

In vivo comparison between the effects of chemically modified hydrophilic and anodically oxidized titanium surfaces on initial bone healing

Hyo-Jung Lee^{1,†}, Il-Hyung Yang^{2,†}, Seong-Kyun Kim³, In-Sung Yeo³, Taek-Ka Kwon^{4,*}

¹Department of Periodontology, Section of Dentistry, Seoul National University Bundang Hospital, Seongnam, Korea

²Department of Orthodontics and Dental Research Institute, Seoul National University School of Dentistry, Seoul, Korea

³Department of Prosthodontics and Dental Research Institute, Seoul National University School of Dentistry, Seoul, Korea

⁴Department of Prosthetic Dentistry, St. Vincent Hospital, Catholic University of Korea, Suwon, Korea

Purpose: The aim of this study was to investigate the combined effects of physical and chemical surface factors on *in vivo* bone responses by comparing chemically modified hydrophilic sandblasted, large-grit, acid-etched (modSLA) and anodically oxidized hydrophobic implant surfaces.

Methods: Five modSLA implants and five anodized implants were inserted into the tibiae of five New Zealand white rabbits (one implant for each tibia). The characteristics of each surface were determined using field emission scanning electron microscopy, energy dispersive spectroscopy, and confocal laser scanning microscopy before the installation. The experimental animals were sacrificed after 1 week of healing and histologic slides were prepared from the implant-tibial bone blocks removed from the animals. Histomorphometric analyses were performed on the light microscopic images, and bone-to-implant contact (BIC) and bone area (BA) ratios were measured. Nonparametric comparison tests were applied to find any significant differences ($P < 0.05$) between the modSLA and anodized surfaces.

Results: The roughness of the anodized surface was $1.22 \pm 0.17 \mu\text{m}$ in Sa, which was within the optimal range of 1.0–2.0 μm for a bone response. The modSLA surface was significantly rougher at $2.53 \pm 0.07 \mu\text{m}$ in Sa. However, the modSLA implant had significantly higher BIC than the anodized implant ($P = 0.02$). Furthermore, BA ratios did not significantly differ between the two implants, although the anodized implant had a higher mean value of BA ($P > 0.05$).

Conclusions: Within the limitations of this study, the hydrophilicity of the modSLA surface may have a stronger effect on *in vivo* bone healing than optimal surface roughness and surface chemistry of the anodized surface.

Keywords: Animal experimentation; Dental implants; Histology; Osseointegration.

INTRODUCTION

Successful osseointegration of titanium (Ti) implants is partly determined by how the implanted materials influence bone responses at the cell-biomaterial interface [1,2]. Such events occurring between the bone and implant surface are influenced by a variety of specific surface properties, including topography, structure, chemistry, surface charge, and wettability [3–6]. Of these, surface topography has been particularly well studied. Researchers have developed numerous additive and subtractive surface modification techniques to improve osseointegration by altering implant surface topography, thus enhancing bone-to-implant contact (BIC) and increasing biomechanical interlocking with bone [7,8]. The clinical introduction of a novel implant surface has also helped to advance the field [9]. This product

pISSN 2093-2278
eISSN 2093-2286



JPIS >
Journal of Periodontal
& Implant Science

Research Article

J Periodontol Implant Sci 2015;45:94–100
<http://dx.doi.org/10.5051/jpis.2015.45.3.94>

Received: Apr. 5, 2015

Accepted: Apr. 29, 2015

*Correspondence:

Taek-Ka Kwon

Department of Prosthetic Dentistry, St. Vincent Hospital, the Catholic University of Korea, 93 Jungbu-daero, Paldal-gu, Suwon 422-723, Korea
E-mail: tega95@naver.com

Tel: +82-31-249-7670

Fax: +82-31-258-3352

[†]Hyo-Jung Lee and Il-Hyung Yang contributed equally to this study.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>).

aims to influence surface charge and wettability in animals via ultraviolet light irradiation [9,10].

Researchers have developed surfaces that are supposedly based not only on micrometre morphology, but also on other characteristics such as hydrophilicity, chemical bonding, and nanostructures [11]. Reports on the wetting behavior of rough surfaces have increased our understanding of the conditions surface topography has to satisfy to induce satisfactory hydrophilicity during contact with bone [12–15]. Extensive hydroxylation/hydration of the oxide layer, together with high wettability, improves interactions between the surface and the water shells around delicate biomolecules such as proteins [16]. Studies using modified sandblasted, large-grit, acid-etched (modSLA) surfaces that enhance hydrophilicity have indicated that bone apposition during the early stages of regeneration is higher after implantation compared with its predecessor (the SLA surface) [17–19].

