublished observations). As for invasion, TM4SF5 expression also increases mRNA and protein levels of MMP2, in addition to its activity [77].

Therefore, TM4SF5 expression correlates with or plays important roles in tumorigenesis in different mechanisms including induction of EMT and gene regulation as well (Fig. 2).

## **5. Concluding remarks**

Considering that such membrane proteins of TM4SF5 may be an important upstream regulator of EMT and invasiveness of cancer cells and their expressions substantially differ in normal and cancer tissues, targeting them could be novel therapeutic targets for the treatment of cancer metastasis. In the future, functional involvement of

TMPRSS4 and/or TM4SF5 in the initiation and progression of tumor needs to be evaluated using transgenic mouse models. Cancer-associated mutations and single nucleotide polymorphisms (SNPs) within the *TMPRSS4* or *TM4SF5* gene also needs to be analyzed in association with cancer risk.

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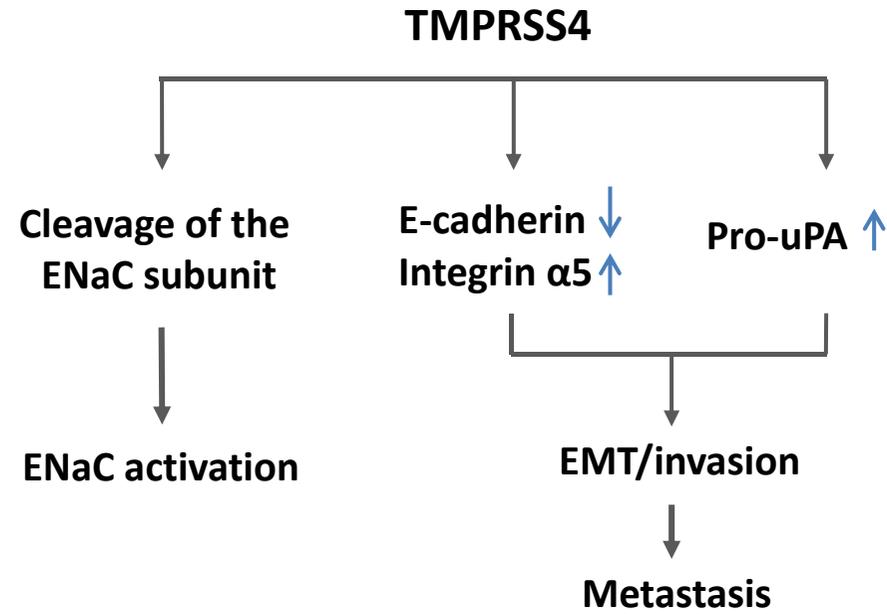
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**Fig. 1**



**Figure 1. Cellular functions of TMPRSS4**

**Fig. 2**

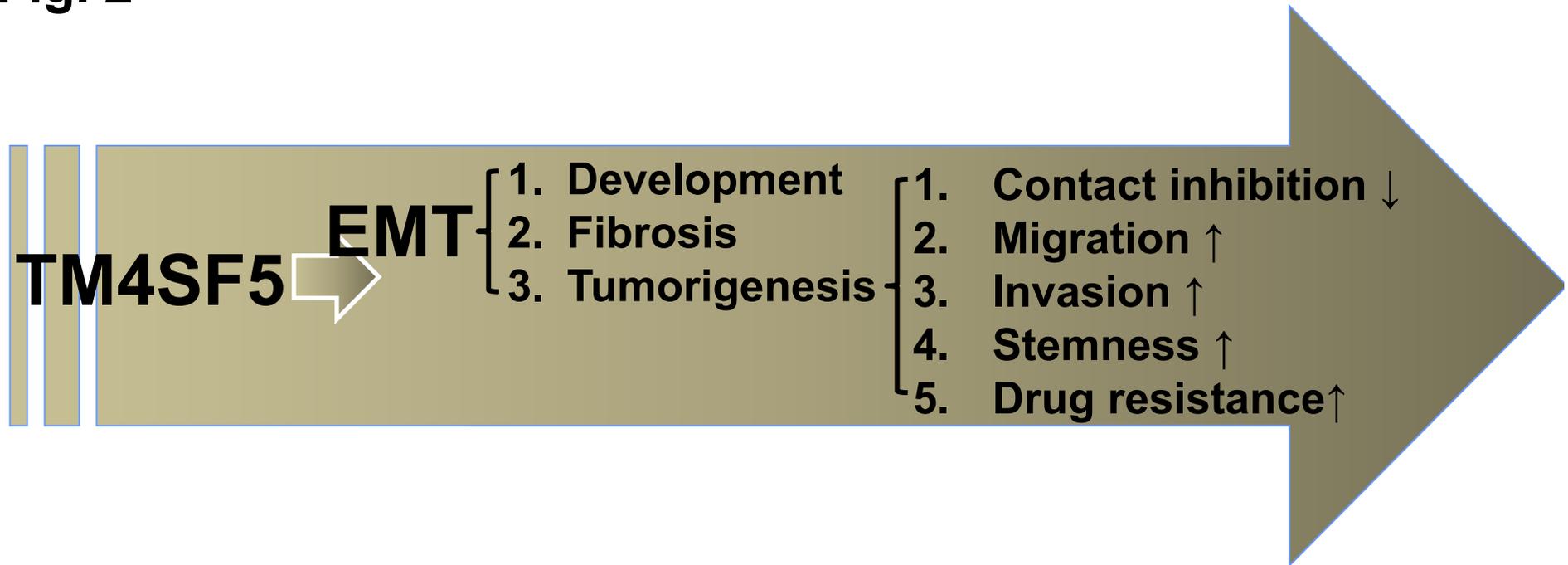


Fig. 2. TM4SF5-mediated EMT is involved in diverse cellular functions, leading to liver tumorigenesis and maintenance in addition to developmental processes.