Anodic oxidation creates a thickened, porous, and moderately roughened titanium oxide layer [20,21]. The anodized titanium surface shows superior osteogenic properties both *in vitro* and *in vivo* despite being hydrophobic [20–24]. Although many studies have examined the longer-term impacts of surface roughness and topography on bone fixation over the long term, there has been relatively little work investigating the effects of these hydrophilic characteristics on the initial bone response [17,19,25,26]. To the best of our knowledge, there were not many studies that have evaluated the effects of hydrophobic oxidized and hydrophilic modSLA surfaces on early bone response *in vivo* [27]. Although the positive effects of the modSLA implants could be easily explained by their hydrophilicity, the clinical relevance needs to be further investigated [12].

We performed histomorphometric analyses to investigate the combined effects of physical and chemical surface factors on *in vivo* bone responses by comparing a modSLA surface and an anodized implant surface in a rabbit tibia model.

MATERIALS AND METHODS

Surface characteristics

Five modSLA (SLActive®, Institut Straumann AG, Basel, Switzerland) and five anodized (TiUnite®, Nobel Biocare AB, Göthenburg, Sweden) implants were used in this study. Both implants were 3.3 mm in diameter and 10.0 mm in length. We performed three surface analyses on each of three implants from both groups: field emission scanning electron microscopy (FE-SEM), energy dispersive spectroscopy (EDS), and confocal laser scanning microscopy (CLSM). The FE-SEM (model S-4700, Hitachi, Tokyo, Japan) was used to produce detailed images of the implant surfaces. The EDS (model EX220, Horiba Ltd., Kyoto, Japan) was used to analyze the element content and components of the modified surfaces; calibrations were performed four times each at four different points. The CLSM (model LSM 5-Pascal, Carl Zeiss AG, Oberkochen, Germany) enabled us to measure the surface roughness of four screw sides (measurement area: 300 µm × 300 µm on a 200× optically and 1.5× digi-

tally magnified image), which were randomly selected from each implant. We measured two roughness parameters: average surface deviation (Sa) and developed surface area ratio (Sdr) [21].

In vivo surgery

This study was approved by the Animal Research Committee of Seoul National University Bundang Hospital (IACUC protocol approval number: BA1101-076/001-01). All procedures, including animal selection, management, preparation, and subsequent surgical protocols, were performed in accordance with the Institute of Laboratory Animal Resources guidelines of Seoul National University Bundang Hospital.

Five male New Zealand white rabbits (each about 6 months of age and weighing 2.5–3 kg) were implanted with a modSLA and an anodized implant; the location of each implant (left or right tibia) was chosen at random. The rabbits showed no sign of illness or disease prior to the study. Prior to surgery, all study subjects were anesthetized with an intramuscular injection of tiletamine/zolazepam (15 mg/kg; Zoletil 50, Virbac Korea Co. Ltd., Seoul, Korea) and xylazine (33 mg/kg; Rompun, Bayer Korea Ltd., Seoul, Korea). The skin of each proximal tibia area was shaved and washed with povidone iodine solution, and each rabbit received an intramuscular injection with 33 mg/kg of Cefazolin (Yuhan Co., Seoul, Korea), a preoperative prophylactic antibiotic. The local anesthetic lidocaine (1:100,000 epinephrine; Yuhan Co.) was injected into each surgical site. The skin was incised with a surgical blade, and each tibia was exposed via full-thickness periosteal flap reflection. The implant sites were prepared on the flat tibial surface using a dental implant drill and profuse sterile saline irrigation.

We performed bicortical drilling as described in a previous study [20]. For the 3.3-mm implants, we used a drill that was 2.8 mm in diameter; gap defects were created with a cortical drill (Astra Tech, Mölndal, Sweden) with a 4.0 mm diameter (Fig. 1). The cortical drill was used monocortically and created a 4.0-mm hole in the upper cortex only. After implant insertion, cover screws were securely fastened and the surgical sites were closed in layers. Muscle and fascia were sutured with absorbable Vicryl sutures (Vicryl 4-0, Polyglactin 910, Ethicon, Johnson & Johnson, Somerville, NJ, USA) and the outer dermis was closed with a silk suture (Mersilk 4-0, Ethicon, Johnson & Johnson). Rabbits were housed in separate cages for 1 week post-surgery, after which they were anesthetized and sacrificed by intravenous administration of potassium chloride.

Histomorphometric analysis

The tibiae of the sacrificed rabbits were exposed so that implants could be surgically removed en bloc with an adjacent collar of bone, which was immediately fixed in 10% neutral formaldehyde. For histomorphometry, the specimens were embedded in light-curing resin (Technovit 7200 VLC, Kultzer, Wehrheim, Germany) prepared as previously described [28]. Undecalcified, cut, and ground sections were prepared using the Exakt® system (Exakt Apparatebau, Norderstedt, Germany) according to the method described by Donath and

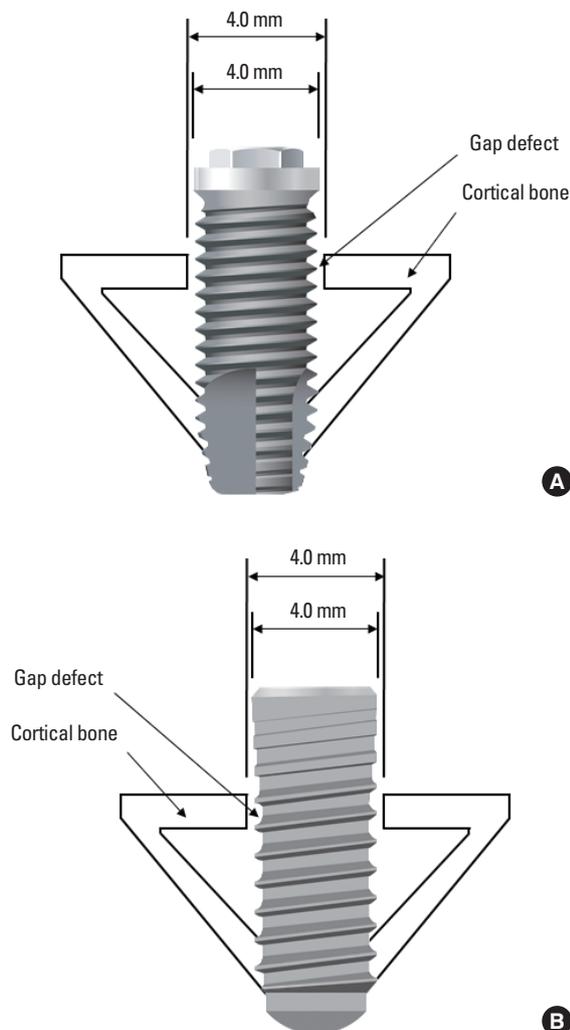


Figure 1. A schematic diagram demonstrating how to insert the implant into the rabbit tibia. (A) An anodized implant and (B) a modSLA implant, both of which were 3.3 mm in diameter, were firmly engaged at the bottom of the cortex in the rabbit tibia. A hole, 4.0 mm in diameter, was equally formed at the upper cortex only, although the thread morphologies were different between the two groups. Note that the threads of the implants were not engaged at the upper cortical area where a gap remained.

Breuner [29]. The specimens were ground to a thickness of approximately 50 μm and stained with hematoxylin and eosin (H&E). Histological examinations of specimens were performed under a light microscope (Olympus BX, Olympus, Tokyo, Japan). BIC and BA percentages were defined and measured in the range of 2 mm below the upper bone crest, as shown in Fig. 2. Histomorphometric analyses were performed on both the right and left sides of each specimen using image analysis software (Kappa PS30C Imagebase, Kappa Opto-electronics GmbH, Gleichen, Germany).

Statistical analyses

The Mann-Whitney U test was used to assess the statistical significance of the difference in surface roughness parameters (Sa and

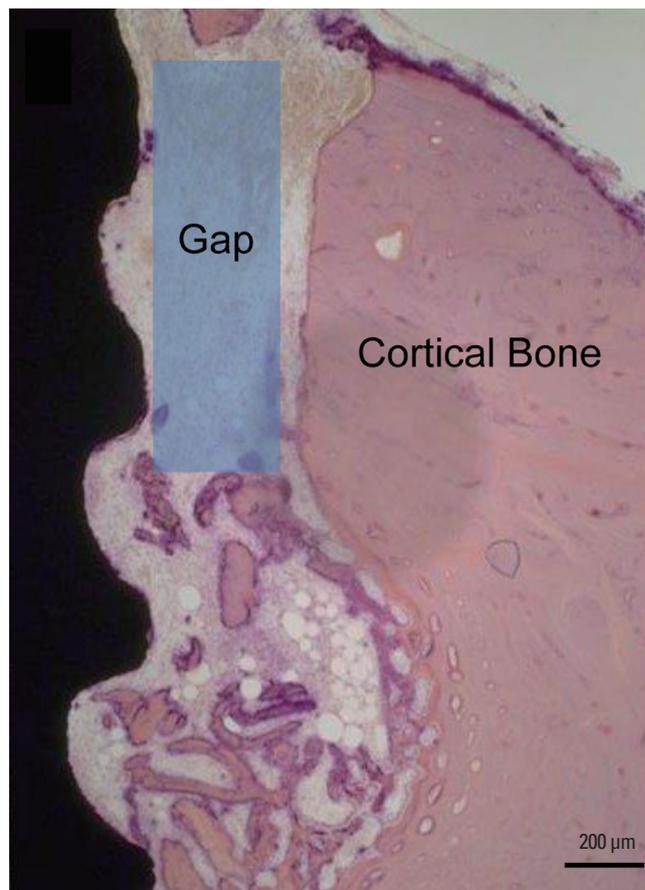


Figure 2. A light microscopic image for histomorphometric analyses (H&E staining, 75 \times magnification). Note the gap space (blue area) that was intentionally made between the implant surface and the cortical bone.

Sdr) between the test and control implants. The Wilcoxon signed-rank test was used to determine statistically significant differences in BIC and BA between the groups. Significance was defined as $P < 0.05$.

RESULTS

Analysis of surface characteristics

The modSLA surface comprised 70.3% titanium, 29.3% oxide, and no phosphorous, while the anodized surface was 26.1% titanium, 69.8% oxide, and 3.5% phosphorus (Table 1). The FE-SEM images of each surface are shown in Fig. 3. Magnification at 5,000 \times revealed the anodized surface to be scattered with many volcano-like porous structures. The modSLA surface had a sharp, irregular pattern produced by the sandblasting and acid-etching processes. At 50,000 \times magnification, the anodized implant was characterized by a relatively smooth surface composed of large micro-pores and small nanopores. This contrasted with the relatively rough surface observed on the hydrophilic modSLA implant, which exhibited beadings of approximately 1-2 μm diameter, along with 0.1-0.2 μm

millet-like prominences.

Surface roughness data collected during the CLSM analysis are shown in Table 2. Anodically oxidized surfaces had significantly lower values for both Sa ($1.22 \pm 0.17 \mu\text{m}$) and Sdr ($26.27 \pm 4.14\%$) than observed for chemically modified surfaces ($2.53 \pm 0.07 \mu\text{m}$ and $139.82 \pm 7.59\%$, respectively) (both $P < 0.001$). Although the modSLA implant screw design is flatter and thus has less total surface area, the modSLA screws are smaller and result in greater osseointegration because of the higher percentage of BIC.

Histomorphometric analysis

All experimental animals healed without complications and at the time of sacrifice all implants were submerged and covered by a healthy ridge of skin. For both types of implant, a favorable bone

response was observed on the implant surface; a small amount of new bone formation was found both within a thread and in the old bone after only 1 week of healing. Osseointegration within the gap defect was notable in some specimens; however, the distinguishing feature of most samples was a visible growth pattern along the border of the gap defect toward the implant surface (Fig. 4). This was observed in both treatments with a statistically significant greater growth pattern seen in the modSLA group. Although the 1-week post-surgical period was insufficient for new bone formation, all specimens showed at least the beginnings of new bone formation or active bone formation on the inner cutting side of the cortical bone and the inner portion of bone marrow. Both implant surfaces were surrounded by small, newly formed trabeculae of woven bone. Although histomorphometric light microscopy revealed that the BIC ratio was significantly higher around the mod-

Table 1. Element content (atomic %) of the implant surfaces according to energy dispersive spectroscopy analysis.

	ModSLA (n=5)	Anodized (n=5)
Ti	70.3±6.4%	26.1±0.9%
O	29.3±6.5%	69.8±1.2%
P	0%	3.5±0.4%
Pt	0.4±0.2%	0.6±0.02%
Ca	0.003±0.005%	0.06±0.07%

Data were shown as mean±standard deviation.

Table 2. Surface roughness of the implant surfaces according to confocal laser scanning electron microscopy analysis.

	ModSLA (n=5)	Anodized (n=5)	P-value
Sa	2.53±0.07 μm	1.22±0.17 μm	<0.001 ^{a)}
Sdr	139.82±7.59%	26.27±4.14%	<0.001 ^{a)}

Data were shown as mean±standard deviation. Sa, average surface deviation; Sdr, developed surface area ratio.

5 point bottom calibration. Focal area 300 $\mu\text{m} \times 300 \mu\text{m}$.

^{a)}Data were analyzed using the Mann-Whitney U test.

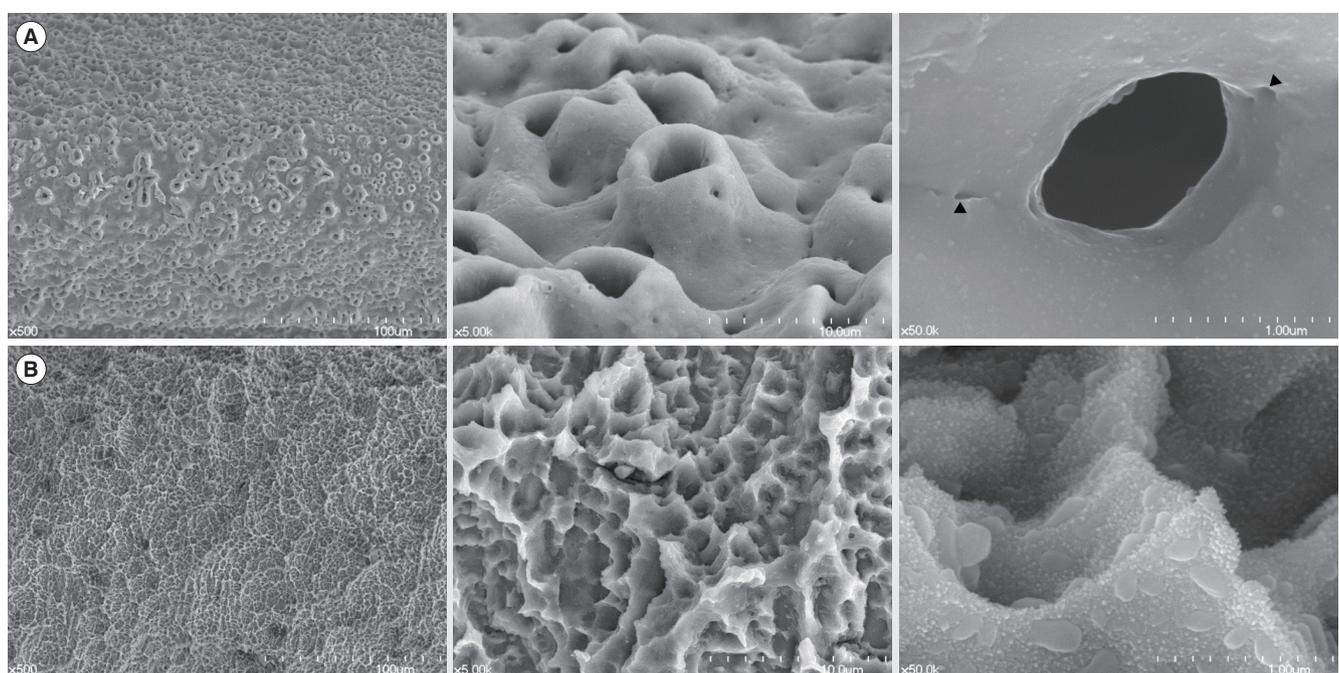


Figure 3. The FE-SEM images of each implant surface at different resolutions (500 \times , 5,000 \times and 50,000 \times from the top). (A) Seen under medium power the anodically oxidized surface has many micropores with elevated margins resembling volcanoes. In a high-power image, a relatively smooth surface composed of large micropores (1–10 μm) and many nanopores with orifices (<1 μm , black arrowheads) are visible. (B) Under medium-power microscopy the chemically modified modSLA surface has a sharp, irregular pattern produced by sandblasting and acid-etching. In a high-power image, 1–2 μm diameter beadings and 0.1–0.2 μm millet-like prominences were observed on the surface.

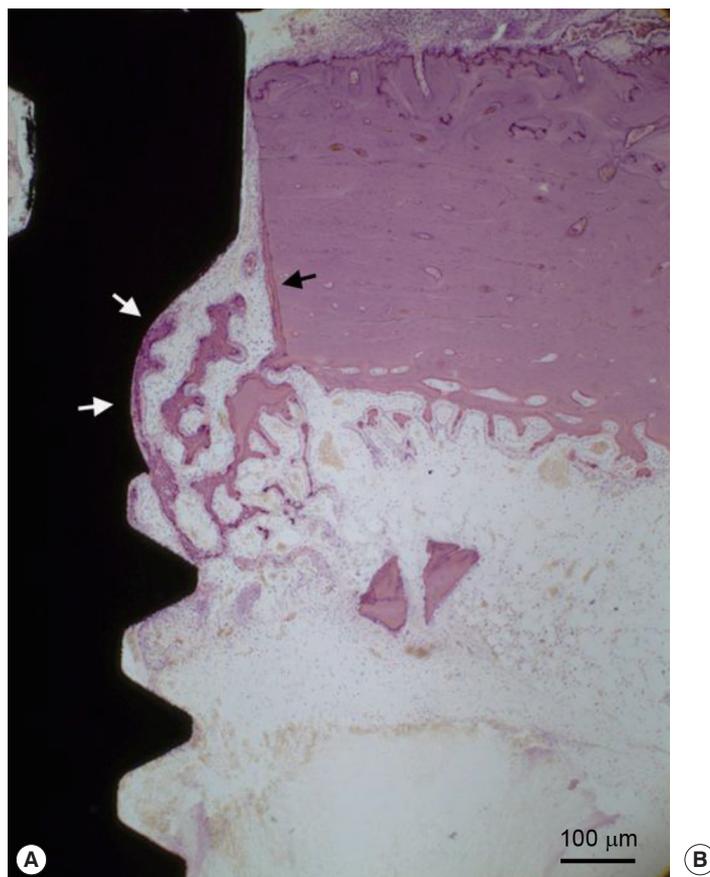


Figure 4. Histologic views of wound healing at day 7 with light microscopic findings (H&E stain, 100× magnification). Newly formed bone was seen from the cutting area of the cortical bone toward the implant surface (black arrows). New bone was also formed from the implant surface (white arrows). Bone formation was found to occur in the marrow area, mainly behind the existing cortical bone. Histologic views were observed around both the anodized (A) and modSLA (B) implants.

Table 3. The bone-implant-contact and bone area 1 week after the surgery.

	ModSLA (n=5)	Anodized (n=5)	P-value
BIC	17.9±19.4%	2.9±8.3%	0.02 ^{al}
BA	14.0±12.0%	4.0±9.0%	0.09 ^{al}

Data shown as mean±standard deviation. BIC, bone-to-implant contact; BA, bone area ratio.

^{al}Data were analyzed using the Mann-Whitney U test.

SLA implants than around the anodized implants ($P=0.02$; Table 3), there was no significant difference in BA between the two types of implant because of a large standard deviation ($P=0.09$; Table 3).

DISCUSSION

We set out to determine whether modSLA implants would possess clinical superiority over anodically oxidized surface implants. Given the results of our *in vivo* experiment, hydrophilicity seems to be very important in bone response.

Thread density but not thread geometry is known to have an ef-

fect on BIC [30,31]. As the anodized implant had a shorter thread pitch than the modSLA implant in this study (Fig. 4), it was considered to have an advantage in BIC. We also found the modSLA surface to be significantly rougher than the anodized surface. Although the modSLA surface is classified as rough, the anodized surface is considered moderately rough, which is known to be more advantageous for bone responses than a "rough" surface [32]. However, we found a significantly higher BIC in response to the modSLA implant. In fact, in terms of the surface treatment based on Sa and Sdr, several studies concluded that modSLA surface showed better performance than conventional SLA surface as well as different surface treatment [17,19,21]. Although the mechanism linking surface properties and osteoblast production is not yet sufficiently understood, the hydrophilic property of the modSLA surface may have a stronger influence on bone response than either the surface configuration of the implant or the surface features resulting from anodic oxidation [19]. These results correlated with the previous study concluding that surface hydrophilicity rather than microtopography affected soft and hard tissue integration [33].

Since the dental implant was first introduced, there have been a

huge number of studies of which the purposes were to find the factors for the improvement of osseointegration. Eventually, we now have diverse dental implants at our hand with various designs, configurations, surface treatment, and modifications. Huge leap of development was already made in the field of dental implants. Therefore, the scientific investigation on only one factor for better dental implant by well-controlled experiments can be so stereotyped but the intuitive comparison among the several dental implants considering each one as an independent entity of the experiment, although somewhat obscure and less scientific, can be more practical and clinically helpful. The well-controlled study on one factor could be even more redundant [6,21]. In such a way, a previous study showed that hydrophilic SLA group showed higher BIC after 10 days than Nobel Biocare Replace Select implant group with oxidized TiUnite surface [34].

We observed many different stages of new bone formation, ranging from no osseointegration at all to a very thin layer of new bone around the original cortical bone, to marked growth toward the implant surface, to complete osseointegration with new bone, which is why such a large standard deviation was observed on histomorphometric analyses. One week of post-surgery recovery time is too short to truly evaluate osseointegration at the bone-implant interface, although the findings of this study support previous results indicating that this process is initiated within the first week of wound healing [35,36]. Further studies are needed to more clearly determine significant histomorphometric differences by controlling the healing period after implant insertion.

Our results suggest that the hydrophilic modSLA surface may have a stronger affinity for bone than the anodized surface during the initial healing period. Somewhat similar results have been indicated by previous studies reporting that a hydrophilic surface implant is associated with better initial bone response [17,37]. Although both surface configuration and hydrophobic properties of the implant surface were found to affect early bone formation, the latter appears to have a more significant effect on BIC for reasons that are yet to be fully elucidated. Various experiments including animal models and immunohistochemistry have reported that blood clots may be formed within 24 hours of implant insertion and that formation of capillaries preceded and accelerated new bone formation [37,38]. We found blood clots close to the hydrophilic modSLA surface, while a previous study demonstrated that the coagulum was partially collapsed at the conventional SLA surface [37]. Within the limitations of this study, the hydrophilicity of the modSLA surface may have a stronger effect on *in vivo* bone healing than optimal surface roughness and surface chemistry of the anodized surface. Further investigations are required to elucidate the interactions between the implant surface factors such as hydrophilicity and the physiology of blood and bone with large sample size at the various different stages during the early healing period.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

ORCID

Hyo-Jung Lee <http://orcid.org/0000-0002-0439-7389>
 Il-Hyung Yang <http://orcid.org/0000-0001-6398-4607>
 Seong-Kyun Kim <http://orcid.org/0000-0001-8694-8385>
 In-Sung Yeo <http://orcid.org/0000-0002-6780-2601>
 Taek-Ka Kwon <http://orcid.org/0000-0003-2961-6135>

ACKNOWLEDGEMENTS

This work was supported by the Seoul National University Bundang Hospital Research Fund (Grant no. 11-2010-029) and the International Research & Development Program of the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT & Future Planning (Grant number: 2014K1A3A1A21001365). The authors are indebted to Professor Emeritus J. Patrick Barron (Tokyo Medical University, Tokyo, Japan and Adjunct Professor, Seoul National University Bundang Hospital, Bundang, Republic of Korea) for his pro bono editing of this manuscript.

REFERENCES

1. Anselme K, Bigerelle M, Noel B, Dufresne E, Judas D, Iost A, et al. Qualitative and quantitative study of human osteoblast adhesion on materials with various surface roughnesses. *J Biomed Mater Res* 2000;49:155-66.
2. Lausmaa J. Surface spectroscopic characterization of titanium implant materials. *J Electron Spectrosc Relat Phenomena* 1996;81: 343-61.
3. Albrektsson T. Direct bone anchorage of dental implants. *J Prosthet Dent* 1983;50:255-61.
4. Lim YJ, Oshida Y, Andres CJ, Barco MT. Surface characterizations of variously treated titanium materials. *Int J Oral Maxillofac Implants* 2001;16:333-42.
5. Taborelli M, Jobin M, François P, Vaudaux P, Tonetti M, Szmukler-Moncler S, et al. Influence of surface treatments developed for oral implants on the physical and biological properties of titanium. (I) Surface characterization. *Clin Oral Implants Res* 1997;8: 208-16.
6. Wennerberg A, Albrektsson T. Effects of titanium surface topography on bone integration: a systematic review. *Clin Oral Implants Res* 2009;20 Suppl 4:172-84.
7. Cooper LF. A role for surface topography in creating and maintaining bone at titanium endosseous implants. *J Prosthet Dent* 2000; 84:522-34.
8. Pilliar RM. Overview of surface variability of metallic endosseous dental implants: textured and porous surface-structured designs.

- Implant Dent 1998;7:305-14.
9. Kilpadi DV, Lemons JE. Surface energy characterization of unalloyed titanium implants. *J Biomed Mater Res* 1994;28:1419-25.
 10. Wang R, Hashimoto K, Fujishima A, Chikuni M, Kojima E, Kitamura A, et al. Light-induced amphiphilic surfaces. *Nature* 1997;388:431-2.
 11. Wennerberg A, Albrektsson T. Suggested guidelines for the topographic evaluation of implant surfaces. *Int J Oral Maxillofac Implants* 2000;15:331-44.
 12. Bico J, Thiele U, Quéré D. Wetting of textured surfaces. *Colloids Surf A Physicochem Eng Asp* 2002;206:41-6.
 13. Rupp F, Scheideler L, Eichler M, Geis-Gerstorfer J. Wetting behavior of dental implants. *Int J Oral Maxillofac Implants* 2011;26:1256-66.
 14. Rupp F, Scheideler L, Olshanska N, de Wild M, Wieland M, Geis-Gerstorfer J. Enhancing surface free energy and hydrophilicity through chemical modification of microstructured titanium implant surfaces. *J Biomed Mater Res A* 2006;76:323-34.
 15. Rupp F, Scheideler L, Rehbein D, Axmann D, Geis-Gerstorfer J. Roughness induced dynamic changes of wettability of acid etched titanium implant modifications. *Biomaterials* 2004;25:1429-38.
 16. Textor M, Sittig C, Frauchiger V, Tosatti S, Brunette DM. Properties and biological significance of natural oxide films on titanium and its alloys. In: Brunette DM, Tengvall P, Textor M, Thomsen P, editors. *Titanium in medicine: material science, surface science, engineering, biological responses, and medical applications*. Berlin: Springer; 2001. p.171-230.
 17. Buser D, Broggini N, Wieland M, Schenk RK, Denzer AJ, Cochran DL, et al. Enhanced bone apposition to a chemically modified SLA titanium surface. *J Dent Res* 2004;83:529-33.
 18. Junker R, Dimakis A, Thoneick M, Jansen JA. Effects of implant surface coatings and composition on bone integration: a systematic review. *Clin Oral Implants Res* 2009;20 Suppl 4:185-206.
 19. Schwarz F, Ferrari D, Herten M, Mihatovic I, Wieland M, Sager M, et al. Effects of surface hydrophilicity and microtopography on early stages of soft and hard tissue integration at non-submerged titanium implants: an immunohistochemical study in dogs. *J Periodontol* 2007;78:2171-84.
 20. Choi JY, Lee HJ, Jang JU, Yeo IS. Comparison between bioactive fluoride modified and bioinert anodically oxidized implant surfaces in early bone response using rabbit tibia model. *Implant Dent* 2012;21:124-8.
 21. Wennerberg A, Albrektsson T. On implant surfaces: a review of current knowledge and opinions. *Int J Oral Maxillofac Implants* 2010;25:63-74.
 22. Iwai-Yoshida M, Shibata Y, Wurihan, Suzuki D, Fujisawa N, Tanimoto Y, et al. Antioxidant and osteogenic properties of anodically oxidized titanium. *J Mech Behav Biomed Mater* 2012;13:230-6.
 23. Le Guehennec L, Lopez-Heredia MA, Enkel B, Weiss P, Amouriq Y, Layrolle P. Osteoblastic cell behaviour on different titanium implant surfaces. *Acta Biomater* 2008;4:535-43.
 24. Schuler RF, Janakievski J, Hacker BM, O'Neal RB, Roberts FA. Effect of implant surface and grafting on implants placed into simulated extraction sockets: a histologic study in dogs. *Int J Oral Maxillofac Implants* 2010;25:893-900.
 25. Ferguson SJ, Broggini N, Wieland M, de Wild M, Rupp F, Geis-Gerstorfer J, et al. Biomechanical evaluation of the interfacial strength of a chemically modified sandblasted and acid-etched titanium surface. *J Biomed Mater Res A* 2006;78:291-7.
 26. Wall I, Donos N, Carlqvist K, Jones F, Brett P. Modified titanium surfaces promote accelerated osteogenic differentiation of mesenchymal stromal cells in vitro. *Bone* 2009;45:17-26.
 27. Hong J, Kurt S, Thor A. A hydrophilic dental implant surface exhibits thrombogenic properties in vitro. *Clin Implant Dent Relat Res* 2013;15:105-12.
 28. Yeo IS, Han JS, Yang JH. Biomechanical and histomorphometric study of dental implants with different surface characteristics. *J Biomed Mater Res B Appl Biomater* 2008;87:303-11.
 29. Donath K, Breuner G. A method for the study of undecalcified bones and teeth with attached soft tissues. The Säge-Schliff (sawing and grinding) technique. *J Oral Pathol* 1982;11:318-26.
 30. Roberts WE, Smith RK, Zilberman Y, Mozsary PG, Smith RS. Osseous adaptation to continuous loading of rigid endosseous implants. *Am J Orthod* 1984;86:95-111.
 31. Steigenga J, Al-Shammari K, Misch C, Nociti FH Jr, Wang HL. Effects of implant thread geometry on percentage of osseointegration and resistance to reverse torque in the tibia of rabbits. *J Periodontol* 2004;75:1233-41.
 32. Albrektsson T, Wennerberg A. Oral implant surfaces: Part 2--review focusing on clinical knowledge of different surfaces. *Int J Prosthodont* 2004;17:544-64.
 33. Schwarz F, Herten M, Sager M, Wieland M, Dard M, Becker J. Bone regeneration in dehiscence-type defects at chemically modified (SLActive) and conventional SLA titanium implants: a pilot study in dogs. *J Clin Periodontol* 2007;34:78-86.
 34. Gottlow J, Barkarmo S, Sennerby L. An experimental comparison of two different clinically used implant designs and surfaces. *Clin Implant Dent Relat Res* 2012;14 Suppl 1:e204-12.
 35. Abrahamsson I, Berglundh T, Linder E, Lang NP, Lindhe J. Early bone formation adjacent to rough and turned endosseous implant surfaces. An experimental study in the dog. *Clin Oral Implants Res* 2004;15:381-92.
 36. Berglundh T, Abrahamsson I, Lang NP, Lindhe J. De novo alveolar bone formation adjacent to endosseous implants. *Clin Oral Implants Res* 2003;14:251-62.
 37. Schwarz F, Herten M, Sager M, Wieland M, Dard M, Becker J. Histological and immunohistochemical analysis of initial and early osseous integration at chemically modified and conventional SLA titanium implants: preliminary results of a pilot study in dogs. *Clin Oral Implants Res* 2007;18:481-8.
 38. Schmid J, Wallkamm B, Hämmerle CH, Gogolewski S, Lang NP. The significance of angiogenesis in guided bone regeneration. A case report of a rabbit experiment. *Clin Oral Implants Res* 1997;8:244-8.